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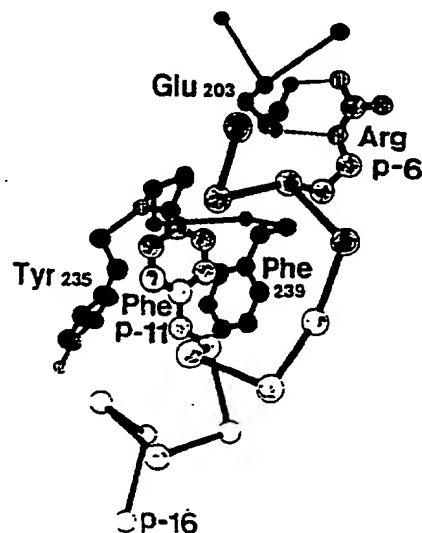
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(54) Title: METHODS OF DESIGNING SPECIFIC AFFECTORS USING THREE-DIMENSIONAL CONFORMATION OF ENZYME/AFFECTOR COMPLEX

(57) Abstract

The present invention includes methods for rational drug design. One exemplary method disclosed herein teaches the preparation of a highly specific affector of a first enzyme when the first enzyme is a member of a class of enzymes having a conserved catalytic core with a plurality of invariant residues. This method comprises identifying a second enzyme that is a member of that class of enzymes and has a known affector. The affector can be an inhibitor or activator of the second enzyme. In the practise of the method, a first complex is formed between the second enzyme and the known affector and data is obtained regarding the three-dimensional coordinates of the invariant residues in the complex. These coordinates are used to form a template. A model is then generated in which the first enzyme is in a conformation with the invariant residues in substantially the same conformation as in the template. Changes in the variable residues of the catalytic core of the first enzyme are compared to the variable residues in the catalytic core of the second enzyme. The second enzyme is modified to include these non-conserved changes and an affector is designed using computer modelling that will induce a conformation on the catalytic core of the first enzyme in which the invariant residues are in substantially the same coordinates as the coordinates of the template when the first enzyme is formed as a second complex with the newly designed affector. The designed affector can be further refined to provide improved affector activity.



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METHODS OF DESIGNING SPECIFIC AFFECTORS USING THREE DIMENSIONAL CONFORMATION OF ENZYME/AFFECTOR COMPLEX

Field of the Invention

5 The present invention relates to rational design of specific affectors for a given enzyme using data obtained regarding the three dimensional conformation of an enzyme/affecter complex. More particularly, it relates to such methods wherein the conformation of the conserved catalytic core of a given enzyme class is elucidated and highly specific affector molecules for a particular member of that class are designed.

Background of the Invention

10 Drug design based on an analysis of the structural features of a molecule is still in its infancy. At present, an analysis of X-ray crystallographic data at best permits the design of broadly acting affector molecules. While these affector molecules can be further refined to impart some selectivity, affector design does not produce molecules having the fine tuned specificity of, for example, an antibody for its antigen. This level of selectivity control is not
15 always necessary; however, therapeutic regimes directed to the control of enzymes involved in certain cancers, genetic disorders, and infectious agents will require this type of selectivity.

Enzymes can be classified into broad families or classes having similar activities, with each enzyme having a specific function. For example, many proteins phosphorylate their
20 substrate. These enzymes are broadly labelled as kinases. A myriad of kinases exist for a myriad of functions. Within this broad group, kinases can be subgrouped based on similarities in substrate, requirements for additional cofactors or similar amino acid residues that are targets for phosphorylation.

Within any given cell, there may be many active members of a given enzyme family.
25 If one member of the family shows aberrant activity, then it may be therapeutically advantageous to alter the activity of this single enzyme to the exclusion of other similar or related enzymes. Such is the case for the protein kinase family where aberrant phosphorylation events can be associated with abnormal cell growth and regulation. This is observed in proto-oncogene related cancers. For example, the pp60^{c-src} protein, needs
30 to be controlled to the exclusion of other protein kinases in order to maintain normal cell metabolism.

Protein phosphorylation as a mechanism for regulating protein activity was first recognized in 1955 with glycogen phosphorylase. Protein phosphorylation and dephosphorylation is widespread and impacts nearly all aspects of growth and homeostasis

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in the eukaryotic cell. Protein kinases catalyze the transfer of the γ -phosphate of MgATP to a protein substrate. The protein kinases, constitute a large and very diverse family of enzymes. Although these enzymes differ in size, substrate specificity, mechanism of activation, subunit composition, and subcellular localization, all, nevertheless, share a homologous catalytic core that has been conserved throughout evolution.

It is not yet possible to regulate a given enzyme at will. While there are hundreds of different protein kinases, only a few of these can be readily purified. Moreover, even among those enzymes that can be purified, many cannot be used for X-ray crystallographic studies. The sequences of many enzymes have been cloned and expressed; however, not all of these are chemically active. Therefore, even if a molecule that cannot be readily purified is cloned and expressed, it may not be functional and thus, would not provide an adequate model for structural studies. Further, even if a recombinant protein is functional, it may not be readily crystallizable. These and other roadblocks have heretofore prevented the design or identification of effector molecules directed to a particular enzyme. Thus, heretofore it has not been possible to provide a method for the design of effector molecules for a given member of an enzyme family.

Brief Description of the Figures

Figure 1 illustrates the general topological arrangement of the catalytic subunits from a number of different protein kinases.

Figure 2 diagrams the placement of the catalytic region within various members of the protein kinase family.

Figure 3 is a stereo view of the electron density for the structure determination. Figure 3A provides the density calculated to 2.7 Å. Figure 3B provides the density calculated with 10.0 to 2.7 Å refined model phases.

Figure 4 is a stereo view of the C- α backbone and includes twenty residues of PKI(5-24).

Figure 5 provides data on the location and orientation of MgATP. Figure 5A illustrates the general localization of MgATP. Figure 5B is a close-up of difference density enclosing an oriented model of AMP.

Figure 6 is an overall two dimensional topology diagram for the C-subunit. of cAPK.

Figure 7 provides stereo views of selected conserved areas.

Figure 8 illustrates the conserved catalytic core of c-AMP dependent protein kinase. Figure 8A is a space-filling model of the catalytic core. Figure 8B is a diagram of the

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conserved catalytic core using the RIBBON program of the PAP package. Figure 8C is a space-filling model identical to A, but includes PKI(5-24).

Figure 9 diagrams the conformation of bound PKI (5-24).

5 Figure 10 illustrates the high affinity binding site interactions between the catalytic subunit and the inhibitor peptide. In Figure 10A the P-11 Phe ring (numbered 356) is shown in relation to three nearby hydrophobic residues of the C-subunit. Figure 10B illustrates the interactions of the P-11 Phe and P-6 Arg sidechains with the C-subunit.

Figure 11 summarizes the interactions that contribute to recognition of the common consensus region of the inhibitor peptide.

10 Figure 12 illustrates the consensus recognition site binding interactions. Figure 12A is an illustration of the electron density corresponding to the anionic P-3 site. Figure 12B illustrates the electron density of the P-2 Arg side chain. Figure 12C illustrates the electron density of the P+1 Ile sidechain.

15 Figure 13 illustrates the catalytic site area. Figure 13A provides the site of catalysis together with the probable catalytic base sidechain of Asp 166 near the β -C of the P Ala. Figure 13B diagrams the consensus recognition site residues Arg-Arg-Asn-Ala-Ile. Figure 13C shows the phosphate of Thr 197 buried among sidechains of cationic and H-bonding residues.

20 Figure 14 is a schematic illustrating the relationship of invariant amino acids at the active site.

Figure 15 is a schematic of the conserved and variable regions of the catalytic subunit.

Figure 16 illustrates the amino acids present in PKI(5-24) that provide important interactions with cAPK.

25 Figure 17 provides a list of the coordinates that define the three-dimensional template.

Figure 18 provides photographs of the crystal forms.

Summary of the Invention

30 In accordance with one aspect of the present invention, there is provided a method of designing a highly specific affector which exerts an effect on the activity of a first enzyme. In this method, the first enzyme is a member of a class of enzymes having a conserved catalytic core. The method comprises the following steps: identifying a second enzyme that is a member of the class in which a first affector can affect the activity of the second enzyme, forming a first complex of the first affector and the second enzyme, obtaining data

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regarding the conformation of the second enzyme at sites greater than 5 Å from the site of catalysis of the second enzyme in the first complex, and designing an effector which induces a conformation on the first enzyme at sites greater than 5 Å from the site of catalysis thereof which is homologous to the conformation of the second enzyme at homologous sites in the first complex, when the effector is formed as a second complex with the first enzyme. Preferably, this method additionally comprises crystallizing the first complex and obtaining X-ray crystallography data therefrom. In a preferred form of this method, all of the members of the class have related functions, and the catalytic cores of all of the members of the class have conserved amino acid residues. In this form of the method, preferably the designing step comprises designing an effector having homologous topography and charge fields that complement the catalytic core of the first enzyme and capable of inducing a conformation wherein the conserved amino acid residues of the first enzyme are in homologous locations to the second enzyme in the first complex. The effectors can be inhibitors, activators or other effectors of enzyme activity. The first effector can be all or a portion of the first enzyme, and the first complex can be a holoenzyme. The class of enzymes can comprise protein kinases or any other suitable class. The second enzyme can be a viral oncogene product or a cellular homologue thereof, such as p60 v-Src from RSV or its cellular homologue, pp60 c-src. The second enzyme can also be cAMP-dependent protein kinase. The second enzyme can be a native mammalian protein kinase or a recombinant protein kinase. In a preferred form of the method, the designing step comprises identifying a potential effector likely to induce a conformation on the catalytic core of the first enzyme which is homologous to the conformation of the catalytic core of the second enzyme in the first complex, when the effector is formed as a second complex with the first enzyme, and determining whether the potential effector induces the conformation through the use of a technique selected from the group consisting of computer modelling, small angle neutron scattering, and circular dichroism. In this preferred method, the potential effector comprises a peptide, and the potential effector comprises a molecule selected from the group consisting of nucleotides, polynucleotides, alcohols, polymers, lipids, carbohydrates, sugars, native or synthetic molecules, protein and organic compounds and combinations thereof. In accordance with this aspect of the invention, the method can include producing the effector. Thus, the present invention also includes the effector produced from the method.

In another aspect of the present invention, there is provided another method of designing a highly specific effector which exerts an effect on the activity of a first enzyme.

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In this method, the first enzyme is also a member of a class of enzymes having conserved residues at an effector binding site. This method comprises the following steps: identifying a second enzyme that is a member of the class in which a first effector can affect the activity of the second enzyme, the first effector having a dissociation constant with the second enzyme of less than 1 μ M, forming a first complex of the first effector and the second enzyme, obtaining data regarding the conformation of the effector binding site of the second enzyme in the first complex, and designing an effector which induces a conformation on the effector binding site of the first enzyme which is homologous to the conformation of the effector binding site of the second enzyme in the first complex, when the effector is formed as a second complex with the first enzyme. In one form of this method, the class of enzymes has a nucleotide binding site and each of the effectors is capable of binding to the nucleotide binding site.

In still another aspect of the present invention, there is provided another method of designing a highly specific effector which exerts an effect on the activity of a first enzyme. In this method, the first enzyme is also a member of a class of enzymes having a conserved catalytic core. This method comprises the following steps: identifying a second enzyme that is a member of the class in which a first effector can affect the activity of the second enzyme, forming a first complex of the first effector and the second enzyme, the first complex having at least three points of contact between the first effector and second enzyme, obtaining data regarding the conformation of the catalytic core of the second enzyme in the first complex, and designing an effector which induces a conformation on the catalytic core of the first enzyme which is homologous to the conformation of the catalytic core of the second enzyme in the first complex, when the effector is formed as a second complex with the first enzyme.

Still another aspect of the present invention provides a crystallized protein kinase/effector complex having stable decay characteristics over 15 minutes and a crystallized protein kinase/effector complex having a Bragg spacing diffraction limit of less than 4Å. Preferably, the crystallized protein kinase of this aspect of the invention exhibits both of these characteristics. The present invention also provides a crystallized complex of a cAMP-dependent protein kinase and an inhibitor thereof. This crystallized complex can be used in an X-ray crystallography procedure to produce data regarding the three dimensional structure of the cAMP-dependent protein kinase in the complex, and this data can be used for designing an inhibitor of a second protein kinase which will induce a similar three dimensional structure on the active site of the second protein kinase as the three

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dimensional structure of the cAMP-dependent protein kinase in the complex. Thus, the present invention also includes an inhibitor designed by this method.

Another preferred method of the present invention involves preparing a highly specific effector of a first enzyme, with the first enzyme being a member of a class of enzymes having a conserved catalytic core with a plurality of invariant residues. This method comprises the following steps: identifying a second enzyme that is a member of the class and having a known effector thereof, forming a first complex of the second enzyme and the known effector, obtaining data regarding the three dimensional coordinates of the invariant residues in the first complex, the coordinates forming a template, generating a model wherein the first enzyme is in a conformation in which the invariant residues are in substantially the same conformation as in the template, identifying a change in the variable residues in the catalytic core of the first enzyme in the conformation of the template when compared to the variable residues in the catalytic core of the second enzyme in the conformation of the template, preparing a modified form of the second enzyme, wherein the modified second enzyme includes a non-conserved change identified through this method, and designing an effector which computer modelling indicates will induce a conformation on the catalytic core of the first enzyme in which the invariant residues are in substantially the same coordinates as the coordinates of the template, when the first enzyme is formed as a second complex with the effector designed in this step. Preferably the identified change is a non-conserved change in the variable residues. In a preferred form of this method, the method also includes forming a third complex of the modified second enzyme and an effector capable of binding thereto, obtaining data regarding the three dimensional coordinates of the invariant residues in the third complex, and using the data obtained in the previous step to design an effector which computer modelling indicates will induce a conformation on the catalytic core of the first enzyme in which the invariant residues are closer to the coordinates of the template than the conformation induced by the effector designed previously, when the first enzyme is formed as a fourth complex with the effector designed in this step. The effector used for computer modelling can be the known effector. Preferably, the method also includes modifying the computer modelling in light of the data obtained through the method prior to designing the effector. Amino acid sequence data relating to the catalytic cores of the first and second enzymes is preferably obtained. Site directed mutagenesis of a recombinantly produced second enzyme can be used in accordance with the method. In one preferred aspect of this method, the coordinates of the template are substantially as shown in Figure 17 and the template can include

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coordinates separated by the distances substantially as shown in Table 4. The effectors can be inhibitors or other effectors. The method can also include preparing the designed effector. Thus, the present invention also includes the effectors prepared through this method, and also includes pharmaceutical compositions containing these effectors.

5 The present invention also includes a method of designing a specific inhibitor for a protein kinase, comprising the following steps: obtaining data regarding the three-dimensional structure of a first protein kinase, and using the data in the design of an inhibitor for a second, different, protein kinase. The first protein kinase is preferably cAMP dependent protein kinase or an analogue thereof. The obtaining step preferably
10 comprises crystallizing a complex of cAMP dependent protein kinase and an inhibitor thereof, and additionally includes obtaining X-ray crystallography data on the crystallized complex obtained from the crystallizing step. Thus, information regarding the structure of the crystallized complex obtained in the crystallizing step through circular dichroism, small angle neutron scattering, or other chemical procedures can be obtained.

15 In a preferred form of the present invention, there is provided the use of the data of Figure 17 or of Table 4 in the design of an effector for a protein kinase.

 Still another aspect of the present invention involves a method of preparing a highly specific inhibitor of a first enzyme. The first enzyme is a member of a class of enzymes having a conserved catalytic core with a plurality of invariant residues. This method
20 includes the following steps: (a) identifying a second enzyme that is a member of the class and having a known first inhibitor thereof, (b) forming a first complex of the second enzyme and the first inhibitor, (c) obtaining data regarding the three dimensional coordinates of the invariant residues in the first complex, (d) designing a second inhibitor which computer modelling indicates will induce a conformation on the catalytic core of the first enzyme in
25 which the invariant residues are in substantially the same coordinates as the coordinates obtained in step (c), when the first enzyme is formed as a second complex with the second inhibitor, (e) preparing the second inhibitor, (f) forming a third complex of the second inhibitor and a third enzyme complexable therewith, the third enzyme having a plurality of the invariant residues, (g) obtaining data regarding the three dimensional coordinates of the invariant residues in the third complex, and (h) using the data obtained from step (g) to
30 design a third inhibitor which computer modelling indicates will induce a conformation on the catalytic core of the first enzyme closer to that in which the invariant residues are in substantially the same coordinates as the coordinates obtained in step (c) than induced by the second inhibitor, when the first enzyme is formed as a fourth complex with the third

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inhibitor. This first inhibitor is in one embodiment of this method an inhibitory domain of the second enzyme. The third enzyme preferably contains at least 5 invariant residues, and can be a naturally occurring enzyme or a mutant enzyme.

5 Still another aspect of the present invention involves a method of determining the efficacy of a first affector in affecting a first enzyme that is a member of a class of enzymes having a plurality of invariant residues among the members of the class. This method includes the following steps: determining the three dimensional coordinates of the invariant residues of a second enzyme in a first conformation wherein the second enzyme is in a complex with a second affector that is a strong affector of the enzyme, determining the three
10 dimensional coordinates of the invariant residues of the second enzyme in a second conformation wherein the enzyme is in a conformation other than the first conformation, identifying the mobile invariant residues of the enzyme, the mobile invariant residues being those invariant residues at coordinates substantially different in the first conformation than in the second conformation, determining the three dimensional coordinates of the mobile invariant residues of the first enzyme when the first enzyme is in a conformation wherein the first enzyme is in a complex with the first affector, comparing the three dimensional coordinates of the mobile invariant residues of the first enzyme in the conformation with the coordinates of the mobile invariant residues of the enzyme in the first conformation, wherein relatively small differences in the coordinates indicate relatively high efficacy of the
15 first affector. The step of determining the coordinates of the first enzyme in the conformation is preferably performed using computer modelling of the conformation. The steps of determining the first and second conformations preferably comprise obtaining X-ray crystallographic data of the enzyme. The second conformation can be a conformation produced by a ternary complex, such as one comprising a protein kinase, a nucleotide and an affector. The second conformation can also be a conformation produced by the second
20 enzyme not complexed with a ligand, or the same enzyme as the first enzyme.

In an additional aspect of the present invention, there is provided a method of designing specific inhibitors of a first protein kinase having a template defined by a plurality of invariant residues present in substantially all protein kinases. This method comprises the
30 following steps: obtaining data regarding the three dimensional coordinates of the invariant residues of a second protein kinase and of the points of contact between the second protein kinase and a known inhibitor thereof, the coordinates being obtained when the second protein kinase is formed as a complex with the known inhibitor, generating a model of the first protein complex wherein the template is defined by the positions of the invariant

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residues in the complex, examining the amino acid residues present in the first protein kinase at positions corresponding to the points of contact in the complex, and designing an inhibitor of the first protein kinase capable of forming ionic and hydrophobic interactions with the amino acid residues. The method of Claim 62, wherein the second protein kinase is cAMP dependent protein kinase. The known inhibitor can be PKI(5-24). For this known inhibitor, the points of contact in the complex preferably comprise positions corresponding to the positions selected from the group consisting of P+1, P-2, P-3, P-6 and P-11 along the known inhibitor. The positions corresponding to the points of contact in the examining step preferably comprise positions within a sphere having a radius of 11 Å, more preferably 6 Å, from the coordinates of the point of contact obtained in the obtaining step. The designing step preferably additionally comprises designing the inhibitor to form appropriate hydrogen bonding with the amino acid residues.

Further details concerning the present invention are provided in the following detailed description.

Detailed Description of the Invention

CITED REFERENCES INCORPORATED BY REFERENCE

A number of articles are specifically cited herein as providing background information useful, but not essential, to those of ordinary skill in the art in the practice of the present invention. As such, the disclosure of each of these articles is hereby explicitly incorporated by reference.

INTRODUCTION

The protein kinase family of enzymes is used as a model for this invention. These enzymes are involved at all levels of regulation in the eukaryotic cell. They act as "transistors" for the cell, receiving signals and amplifying the message inside the cell. Protein kinases receive hormone signals from outside the cell. They are involved in cell growth, for cellular homeostasis, and for triggering the steps of mitosis.

In addition, many oncogenes code for protein kinases. These oncogenic protein kinases are also very diverse in their structure and location within the cell. However, all are derived from normal cellular components and all, in one way or another are defective in their ability to be turned off. In other words, they are constitutively active in contrast to their protooncogene counterparts which are turned off in the absence of the appropriate signal. Thus, protein kinases are not only an essential part of normal cell growth and division, but, can lead to oncogenesis when their normal function becomes genetically impaired.

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Diversity is a hallmark of the protein kinase family. For example, growth factor receptors, such as the insulin receptor, are large proteins with a major extracellular domain for binding growth factor, a single membrane spanning domain, and an intracellular protein kinase domain that is activated in response to growth factor binding. The kinase activity is limited to a specific domain of the protein. Control of the insulin receptor may play an important role in the control of diabetes. Protein kinase C is activated by diacyl glycerol and Ca^{2+} and is also activated by the tumor promoting phorbol esters. It is a cytoplasmic protein that in its active state is associated with the plasma membrane. Another protein kinase, cdc2, associates with cyclin B and is an essential trigger for mitosis. The transforming protein in Rous Sarcoma Virus, pp60^{v-src} is anchored to the cytoplasm surface of membranes. In spite of the diversity in size, subunit composition, location in the cell, and mechanism of activation, all protein kinases share a common enzymatic activity and a conserved catalytic core, indicating that all have likely evolved from a common functional precursor. Thus, one aspect of the present invention provides a method for developing highly selective inhibitors for members of the protein kinase family.

The first protein kinase to be purified was phosphorylase kinase. The second was phosphorylase kinase kinase, later renamed cAMP-dependent protein kinase (EC2.7.1.37:ATP:protein serine phosphotransferase) when its broader substrate specificity was appreciated. Not only was cAMP-dependent protein kinase (cAPK) one of the first protein kinases to be characterized, it also is one of the simplest and best understood biochemically. Its simplicity is due primarily to its mechanism of activation, which involves subunit dissociation. With the exception of the oncogenic enzymes, all protein kinases typically are maintained in an inactive state in the absence of the appropriate activating signal. In the case of cAPK, the ligand triggering activation is cAMP, one of the first recognized second messengers for hormone signalling. In the absence of cAMP, the enzyme is sequestered as an inactive holoenzyme containing two regulatory (R) and two catalytic (C) subunits. When intracellular cAMP levels are elevated, the cyclic nucleotide binds to the R-subunit, thus causing the complex to dissociate into a R_2 dimer and two free and active C-subunits. The general consensus sequence recognized by the C-subunit is Arg-Arg-X-Ser/Thr-Y, where X is any small residue and Y is a large hydrophobic group. The conserved catalytic core found in all protein kinases is contained within this relatively simple monomeric C-subunit.

This invention provides the first crystal structure of a protein kinase with its catalytic subunit intact. Knowledge of the conformation of the catalytic structure of cAPK is central

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to the understanding of protein kinase activity. Not only is the structure of the cAMP-dependent protein kinase catalytic site provided, but, the crystals contain a bound inhibitor peptide. This inhibitor peptide, PKI(5-24), is a fragment of the heat stable protein kinase inhibitor (PKI). This peptide includes the consensus features common to all peptide
5 substrates and inhibitors of cAMP-dependent protein kinase. In addition, it contains other features that convey unique high affinity binding characteristics. Thus, precise properties of binding and interaction are described. From this data, a template is derived from which all other protein kinases can be modelled and from which other inhibitors can be designed.

One of the more important questions regarding protein phosphorylation is how the
10 targeted protein substrate is recognized by a specific protein kinase. This question has remained particularly elusive until now because the determinants for peptide recognition are widely dispersed and in some cases well-removed from the actual site of phosphotransfer. Owing to its simplicity as well as its relative ease of purification, the catalytic or C-subunit of cAMP-dependent protein kinase serves here as a prototype for identifying functional sites
15 that are involved in substrate recognition and catalysis. Chemical analyses and procedures, such as affinity labeling, group specific labeling, and fluorescence energy transfer all have provided clues about regions involved in peptide recognition, MgATP binding, and catalysis. Substrate analogues provide indirect information about binding sites important for effector molecule specificity. Further NMR, circular dichroism, small angle neutron scattering
20 (SANS) and other chemical procedures offer further insight into the structure of the enzyme. However, X-ray crystallography provides a comprehensive three dimensional structure that can confirm and integrate these other techniques.

The expression of the C-subunit in *E. coli* not only facilitated these structural studies, but also has permitted recombinant approaches to be used to further modify the active site
25 of cAPK and thereby mimic the reactive site of other protein kinases. Information to aid in these studies is obtained from sequence data available for the protein kinase family. Hanks et al., Science 241: 42, 1988, is one source of such data. Such sequence comparisons have identified highly conserved regions including several invariant residues, variable regions, and places where inserts and deletions can be tolerated. Both chemical and
30 sequence information are used here to verify the structure data obtained from the X-ray diffraction studies. As will be disclosed herein, this body of information permits the design of other effector molecules specific for other protein kinases. Further, this information serves as guidelines for the design of specific effector molecules for enzymes from a wide variety of enzyme families.

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The existing basis for the design of specific inhibitors for protein kinases, in the absence of the three dimensional structure provided herein, relies on the use of synthetic peptides based primarily on the sequences of known substrates and inhibitors. In the case of cAMP-dependent protein kinase, there are some very specific high affinity peptides available. Existing inhibitors also include nucleotide and nucleoside derived compounds found through traditional means. However, these nucleoside and nucleotide inhibitors do not generally exhibit the type of specificity observed with peptide inhibitors. In general, such specific peptide inhibitors are not available for other protein kinases. Specificity for cAMP-dependent kinase improves with the addition of amino acids postulated to lie outside of the catalytic core. We have discovered that these regions are also important for inhibitor design. Knowledge of these sites provides a "lock" to permit for the first time the tailoring of inhibitors for any given protein kinase. Thus, one important aspect of the invention lies in the design of the "lock", that requires an understanding of the three dimensional structure of the complex of the catalytic subunit of cAMP-dependent protein kinase, with its very potent specific inhibitor, PKI(5-24).

Disclosed herein is a template gleaned from the crystal structure of the catalytic subunit of cAMP-dependent protein kinase. Just as the chemical information derived from the C-subunit serves as a framework for interpreting the entire kinase family, the structure of cAPK provides information for the creation of a template for viewing the conserved catalytic core of all eukaryotic protein kinases. This invention further provides a model for the identification and design of molecules capable of interacting with the catalytic core of a given enzyme by analyzing the conserved catalytic core of another member of that enzyme class.

X-RAY CRYSTALLOGRAPHY

X-ray crystallography permits three dimensional molecular analysis of a protein at the atomic level. Analysis requires the production of crystals and crystal production requires a pure concentrated product. Further, complexes of a protein of interest together with a second interacting molecule provides information on the conformational changes occurring within a protein in response to that second molecule. X-ray crystallography of a protein with its substrate, an antibody or a drug can provide information for rational drug design.

An X-ray diffraction pattern taken from a crystal looks like an array of spots of varying intensities. Each spot is related to one of the Fourier coefficients of the electron density pattern in the crystal. Thus, the electron density in the crystal can be reconstructed if a sufficient number of diffraction spots can be measured and the relative phase angles of

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the Fourier coefficients can be determined. Thus, a crystallized enzyme used in the practice of certain aspects of the present invention should be of sufficient quality to obtain these measurements. For example, the spots of varying intensity in the diffraction pattern decay over time. It is quite difficult to work with diffraction patterns with half lives of less than 5 10 hours. However, it is possible to work with diffraction patterns having half lives as short as about 15 minutes to 3 hours, depending on the amount of structural data desired to be obtained. Further, it is believed possible to work with crystals of even shorter half lives using equipment and computer programs more advanced than commonly available today. Additionally, not all crystals are of equal quality and poor crystals have large Bragg spacing 10 diffraction limits. Thus, a workable crystal should have a Bragg spacing diffraction limit of less than 4 Å.

Determination of phase angles uses isomorphous replacement to insert atoms into defined positions in the crystal for diffraction data measurement. These angles provide information that permit the production of an electron density map. The map is then used 15 to build an atomic model from which three-dimensional coordinates are measured that define the structure of the crystallized molecule.

MODEL SYSTEM

X-ray crystallography has been employed for the rational design of drugs and other interacting molecules. However, to date, the rational design of effector molecules has been 20 limited to a study of the active site of the protein/effector molecule interaction. Potential effectors designed from this information have not been obtained by looking at interactions beyond the active site. We believe that these interactions assist in binding and thereby contribute to binding specificity. Thus, we have discovered that it is these interactions in concert with information obtained from the active site that make the design of specific 25 effector molecules a possibility. Moreover, this information additionally permits the design of specific effector molecules for related but nonidentical enzymes.

Many enzymes within a cell have evolved from common progenitors. These enzymes share common enzymatic activities and one example is the protein kinase family. Since the functions of the enzymatic families or classes are broadly conserved, at least a portion of the 30 catalytic site is also conserved. Therefore to a large extent rational drug design relies on the identification of the familial similarities and hence drugs are designed to react broadly within a given family or class.

While all members of an enzymatic class may provide a similar activity, such as phosphorylation or dephosphorylation, each member may have only one specific target.

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Thus, successful rational drug design based on group similarities would provide molecules that also interact broadly. Where there are many members of an enzymatic class acting specifically within a restricted locale such as a single cell, a broadly acting drug would interact with any number of enzymes from the group. Thus, the interaction would be
5 general and not specific. In contrast, the present invention advantageously is capable of providing effectors with highly specific interactions for a given member of an enzyme class.

Previous methods for rational drug design require the crystallization of the target molecule of interest. However, the production of useful crystals is both difficult and time consuming. It first depends on the ability of the target molecule to be isolated and purified
10 in sufficient quantity for crystallization. A large number of crystallization conditions often need to be tested and once a crystal is made that is of sufficient quality, additional crystals often need to be produced in order to have enough material for analysis. Further, not all molecules are readily purified or readily crystallized. Advantageously, the present invention discloses a method whereby only one enzyme within a family of enzymes need be
15 crystallized.

This invention teaches a method for the identification and design of specific molecules interacting with a specific enzyme wherein the specific enzyme is a member of a broadly acting enzymatic group or class.

The particular enzyme class chosen for this invention is preferably one that has the characteristics generally associated with an enzyme class developed from a divergent
20 evolutionary pathway. That is, an enzyme class in which it is possible to identify similarities within the catalytic core of all members of the class. Enzymes with similar activities that have evolved from convergent evolution will not necessarily share these constant residues and a model or template employing invariant amino acids as anchors would then not be
25 possible. A variety of enzyme families are postulated to arise from divergent evolution, and thus would be expected to serve as a preferred class of enzymes for design of effector molecules within the context of the present invention. Such enzyme families include, but are not limited to, the protein kinases, phosphorylases, and several groups of proteases.

For purposes of illustration only, the present invention is described using the protein
30 kinase family as a model system. As discussed above, these enzymes are essential for many aspects of cell regulation. Over 100 individual protein kinases have been identified. Thus, the successful design of effectors to manipulate the activity of a kinase can provides an invaluable tool for research as well as for the design of a wide variety of therapeutics and diagnostics.

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Thus, for example, development of effective specific inhibitors of oncogenic kinases is believed to lead to the development of anti-neoplastic treatments. In addition, specific inhibitors of kinases involved in hormone regulation will be useful in artificially regulating the secretion and regulation of such hormones. Also, since many neuro-transmitters are regulated by kinases, development of new effectors could potentially impact on diseases of the nervous system. Further, platelet aggregation and clot formation might also be regulated through novel effectors of kinases developed through the methods of the present invention. Many other therapeutics are believed possible through the development of novel specific effector molecules.

The model system used in connection with this invention uses cAMP-dependent protein kinase together with a 20 amino acid inhibitor peptide, PKI(5-24), to establish a "lock" for specific effector design. This inhibitor is unique in that it interacts only with the cAMP-dependent protein kinase. Therefore, cocrystallization of this inhibitor with cAMP-dependent protein kinase permits the visualization of the conformation of an enzyme in association with its specific inhibitor.

The general requirements for peptide recognition by the catalytic subunit were characterized originally using synthetic peptides. From those studies evolved a general consensus sequence that includes two basic residues, typically arginine, followed by one intervening small residue preceding the site of phosphotransfer. The phosphate acceptor site, either Ser or Thr, is followed immediately by a large hydrophobic residue. The typical peptide substrate used for this kinase, based on the phosphorylation site in pyruvate kinase, is shown in 1. Replacement of the Ser with an Ala generates an inhibitor peptide with poor affinity. In contrast is the inhibitor peptide, PKI(5-24), derived from the thermostable protein kinase inhibitor (PKI). This peptide, seen in 1, encompasses the consensus sequences described above but also contains additional unique features that convey high affinity binding. The particular features that contribute to high affinity binding were defined using peptide analogues, and the most relevant ones are also indicated in 1. A general structure of the peptide in solution was deduced using circular dichroism (CD) and NMR spectroscopy. The peptide, PKI(5-24), was co-crystallized with the catalytic subunit of cAPK, and the structure of that peptide as well as its interaction with the protein are discussed here.

The folding of the polypeptide chain and the mechanism of catalysis is conserved in all protein kinases. There are 8-9 invariant residues scattered throughout the core for all protein kinases. The crystal structure reveals that most of these invariant residues are

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clustered three dimensionally around the site of catalysis providing an interconnected network. The regions involved in peptide recognition extend over a wide area on the surface of the enzyme and until this structure was solved there was no understanding of the details of the peptide recognition sites. The structure of the catalytic subunit thus serves
5 as a framework from which a template for the entire protein kinases family can be produced. This structure provides, for the first time, a true molecular basis for the design of effectors that will selectively target any given protein kinase. Thus, it is an object of this invention to provide a method for the identification and design of molecules interacting with the catalytic core of a protein kinase by preparing a template from the analysis of the
10 catalytic core of the cAMP dependent protein kinase.

The ability to design effector molecules that act on a given enzyme using information obtained by X-ray crystallography is dependent on the formation of crystals of purified enzyme. Methods for crystal production vary greatly and one cannot predict how readily a given molecule or complex will crystalize. However, those skilled in the art will recognize
15 that a variety of methods for crystallizing can be attempted for any given enzyme, and that successful crystallization can be expected of a variety of enzymes. Rational drug design additionally requires information about the interaction of a known effector in order to accurately predict a potential effector's effect on the catalytic core of the enzyme. Thus, crystals of the complex of effector molecule and enzyme together are used to gather
20 information on the conformation of the enzyme in its inhibited conformation. Thus, in addition to information about the catalytic core of an enzyme family and the identification of additional sites adjacent to the core that permit the specific design of inhibitors, the present invention provides an improved method for the crystallization of complexes.

An important feature of certain aspects of this invention is the production of an enzyme/effector template. In order to generate this template, the effector chosen for
25 production of enzyme/effector complex should have a high affinity for a particular enzyme. The initial effector molecule chosen should preferably have a K_d less than 1 μM , and more preferably less than 100 nM, in order to provide a conformation resulting from high affinity interactions. Once the specific interactions are understood it is contemplated that effector
30 molecules having a variety of K_d ranges could be selectively designed for various purposes. Thus, in the model system chosen to illustrate this invention, PKI(5-24) is used as an effector of cAPK, with a K_d of approximately 60 nM. Those of ordinary skill in the art will recognize that other inhibitors with K_d less than 100 nM could also have been chosen to illustrate this aspect of the present invention.

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For example, cGMP dependent protein kinase has an inhibitor with a K_i of approximately 6 nM, and an inhibitor for cAPK described by Ricouart et al. is characterized in the 4 nM range (J. Med. Chem., 34: 73-78, 1991). These K_i values are roughly equivalent, however, not identical, to the expected dissociation constants (K_d 's).

5 The PKI(5-24) inhibitor peptide used here is highly specific and is rather large in that it extends beyond the catalytic core. Other known peptides and effector molecules for kinases are not as specific. We have discovered that the interactions beyond the catalytic core provide the high specificity of PKI(5-24) for cAPK.

10 As stated above, it is the crystallization of the enzyme with its specific inhibitor together with the analysis of the relationship of the inhibitor to both the catalytic core and to areas surrounding the core that provide data for the particular protein kinase "lock". The "lock" comprises the three dimensional structure and ionic, hydrophobic, hydrogen bonding and other interactions of the non-conserved variable residues with the specific effector structure. The lock is defined by the invariant residues of the exemplary structure. When
15 an enzyme of the class is affected by a specific effector, the backbone atoms of these invariant residues must be in substantially the same relative coordinates in all members of the enzyme class. Thus, with knowledge of sequence information of the particular enzyme for which the effector is being designed, knowledge of the lock formed by the invariant residues can be obtained. The coordinates of the invariant residues position the variable
20 residues of the lock in space and thereby permits the design of other specific inhibitors and effector molecules for other protein kinases.

 The lock consists of the site of phosphotransfer (P site) with recognition sites for flanking sequences. The flanking sites can be identified by the number of amino acid residues separating that site from the P site. Thus, the first amino acid residue in the
25 direction moving toward the carboxy terminus is designated P+1, and the following residues are designated P+2, P+3, P+4 and so on. Similarly, the residues on the side moving toward the amino terminus are designated P-1, P-2 and so on.

 The sites for recognition of the peptide are not identical between members of the protein kinase family, and the chemical content is unique for each given protein kinase. The
30 sequence of the given protein kinase is built into the coordinates of the C-subunit using the invariant residues. The position of these invariant residues can be identified using X-ray crystallographic data, such as the data disclosed herein in Figure 17. This data provides the coordinates for each non-hydrogen atom in cAPK. It is the locations of these invariant residues which serve to define the template common to all protein kinases. This template

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can then be used to model the three dimensional coordinates of the variable as a basis to design highly specific effector molecules.

5 The effector molecules to be designed could be polypeptides, nucleic acids and their analogues, combinations of nucleotides and peptides, organics or any other molecule capable of specific interaction with a given enzyme. The essence of the design of a specific inhibitor for a given protein kinase is based on the three-dimensional fit of the specific inhibitor into the provided "lock", or template, provided by the known structure of cAPK.

10 The template defined by the invariant or other highly conserved residues can be used to define the region immediately flanking the phosphorylation site and, in addition, can incorporate more distant parts of the molecule to enhance specificity and affinity. The peptide recognition site serves in the same manner as the antigen recognition site of an antibody. This site extends over a large surface of the enzyme and provides a unique lock for the design of a wide variety of effector molecules, including both peptide and non-peptide effectors. Each particular protein kinase has a different and unique chemical content at each individual site. Thus, the "lock", is unique for each protein kinase.

15 The "lock" of any particular enzyme represents a topological map with defined sites, positions of which vary between members of the enzyme class. As an analogy, each kinase can be thought of as functioning in a manner similar to a specific antibody in that it recognizes only a very specific set of proteins to phosphorylate. However, each of the kinases has a conserved template, the positions of which will not substantially vary between kinases in an inhibited conformation. Thus, using computer modelling together with known sequence information regarding a particular kinase, the invariant residues of the kinase can be placed in the template conformation, and the approximate positions of the variable residues can be predicted.

25 The lock provides the information from which other specific effector molecules can be designed. It provides information on topology, charge interactions and the points of contact both within the catalytic core and around the core that suggest the design features important for the production or identification of novel effector molecules. Thus, the goal is to design an effector having homologous topography and charge fields that complement the catalytic core of the lock of the enzyme. Computer modelling can be used with these factors to design an effector capable of inducing a conformation where the conserved amino acid residues of the enzyme are in homologous locations to the template.

30

BRIEF DESCRIPTION OF EFFECTOR DESIGN

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The basic steps toward achieving this invention are provided briefly here and in detail below. A class of enzymes is first identified wherein at least one enzyme of the class has a highly specific affector molecule. Then the inhibitor is tested for specificity and, preferably, the inhibitor sequence is reduced in size until a minimum sequence having the
5 desired specificity is obtained. Sequence data from related enzymes is analyzed so that a consensus region that forms the catalytic core can be identified. Crystals of affector molecule together with the model enzyme are subjected to multiple isomorphous replacement techniques to prepare heavy atom derivatives. This permits the location of heavy atoms within the structure to be identified and additionally permits multiple
10 diffraction patterns to be combined to deduce phase angles for calculation of the electron density of the structure. Those of ordinary skill in the art will recognize that other techniques can be used to deduce phase angles and to improve the accuracy of previously deduced phase angles.

A three-dimensional structure can be obtained from the electron density data using
15 a computer program such as TOM/FRODO. Further, a computer program, such as X-PLOR, can be used to improve the accuracy of the initial three-dimensional structure. There are a variety of computer programs available for analyzing X-ray crystallographic data. Those used in the development of the model system for this invention are cited herein. Those of ordinary skill in the art will recognize that many other such computer programs
20 providing similar functions could also have been used. From this data, the points of contact are identified both within the catalytic core and the surrounding region. Invariant amino acids and consensus recognition sequences are identified. The data is further analyzed against available chemical data such as NMR, CD, SANS data and other data resulting from chemical procedures. This chemical data can provide additional information for the
25 structural model.

The coordinates of the invariant amino acids residing in the conserved catalytic core and the surrounding invariant residues in the enzyme/affector complex provide the template to be duplicated in other members of the enzyme class. The lock of the enzyme for which the affector is to be designed is then built by replacing the variable amino acids of the
30 catalytic subunit of the template enzyme with the amino acids of the new enzyme. Any gaps in the sequence alignment between the enzyme used to generate the template and the enzyme for which the lock is being modelled generally occur within loops. These loop regions can be modelled separately using the structural data accumulated in a data bank, such as the Brookhaven data bank. The model of protein kinase can then be corrected and

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refined using an energy minimization procedure and using molecular dynamics to eliminate stearic and electrostatic clashes. The resulting model of the catalytic core of the protein kinase under investigation is then inspected for amino acid content of the enzyme's surface which interacts with the proposed inhibitor.

5 In accordance with one aspect of the present invention, another member of this enzymatic class can then be analyzed in the context of this template. If the new enzyme can be crystallized, then the information obtained from the crystallization is merged with the "lock" structure. However if the new enzyme is not accessible or is not crystallizable, the enzyme can still be incorporated into the three-dimensional lock. The ability to incorporate
10 the new enzyme into the template is dependent on the identification of conserved residues within the catalytic core of the new enzyme that are complementary to the conserved residues in the model enzyme. The template establishes the coordinates for these residues in three-dimensional space as well as providing coordinates for the three-dimensional surface of the catalytic core and adjacent regions.

15 For cAPK and the protein kinase family, the invariant residues are identified and summarized in the review by Hanks et al., supra. The template permits a comparison of the new enzyme catalytic core surface with cAPK. Residues within the catalytic core that are different from those of cAPK are studied to determine how those differences in the new enzyme might alter the surface of the core or change the structure of a new effector
20 molecule. Recombinant cAPK can then be subjected to site-directed mutagenesis to change residues specific to cAPK into residues found in the new enzyme. This recombinant protein can be crystallized.

 A novel effector molecule can then be synthesized that complements the electrostatic charges and topography of both the catalytic core and identified surrounding regions of
25 interest for the new enzyme. The points of contact, hydrophobic pockets, site of phosphotransfer, topography and stearic interactions are assessed and the effector molecule can then, if necessary, be subjected to random mutagenesis or site-directed mutagenesis to improve the effector/enzyme interaction. This model effector molecule together with recombinant mutated cAPK, are tested with the target enzyme for effector activity. The
30 effector molecule is finally tested with the native new enzyme. Fluorescent tags bound to the effector can be used to assess binding to the new enzyme in the cell. Alterations in enzyme function can be detected by gel electrophoresis and complexes of enzyme and effector can be isolated and purified for further analysis. Thus, new enzyme purification and crystallization is not required for effector design.

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As discussed above, generation of new effectors is not limited to peptides. A variety of chemically synthesizable compounds can be used.

The model can be tested by a variety of methods. For example, kinetic determination of inhibition constants of novel inhibitors can be measured. Also, CD, SANS and other chemical procedures can be used to assess the extent of the conformation changes due to binding of the effector. If a mutated form of the enzyme has been prepared, cocrystallization of the effector with this mutated form can be performed and the points of contact can be determined and compared with the modelled points of contact.

ENZYME FAMILY CHOICE AND IDENTIFICATION OF CATALYTIC CORE

This invention relates particularly to enzyme families formed by divergent evolution. Once an enzyme family of interest is identified, an individual enzyme is chosen from a group of enzymes that share invariant residues within their postulated active sites.

The enzymatic or active site within a given protein kinase can be broadly identified through biochemical means. When the enzyme exists as a group of subunits, enzymatic activity is often restricted to one of those subunits. Thus, prior to performing these biochemical means, the enzymatic subunit can be purified from the holoenzyme. The active site can be further localized by systematically reducing the subunit size and assessing enzyme activity with each reduction. In one method, the various mRNA sequences encoding the related enzymes are reversely transcribed and cloned. Sequence information can then be obtained from the catalytic region for a number of enzymes of the same class. Similar amino acid residues within the catalytic subunit are aligned in order to visualize homologous regions. Invariant amino residues can be identified among the class which are either present in all known members of the class or substantially all members of the class. At least a plurality of these invariant residues are believed necessary for enzyme activity within the catalytic subunit. Thus, the invariant residues can further define the catalytic core.

For the protein kinase family, invariant amino acid residues are located within the catalytic core and are boxed in by a solid line in Figure 1. Figure 1 illustrates the general topological arrangement of the catalytic subunits from a number of protein kinases.

Figure 2 illustrates that while the catalytic regions from members of the protein kinase family share some striking similarities, the placement of this active region within the enzyme, the size of the enzyme and the regulatory regions of the enzyme vary considerably. The conserved catalytic core is denoted in Figure 2 as solid black areas and regulatory regions are cross-hatched. Additional information regarding the use of cAMP-dependent

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Protein Kinase as a model for the protein kinase family can be found in a review by S. Taylor (J. Biol. Chem. 264:8443-8446, 1989.).

5 It is known from an analysis of the catalytic core of the protein kinase family that the core is included in a conserved 300-residue segment. Site-directed mutagenesis of recombinant enzyme sequences has been used to identify particular residues critical to enzymatic function. In the cAMP-dependent kinase an invariant lysine residue at position 72 has been shown to be important by site-directed mutagenesis and a triad of glycines is thought to be associated with ATP-binding.

10 The enzyme exists as a tetrameric holoenzyme composed of a dimer of regulatory subunits and two catalytic subunits. cAMP binds to the regulatory dimer yielding dissociation of the enzyme into an $R_2(cAMP)_4$ complex and two active catalytic (C) subunits. It is the active C-subunit that phosphorylates serine or threonine residues on substrates having the consensus sequence Arg-Arg-X-Ser/Thr-Leu.

AFFECTOR MOLECULE FOR TEMPLATE DEVELOPMENT

15 In a preferred form of the present invention, the enzyme used to establish the template or lock is, advantageously, a molecule that binds with high affinity to its effector, preferably with a dissociation constant less than 1 μ M. For example, there are many such known effectors, such as inhibitors and activators, of various protein kinases. Kinases with a regulatory subunit are known that are inhibited by a peptide encoding the regulatory subunit binding site. Similarly, kinases that possess an autoinhibitory portion are also known. Thus, for such a kinase, this autoinhibitory region could be cleaved away from the core enzyme, purified and analyzed to provide a minimal high-affinity inhibitory sequence.

20 There are several inhibitors of cAMP-dependent kinase. The regulatory subunits can function as physiologic inhibitors as can the heat stable inhibitor protein (PKI). These inhibitors share a substrate-like sequence based on the arginine doublet, N-terminal to the position of the phosphorylation site in a normal substrate. Peptide fragments containing the consensus sequence bind the C-subunit in a manner analogous to a real substrate. PKI has an alanine in place of the phosphorylatable residue. While the PKI sequence is clearly inhibitory, the addition of a 15 residue stretch N-terminal to PKI increases inhibitory activity. Thus, residues external to the catalytic site are believed to be relevant in providing potent, high-affinity, inhibition and for improving the specificity of an inhibitor.

30 A protein that is a high-affinity inhibitor of an enzyme can be dissected to find a smaller fragment, if it exists, that still contains high-affinity inhibitory activity. At least three factors are useful in this dissection: 1) an ability to produce chemically defined fragments

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of the larger inhibitor, either by synthesizing peptides or by cleaving the inhibitor with reagents such as cyanogen bromide or proteases, that cut at short amino acid sequences of a specific type for each reagent, 2) an ability to isolate specific fragments of the larger inhibitor from the mixture of fragments resulting from cleavage of the larger inhibitor, and
5 3) an ability to assay chemical species for inhibition of the enzyme of interest.

To carry out the isolation of a potential smaller inhibitory region of a larger inhibitor, one can cleave the inhibitor into fragments using a protease. Then one can separate the resulting fragments using HPLC and assay the fractions for high-affinity inhibition of the target enzyme. If no fraction is found that exhibits the desired inhibition,
10 the cleaving reagent may have cleaved at a location that splits the inhibitory portion of the protein, destroying its ability to inhibit. In this case, it would be desirable to obtain other cleavage patterns until an inhibitory fragment is found. After obtaining the smallest possible inhibitory fragment using proteolytic cleavage of the intact inhibitor, one can chemically sequence the fragment as a step toward further defining the smallest fragment still having
15 high-affinity inhibitory activity. With knowledge of the amino acid sequence, one can then use peptide synthesis to construct progressively shorter subsets of this fragment. These shorter subsets can then be assayed for inhibitory activity. Proceeding in this manner will thus allow definition of the smallest sequence, present in the larger inhibitor that still possesses high-affinity inhibitory activity toward the target enzyme. Methods for
20 determining inhibition constants for tight-binding inhibitors are found in Biochem. J. 127: 321-333, 1972 by P. Henderson. Methods for determining the inhibitory region PKI(5-24), are provided by Scott et al. in Proc. Natl Acad. Sci. USA 82:4379-4383, 1985.

OBTAINING THREE DIMENSIONAL STRUCTURE DATA

In order to obtain data on the conformation of the template of the enzyme formed
25 by binding of the effector thereto, a variety of techniques can be used. These techniques include, circular dichroism, small angle neutron scattering, diffraction methods, including any combination of multiple and single isomorphous replacement, single or multiwavelength anomalous scattering methods, molecular replacement methods maximum entropy phasing, solvent-flattening methods and so-called "direct" methods used primarily to solve small-
30 molecule structures. However, in the preferred embodiment, X-ray crystallography is used in order to generate specific coordinates for each of the non-hydrogen atoms in the complex. Coordinates for the hydrogen atoms could additionally be obtained using neutrons. Thus, following the isolation of an exemplary protein and effector and following or during the sequence analysis of related enzymes, crystals of enzyme and effector protein are generated.

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The crystals can be generated from enzyme purified from natural tissue or from enzyme generated by recombinant means. Provided below are examples pertaining to the production of crystals using the recombinant mouse C_{α} -subunit of cAMP dependent protein kinase and purified cAMP-dependent protein kinase from porcine heart. Nelson et al describe the purification schemes for porcine heart cAMP dependent kinase (J. Biol. Chem. 256:3743, 1981.) and Slice et al. disclose the methods for the generation of recombinant mouse C_{α} -subunit in E. coli (J. Biol. Chem. 264:20940, 1989). The sequence data for cAPK was published by Uhler et al. (J. Biol. Chem. 261:15360-15363, 1986).

The steady state kinetics of the C-subunit, purified from E. coli are identical to the mammalian C-subunit, although the E. coli protein is more labile to heat denaturation. Unlike the mammalian enzyme, the recombinant C-subunit lacks a myristoyl group at its amino terminus. For a review of protein crystallography see Protein Crystallography, 1976, T. Blundell and L.N. Johnson, Academic Press, New York. Information on circular dichroism and neutron scattering is found in Biophysical Chemistry. Part II: Techniques for the Study of Biological Structure and Function, C.R. Cantor et al. (W.H. Freeman and Co., San Francisco, 1980).

Example 1

Porcine Heart C-subunit Crystal Forms

Reagents were obtained from the following sources: threo-1, 4-dimercapto-2,3-butanediol (DTT, dithiothreitol; Aldrich, Milwaukee, WI); N,N-bis(2-hydroxyethyl)glycine (Bicine; Aldrich); methanol (Fisher Scientific); ammonium acetate (Aldrich); polyethylene glycol (Dow, Midland, MI).

The peptide inhibitor PKI(5-24) was synthesized at the La Jolla Cancer Research Foundation (La Jolla, CA) and modified in our laboratory. These modifications are described in detail below. The sequences of the peptide inhibitors are: (1)PKI(5-24); TTYADFIASGRTGRRNAIHD, (2)PKI(5-24), tyrosine iodinated: TTY*ADFIASGRTGRRNAIHD. The peptide sequence abbreviations follow either of the two standard abbreviation schemes for amino acids; the three letter code or the single capital letter designation. Both are standard abbreviations and are well understood by those of skill in the art.

The porcine C-subunit was purified to a single band on SDS-polyacrylamide gels and used for crystallization. Two crystal forms were prepared. Photographs of the porcine heart apoenzyme (cubic); and the porcine heart C:MgATP:PKI(5-24) ternary complex (hexagonal) are provided as Figures 18A and 18B.

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The first crystal form used the hanging-drop vapor diffusion method. A drop of protein mixed with precipitating agents is suspended from a microscope cover slip and allowed to equilibrate through the gas phase against a larger reservoir.

Specifically, 30- μ L drops of 3-4 mg/mL protein solution were suspended and allowed to equilibrate against approximately 1mL of reservoir solution in wells of plastic Linbro tissue culture trays over a time of several days to weeks. Both new forms, as well as the earlier P2₁ form, were grown at 4°C. The porcine heart C-subunit was concentrated to 8-12 mg/mL and subjected to a final dialysis before attempting crystallization. The specific recipe for obtaining the first new form was the following: drop- 1/3 protein in 50 mM (NH₄)₂HPO₄ and 5 mM 2-mercaptoethanol (pH 8.0-8.2); 1/3 150 mM NH₄CH₃COO, 50 mM (NH₄)₂HPO₄, and 10 mM dithiothreitol (DTT) (pH 8.1-8.2); and 1/3 reservoir composed of 8-9% PEG-400, 17-20% MeOH, and 10 mM DTT. Crystals of the second form were obtained from the same conditions as the first new form when the drop contained, in addition to the protein, MgATP and a 20-residue peptide inhibitor [PKI(5-24)] in the molar ratio 20:5:1:1 ATP:Mg²⁺:PKI(5-24):C-subunit. The same crystal form was subsequently grown from a drop containing 1/3 protein in 50 mM bicine, 100 mM NH₄CH₃COO, and 5 mM 2-mercaptoethanol (pH 8.3); 1/3 MgATP and PKI(5-24) in 10 mM DTT in the same ratio to protein as before; and 1/3 8 mM DTT and 8% PEG-400. The reservoir contained 8% PEG-400, 15-20% MeOH and 7mM DTT. The first new crystal form could also be grown in the presence of the Mg²⁺ and the non-hydrolyzable ATP analogue adenosine 5'β, γ-methylenetriphosphate (AMP-PCP). The second new crystal form, representing the ternary complex, could be grown with CoCl₂ or CdCl₂ substituted for MgCl₂ in the crystallization. The transition from one crystal form to another caused only by addition of MgATP and the peptide inhibitor PKI(5-24) suggests that a significant conformation change may occur upon their binding.

The space groups of the new crystal forms were determined to be P4₁32 (cubic) (Figure 18A), and P6₁22 (hexagonal) (Figure 18B), respectively. Space groups were determined and all diffraction data were measured at the University of California, San Diego Research Resource Laboratory at 4°C using graphite-monochromated CuK_α X-rays from either the Mark II Elliot GX-6 rotating anode diffractometer operating at 2 kilowatts or the Mark III Rigaku RU-200 rotating anode diffractometer (available from Rigaku USA, Danvers, MA) operating at 5 kilowatts, each equipped with two Xuong-Hamlin multiwire area detectors (available from San Diego Multiwire Systems, San Diego, CA).

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Preferably, area detector data collection is used. One facility offering equipment to support this data collection technique is The Resource Research Laboratory. This facility is a geographically designated, NIH supported facility to promote the use of X-ray crystallographic techniques. In speed, signal-to-noise ratio, and data precision, area detector data collection far surpasses standard diffractometer or film data collection. On the average, data collection is 50 times faster; consequently complete high resolution data sets can frequently be collected from a single crystal in one or two days. The space groups and lattice constraints of the crystal forms were determined to be the following: $P4_132$, $a=b=c=169.24$ Å; and $P6_122$, $a=b=80.3$ Å, $c=293.0$ Å. Calculations using an average reciprocal density of 2.7 Å³/D yield to the nearest unit 2 and 1 C-subunit monomers/asymmetric unit. The $P4_132$ form diffracts typically to 3.2 Å. Pictures of the cubic and hexagonal crystal forms can be seen in Figure 18

Because the $P6_122$ crystal form had diffraction better in extent and decay characteristics than the $P4_132$ form and because of the greater biochemical interest of a ternary complex, work concentrated on solving the hexagonal ternary complex crystal form. The lack of phase angles for a similar protein structure prohibited an initial structure solution for the C-subunit in the $P6_122$ form using molecular replacement techniques, so a structure solution using standard multiple isomorphous replacement (MIR) techniques was attempted. Both of these are techniques known to those of ordinary skill in this art. Briefly MIR involves introduction into the space group asymmetric unit of a relatively heavy reference atom that, after being located through difference Patterson analysis, enables the needed phase angles to be determined. The reference atom can be found with 6 -Å data, and with its location and generation of phase angles the fundamental crystallographic problem of a protein structure solution is solved and an electron density map can be calculated. Subsequent work on a protein structure focuses on incrementally improving the degree of detail visible in the electron density map through acquisition of higher resolution data and accompanying phase angles.

The procedure used to search for heavy-atom derivatives was to soak or co-crystallize C-subunit with heavy atoms based on the empirical success record or various heavy atom reagents and on known C-subunit chemical information, such as the availability of two free thiol groups and the obligatory use of a divalent cation in catalysis. Soaks in Au, Hg, and Pt compounds yielded precession picture diffraction changes but uninterpretable 6 Å difference Patterson maps. A 4.8 -Å data set from a $Na_2U_2O_7$ soak yielded an apparent Patterson solution through examination of isomorphous difference and $(1/\text{variance})$ -

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weighted anomalous difference Patterson maps, but the site quality was not high and attempts to reproduce or improve the soak failed. Isomorphous crystals grown with Co^{2+} or Cd^{2+} substituted for Mg^{2+} proved useless since neither metal could be located (location of Co^{2+} through Patterson analysis was improbable anyway due to its lightness), although with phases their positions could reveal metal site number and location. Co^{2+} and Cd^{2+} were chosen for co-crystallization based on their reported ability to support nucleotide binding to the C-subunit and support catalysis, although at a reduced rate.

The single most important modification in the crystallization protocol that led to the formation of crystals in a different space group was the careful selection of polyethylene glycol in combination with various low molecular-weight alcohols. Commercially available polyethylene glycol contains various contaminants that may cause problems in the achievement of stable and reproducible crystallization conditions. All commercially available polyethylene glycols (PEG) were examined with the aim of detecting the presence of ionic species.

The lowest level of ionic contaminants was detected in PEG manufactured by Dow Chemical. It is this PEG that was selected for further crystallization experiments. PEG from other sources appeared to be generally more contaminated and also exhibited large differences in contamination between batches. In our experiments, several molecular weights of PEG were used along with several low-molecular-weight alcohols.

The catalytic subunit crystallized in the hexagonal space group with the introduction of PKI(5-24) and MgATP , whereas in its apo form it crystallized in the cubic space group using otherwise identical crystallization conditions, indicates that the hexagonal crystal may arise as a result of a different conformational state of the enzyme.

Example 2

Mouse recombinant C-subunit Crystal Forms

One of the most promising directions for combining crystallographic methods with those of molecular biology is the development of highly effective vectors for expressing large amounts of protein for crystallization. Expression of protein in *E. coli* also provides a mechanism for eliminating posttranslational modifications which may hinder crystallization and in addition permits structure-function studies on mutant forms of the protein following the generation of mutant containing crystals.

The recombinant murine catalytic subunit, whose expression and purification was described by Slice et al., is devoid of myristic acid at the N terminus and differs by nine amino acids from the porcine heart catalytic subunit used in the earlier crystallizations. It

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has been shown, however, that N-terminal myristoylation is not necessary for C-subunit function. Additional differences between the porcine heart and recombinant mouse C α proteins include the presence of additional phosphorylation sites (Ser 10 and Ser 139) in the recombinant protein.

5 Crystals were prepared from a binary complex of the recombinant mouse C α -subunit with a bound, high-affinity ($K_i = 3$ nM) inhibitor peptide. The peptide (PKI95-24) derived from the N-terminal region of the naturally occurring thermostable protein kinase inhibitor protein (PKI), is the same peptide inhibitor used for the porcine heart ternary complex crystal. The steady state kinetics of the C-subunit purified from E.coli, are indistinguishable
10 from those of the mammalian C-subunit, although the E. coli protein is more labile to heat denaturation.

 The recombinant protein was crystallized using a small variation of the porcine heart ternary complex (hexagonal) conditions. A photograph of an exemplary crystal is provided in Figure 18C. First a ternary complex was prepared with MgATP and PKI(5-24). A
15 C:PKI(5-24) binary complex was obtained after small-angle neutron scattering experiments showed that for the recombinant mouse C-subunit, the PKI(5-24) peptide alone, without MgATP, was able to cause a significant decrease in the radius of gyration. The ternary complex crystal form diffracted to at least 2.7 Å on the Mark III and was of orthorhombic space P2 $_1$ 2 $_1$ 2 $_1$. A data collection strategy following the procedure of Xuong, et al., (Acta
20 Cryst. B41: 267, 1985) was developed. Equipment for use with this procedure is available from San Diego Multiwire Systems of San Diego, California. The procedure allowed an asymmetric unit of data to be collected in 3 ω -sweeps totaling about 140° (with appropriate choice of ϕ and χ settings and a crystal mounted with one of the axes parallel to the capillary axis). Data collection took about 16 h for a >90% complete 2.7-Å data set from one crystal
25 with R_{sym} on the intensity of 4-6%; in the same period the average reflection intensity decayed approximately 15%.

 Crystals were generally soaked or mounted in a stabilizing solution prepared as the crystallization drop, but with the addition of the initial reservoir MeOH percentage and the omission of C-subunit and PKI(5-24). It was discovered that Cd $^{2+}$ could be substituted for
30 Mg $^{+2}$ in crystal growth, as with the porcine heart ternary complex crystal. It was also discovered that elevating the MgCl $_2$ to ten times the starting mother liquor concentration, after crystal growth had stopped, altered the cell dimensions slightly (<1%) and resulted in a different pattern of heavy-atom binding.

Example 3

Recombinant Binary-Complex Structure Solution

The binary complex crystal was nearly isomorphous with the ternary complex crystal, differing by less than 1% along any axis and had the same space group with $a=73.62 \text{ \AA}$, $b=76.53 \text{ \AA}$, $c=80.14 \text{ \AA}$. The asymmetric unit contains one C:PKI(5-24) complex and has a calculated solvent content of 0.53. Mercury reagents were co-crystallized with the C:PKI(5-24) complex by exposing it to 1-mM reagent for six hours, followed by dialysis to remove excess Hg reagent. Native and co-crystallized 4-(hydroxymercuri)benzoic acid (PHMB) derivative diffraction data were measured on the Mark III diffractometer. Data from two native crystals were merged to provide a more complete data set for the initial phase computation, phase extension work, and initial refinement cycles, but later refinement used data from only a single crystal that yielded better quality data. Data from two PHMB co-crystals, one with measured Bijvoet mates, were kept separate and used to compute initial phases, which were improved at constant 3.5- \AA resolution using the solvent flattening approach of Wang (Methods Enzymol. 115:90, 1985), with a slightly conservative solvent content of 0.50. Additional phases from inversion of the solvent-flattened map were added incrementally using 6 additional envelopes to a final resolution of 2.7 \AA . Starting with minimap α -carbon coordinates, the program TOM/FRODO (available from Christian Cambillau, University of Marseille, Marseilles, France) was used with the resulting map to model in the sequence of PKI(5-24) and 257 of the 350 residues in the C-subunit. This partial model was refined using X-PLOR (available from Axel T. Brunger, Yale University, New Haven CT), and calculated structure factors were combined with Wang structure factors in a resolution-dependent manner using equally weighted ABCD coefficients to yield improved maps. ABCD coefficients are described by Hendrickson et al. (Acta Cryst. B26: 136, 1970). Structure solution statistics are summarized in 2, and a sample of electron density of the structure determination is shown in Figure 3, described in detail below.

A number of crystal forms of the catalytic subunit of cAPK have been obtained thus far. All of the crystal forms of the different complexes of the catalytic subunit, with the exception of the monoclinic crystal of the apoenzyme, were obtained under identical crystallization conditions and these are described above. The crystals in different space groups therefore very likely result from conformational states of the enzyme. Crystals of both the binary and ternary complexes with PKI(5-24) exhibited better diffraction characteristics than crystals of the apoenzyme.

Our results also indicate that the ternary complex of the murine catalytic subunit expressed in *E. coli* produced a crystal of better quality than did the ternary complex of the

catalytic subunit purified from porcine heart. It is difficult to conclude whether this was due to the absence of myristic acid, the amino acid differences between the two forms, microheterogeneity in the mammalian enzyme, or a combination of these factors. It may suggest, however, that another way to improve the quality of crystals is to mutate the protein and to cocrystallize mutants if crystallization of the wild type fails.

Three factors are important for reproducible crystallization. First, the salt of the eluting buffer of the last column must be chosen carefully. Second, the purity of the protein must be verified with isoelectric focusing gels. The protein must not contain typical additives, such as glycerol and should not be frozen prior to crystallization. Third, all reagents used for crystallization must be of the highest degree of purity. If all of these conditions are met, it is possible to obtain, in identical crystallizations, three different crystal forms representing two different conformational states of the enzyme. Some of those crystals, such as those of the ternary complex with PKI(5-24), are of much better quality than the other crystals.

The current model consists of C-subunit residues 15-350 and 1-20 of PKI(5-24) and has been partially refined using X-PLOR to an R-factor of 0.195 with r.m.s.(root mean square) bond length deviation from ideality of 0.024 Å. The location of the MgATP-binding site was determined by difference Fourier synthesis with the nearly isomorphous ternary complex crystal, which showed clear density for the adenine, ribose, and α -PO₄ for the low-[Mg²⁺] ternary complex crystal. The high-[Mg²⁺] difference density showed additional features that could contain the β - and γ -PO₄ as well as metal ion(s), but an unambiguous assignment of atoms to this density could not be made.

Diffraction data is summarized in Table 1. Definitions for Table 1 are as follows: f_h , calculated heavy-atom structure factor amplitude; F_p , measured native structure factor amplitude; F_{ph} , measured derivative structure factor amplitude; ΔF_{anom} , calculated Bijvoet difference; E_{iso} , r.m.s. isomorphous lack-of-closure, E_{anom} , r.m.s. anomalous lack-of-closure; $R_c = \sum |F_{ph} \pm F_p| - f_h / \sum |F_{ph} - F_p|$.

All diffraction data were measured at 4°C using graphite-monochromated CuK_α X-rays from the Mark III Rigaku RU-200 rotating anode diffractometer equipped with two Xuong-Hamlin multiwire area detectors. Paired runs starting from settings (ω, ϕ, χ) and ($\omega, \phi + 180, -\chi$) were used to collect Bijvoet mates (inverse beam method). Data reduction and derivative-to-native scaling were done using the UCSD area detector data processing programs (available from San Diego Multiwire Systems). $R_{sym} = \sum |I_{obs} - I_{avg}| / \sum I_{avg}$ and is shown for merged Friedel pairs.

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Native-1 was used for native. Hg positions of the PHMB (4-(hydroxymercuri)benzoic acid) co-crystal derivative were found from a difference Patterson synthesis. The heavy-atom sites in relation to the model suggest heavy-atom binding at Cys 343 (major site) and Met 58 (minor site). Positional and relative occupancy refinement of two common sites (relative occupancies 2.66, 1.87 for PHMB-1), and calculation of native phases and corresponding ABCD coefficients, were done using the program HEAVY (available from the Protein Data Bank, Brookhaven National Laboratory, Upton, NY). Solvent flattening used the Wang program package (Bi-Cheng Wang, University of Pittsburgh, Pittsburgh, PA) on imported initial ABCD coefficients and phases to 3.5 Å. Molecular envelopes were calculated with solvent content 0.50 rather than 0.53 calculated for the cell. After 3 envelopes at 3.5 Å, the resolution was extended incrementally in 6 shells to a final resolution of 2.7 Å. After convergence at 3.5 Å, the mean phase change/reflection was 36.6° and the mean figure of merit was 0.84; the map inversion R-factor was 0.181. Phase extension added 6786 phases from 5914 in the 3.5-Å starting set; 261 unobserved reflections were estimated by map inversion in the 2.7-Å set.

X-PLOR Version 2.1 was used exclusively following recommended protocols provided in the accompanying manual. Simulated annealing was performed according to a slow-cooling protocol (Brunger et al. Science 235:458-460, 1987) between either 3000K or 4000K and 300K, followed by 120 cycles of conjugate-gradient minimization. Refinement began with the partial model of Stage A to improve the coordinates for phase combination. Combined maps were calculated using the Hendrickson-Lattman scheme. Wang phases were used to 6 Å combined ones between 6 Å and 3.5 Å or 3.0 Å, and calculated phases between 3.5 or 3.0 Å and 2.7 Å. The corresponding weighted amplitudes were $m_{\text{Wang}}F_o$, $m_{\text{comb}}(2F_o-F_c)$, and $m_{\text{sim}}(2F_o-F_c)$. The model was completed by iterative refinement and building in areas not included in refinement of partial model. Refinement and R-factor ($= \sum |F_o-F_c| / \sum F_o$) calculations used $F/\sigma > 2$ reflections (12024 Native-1 reflections; 10194 Native-2 reflections beginning with Stage B). The current R-factor of 0.195 is for 2939 atoms (no solvent atoms) with individual B-factors (r.m.s. $B = 17.6 \text{ \AA}^2$). R.M.S. bond length and angle deviation from ideality are 0.024 Å and 4.3°.

In summary, the crystals were grown as described above using a 5-10% molar excess of PKI(5-24) and were determined to be of space group $P2_12_12_1$ with $a = 73.62 \text{ \AA}$, $b = 76.52 \text{ \AA}$, $c = 80.14 \text{ \AA}$. The asymmetric unit contained one C:PKI(5-24) complex and had a calculated solvent content of 0.53. Native and co-crystallized 4-(hydroxymercuri)benzoic acid (PHMB) derivative diffraction data were measured using Xuong-Hamlin area detectors.

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Data from two native crystals were merged to provide a more complete data set for the initial phase computation, phase extension work, and initial refinement cycles, but later refinement used data from only a single crystal of better quality. Data from two PHMB co-crystals, one with measured Bijvoet mates, were kept separate and used to computer initial phases, which were improved at constant 3.5-Å resolution using the solvent flattening approach of Wang with a slightly conservative solvent content of 0.50. Additional phases from inversion of the solvent-flattened map were added incrementally using 6 additional envelopes to a final resolution of 2.7 Å. Starting with a minima alpha-carbon coordinates, the program TOM/FRODO was used with the resulting map to model in the sequence of PKI(5-24) and 257 of the 350 residues in the C-subunit. This partial model was refined using X-PLOR, and calculated structure factors were combined with Wang structure factors in a resolution-dependent manner, using equally weighted ABCD coefficients to yield improved maps. Details on the combination can be found in Allured et al. Proc. Natl. Acad. Sci. USA 83:1320, 1986 and Remington et al. J. Mol. Biol. 158:111, 1982. An example of the electron density of the structure determination is shown in Figure 3.

Figure 3 is a stereo view of the electron density for the structure determination. Portions of the latest refined model of 3 β-strands are shown (top to bottom from left): 112-106, 114-121, 75-69. Figure 3A provides the 1.5-σ experimental density calculated to 2.7Å using phases after Wang improvement and extensions. Figure 3B provides the 1.5-σ(2F_o-F_c) density calculated with 10 to 2.7-Å refined model phases. The current model consists of C-subunit residues 15-350 and 1-20 of PKI(5-24) and has been partially refined using X-PLOR to an R-factor of 0.195 with r.m.s. bond length deviation from ideality of 0.024Å. The structure of the catalytic subunit and effector molecule are described below.

Example 4

Structural Analysis of the Catalytic Subunit

A stereo view of the backbone structure of the C-subunit with the bound peptide is shown in Figure 4. Residues 15-350 of the C-subunit and the twenty residues of PKI(5-24), in bold print, of the partially refined model are shown. The overall dimensions of the monomer (65Å x 45Å x 45Å) indicate a slightly elongated molecule. Earlier hydrodynamic measurements showing a Stokes radius of 26.1Å, a frictional coefficient ration (f/f₀) of 1.19, and a radius of gyration of 20Å are consistent with this structure. The most striking feature of the overall molecular architecture is its bilobal shape with a deep cleft between the two lobes. The core of the small lobe is associated primarily with the amino-terminus, while the core of the large lobe corresponds to the C-terminal region of the protein. The cleft

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between the lobes is filled by a portion of the bound inhibitor peptide in the binary complex. A difference Fourier map of the ternary complex containing both peptide and MgATP places MgATP at the base of that cleft (Figure 5). The 3.5- σ positive density contours for the ($F_{\text{ternary}} - F_{\text{binary}}$) difference Fourier were calculated using refined model phases in 10 to 2.7-Å range and are shown superimposed on the partially refined backbone model. Figure 5A illustrates the general localization of MgATP while Figure 5B is a close-up of difference density enclosing an oriented model of AMP.

The cleft is clearly the site of catalysis, and the peptide-induced conformational changes, observed by both SANS and circular dichroism, may be associated with a closing of this cleft. SANS established that in the absence of inhibitor and MgATP the enzyme adopted a more expanded conformation than that adopted by the enzyme in the binary complex of the enzyme and the peptide inhibitor, or the ternary complex of the enzyme, inhibitor and MgATP. This technique was also used to show that binding of the inhibitor to the enzyme did not require MgATP. Neutron scattering, in particular, established that the apo form of the enzyme adopts a more expanded conformation than the ternary complex containing MgATP and PKI(5-24). Furthermore, PKI(5-24) alone, but not MgATP, was sufficient to induce this conformational change. SANS and CD are techniques known to those of ordinary skill in this art. Accordingly, no further descriptions of these techniques are necessary.

Most of the predictions of secondary structure made prior to this crystallographic study of the C-subunit are quite inaccurate and do not correlate well with the actual structure that is provided herein. The prediction of the secondary structure by Benner et al., *Adv. Enzyme Regulat.* 31:121, 1991, is somewhat more accurate. It is based on chemical information and homologies within the protein kinase family and is accurate within the small lobe. However, detailed and accurate information on the structures of the protein kinase family has not been available until the discoveries presented herein.

The amino-terminus of the C-subunit begins with an amphipathic α -helix that lies primarily along the surface of the larger lobe. This N-terminal region differs in the recombinant and mammalian enzymes, since the recombinant protein lacks a myristoyl group at the N-terminal glycine. In the crystal structure, the first 14 amino acids are not visible. However, the surface of the enzyme in this N-terminal region is hydrophobic, suggesting a possible site for the N-terminal myristoyl moiety of the mammalian enzyme. The myristoyl group stabilizes the C-subunit but does not promote association with membranes.

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The smaller lobe, consisting of residues 40 through 125, is associated primarily with the binding of the nucleotide and is characterized by a dominance of β structure. Five antiparallel β -strands comprise the core of this domain. The only helical element in the small lobe is inserted between β -strands 3 and 4 and lies on one side of the plane of the β -sheet. It consists of two parts: a two turn helix B, followed by a sharp break and a five turn helix, helix C. Based on a difference Fourier map (Fig. 5) with a ternary complex of the recombinant C-subunit containing MgATP and PKI(5-24), and supported by chemical evidence discussed below, it is clear that this small lobe is the primary site for interaction with MgATP. As seen in Figure 5, the density based on the difference map is consistent with the adenine moiety of the nucleotide oriented towards the base of the cleft beneath the β -sheet, with the phosphates facing outwards, towards the edge of the cleft. This structure is distinct from the Rossmann fold that is characteristic of many nucleotide binding proteins.

The larger lobe, in contrast, is remarkable for its predominance of helical structure. Seven helices are found in this C-terminal domain. A particularly unusual feature are the antiparallel hydrophobic helices, helix E (residues 140 through 159) and especially helix F (residues 218 through 233), that extend right through the core of this domain. The only region of β -structure in this lobe is located on the surface of the cleft at the interface between the two lobes where four antiparallel β -strands form a sheet. Most of the regions important for peptide recognition, as well as some conserved residues likely to be involved in catalysis, are located within this larger lobe.

The C-terminal 70 amino acids, residues 281 through 350, extend over a large portion of the surface of the enzyme from the bottom of the large lobe to the top of the small lobe. The part of this extended chain that passes through the region linking the two lobes appears to participate in recognition of both the peptide and the nucleotide, even though these amino acids are outside the conserved catalytic core. The other extended chain connecting the two lobes of the enzyme, residues 120 through 127, likewise, passes through this linker region between the small and large lobe and also participates in peptide recognition. Hence, this linking region consisting of both chains may contribute in part to the observed peptide-induced conformational changes described earlier. An overall two dimensional topology diagram for the C-subunit of cAPK is presented in Figure 6. Residues corresponding to the secondary structure elements are as follows: β -strands - 1:43-48, 2:57-63, 3:67-75, 4:106-111, 5:115-120, 6:161-164, 7:171-175, 8:178-183, 9:188-191; α -helices - A:15-31, B:76-82, C:84-97, D:-128-135, E:140-159, F:218-233, G:244-252, H:263-272, I:288-293, J:301-307.

CORRELATION OF STRUCTURAL DATA WITH CHEMICAL DATA

As discussed above, chemical data can be used to confirm the correct interpretation of the electron density map. Chemical analysis has been used as a way to obtain structural data in the absence of X-ray crystallography. Since the protein kinase family is an enzymatic group of major import, a significant body of chemical data is available. While this data cannot be used to predict a three-dimensional structure for effector modelling, it does provide a body of data that can be used to confirm and ensure the consistency of the three-dimensional structure. Thus, once a crystal structure is obtained for a model enzyme and its effector, the chemical data present in the literature can be used to examine the consistency of the model before proceeding to the design step. The three dimensional structure of the enzyme-effector complex should provide a solid explanation for the earlier chemical data. Information provided from chemical data together with structural data is used to obtain both the template and the "lock" derived therefrom.

For example, evidence for localizing the nucleotide binding site near the amino-terminus first came from affinity labeling with an analogue of MgATP, fluorosulfonyl benzoyl adenosine (FSBA). Labeling with a hydrophobic carbodiimide, DCCD, identified two carboxyl groups near the MgATP binding site, Asp184 and Glu91, and, furthermore, established that Asp184 could be readily cross-linked to Lys72 in the apoenzyme. The structure of the binary complex without bound MgATP (Fig. 7) confirms that all three residues are localized in close proximity to one another, while the difference Fourier map with the ternary complex places these residues close to the γ -phosphate region of MgATP (see Fig. 5). Figure 7 provides stereo views of selected conserved areas. $1.5\text{-}\sigma$ ($2F_o - F_c$) electron density (10 to 2.7\AA) is shown superimposed on the latest refined coordinates. In Figure 7A the sidechains of the invariant Lys72, Glu91, and Asp184 are shown in proximity to each other. Figure 7B shows the catalytic loop, Arg-Asp-Leu-Lys-Pro-Glu-Asn (165-171), together with part of PKI(5-24). Arg 20 of PKI(5-24) is labeled as 365. Lys72 is on β -strand 3, and Glu91 lies along the edge of the C-helix that faces the cleft. Asp184 is located on the loop connecting β -strands 8 and 9, and this loop also lines the cleft. All three residues are invariant in every protein kinase. Therefore these residues can be used as anchors for modeling the three dimensional structure of other protein kinases.

The MgATP binding site was defined more globally by differential labeling with acetic anhydride. By describing the reactivity of each lysine side-chain in the presence and absence of substrates, it was shown by Buechler et al., *Biochemistry* 28:3018-3024 (1989), that the specific protection afforded by MgATP was localized exclusively to residues in the

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small lobe. In addition, to Lys72, MgATP protected Lys76 and Lys47 against modification by acetic anhydride. These protected lysines also flank the conserved glycine-rich loop that lies between β -strands 1 and 2. Based on the difference Fourier shown in Figure 5, this loop is close to the phosphates of MgATP.

5 Chemical studies using an affinity analogue have shown that Cys 199 is important for peptide binding. Modification of Cys 199 leads to loss of activity, and MgATP protects against inactivation. In contrast, Cys 343 can be covalently modified with no concomitant loss of activity. The structural analysis reported here indicates that Cys199 is on the surface of the cleft that interacts with the C-terminus of the inhibitor peptide, and Cys343 is on the
10 surface of the small lobe. This distance measured between the two α -carbons of Cys199 and Cys343 in the crystal structure is 24Å. Thus, some of the chemical data is confirmed by the crystal structure.

CONSERVED REGIONS AND THEIR FUNCTIONS

15 The fact that all known protein kinases share a conserved catalytic core that is homologous to the C-subunit provides information that independently highlights important regions. This conserved catalytic core begins with the β -1 strand in the small lobe and extends through Arg280 in the large lobe (Hanks et al., *supra*) The two lobes comprising this conserved catalytic core can be seen clearly in Figure 8. Figure 8A is a space-filling model of the catalytic core (residues 40-280) shared by all protein kinases. The small lobe
20 corresponding to the nucleotide binding fold 1 (residues (40-126); the larger lobe 2 (residues 127-280). In this model the bound peptide is not shown. Figure 8B is a diagram of the conserved catalytic core using the RIBBON program of the PAP package (J. P. Priestle, J. Appl. Cryst. 21:572, 1988 and available from the Molecular Simulation Laboratory at the University of Minnesota, Minneapolis, MN). Regions of the linear sequence noted by
25 Hanks et al., *supra*, are indicated. The protein kinase having the largest insert at each position is designated using the following notation to define each insert: Gene/Protein Name: NH₂-terminal C-subunit residue no. (insert length) COOH-terminal C-subunit residue no. The inserts are CDC7:64(14)65, KIN1:83(26)84, PKC- γ :98(6)99, c-mos:113(5)114, PDGFR:137(99)138, CDC7:196(82)197, ran⁺1:210(23)211,
30 HSVK:240(11)241, CDC7:260(93)261, 7less:178(7)179. Figure 8C is identical to Figure 8A, but includes PKI(5-24) 3. Within this conserved core are nine invariant amino acids, as well as several highly conserved residues. Most of these conserved residues contribute directly to either MgATP binding or catalysis. Others, such as Arg280 and Asp208, exist as ion-pairs

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and link two segments of the polypeptide chain that are widely separated in the linear sequence.

In addition, to providing information on conserved residues, sequence comparisons among protein kinases also identify inserts, sometimes quite sizable, that lie within the catalytic core. These inserts were noted by Hanks et al., *supra*, but their conformation in the overall structure of the catalytic subunit and their relationship to other regions of the catalytic core is described for the first time here. The locations of these inserts are indicated in Figure 8. All inserts invariably are located at loops on the surface of the protein and can be accommodated within the tertiary structure.

The structures of several important regions of the catalytic subunit are described below. Two highly conserved loops, as well as a triad of invariant charged residues, appear to be particularly important for nucleotide binding and catalysis. However the regions important for recognition of the peptide substrate are quite variable and were not available until the crystallized structure was analyzed. Predictions based on these variabilities are heretofore undescribed.

GLYCINE-RICH LOOP

The glycine-rich segment, Gly⁵⁰-Thr-Gly⁵²-Ser-Phe-Gly⁵⁵, was identified originally as part of the MgATP binding site based on its proximity to Lys72 and on differential labeling with acetic anhydride, since all of the lysines flanking this region, Lys47, Lys72, and Lys76, are protected in the presence of MgATP. The specific structural explanation obtained from crystallographic data for the protection of Lys47 is due to ionic pairing with the side chain of Glu333 while Lys76 ion-pairs with Glu346. Thus, conformation changes that occur around the glycine-rich loop as a consequence of MgATP and peptide binding are understood from the structural data in combination with known chemical data.

A glycine-rich motif is associated with many nucleotide binding sites, and this region has been the subject of much speculation and model building. The Rossmann fold, found in many nucleotide binding sites, contains a sheet of mostly parallel β -strands containing a glycine-rich loop. A similar motif containing a glycine-rich loop is found in other proteins such as adenylate kinase and p21 ras. The protein kinase fold found in the C-subunit and conserved in over one hundred protein kinases, does not conform to either of these motifs; it forms a unique nucleotide binding site. The uniqueness of this site is summarized as follows: (1) The glycine-rich segment lies at a sharp turn that joins two antiparallel strands at the beginning of the β -sheet. (2) The phosphate binding site is not dominated by a helix whose dipole points towards the phosphate. (3) The nucleotide does not lie along the edge

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of the β -sheet. (4) An invariant Lys does not immediately follow this loop. Instead, the invariant Lys in the protein kinases, Lys72, is located in the β -3 strand and is a part of the stable scaffold of the structure. The single conserved element in each of these motifs is the glycine-rich loop whose apparent function is to serve as a phosphate anchor so that the γ - PO_4 is poised for transfer.

CATALYTIC LOOP

Another highly conserved loop in the C-subunit extends from Arg165 through Asn171 and can be termed the catalytic loop (Fig. 7B). This catalytic loop, Arg-Asp-Leu-Lys-Pro-Glu-Asn, contains 2 invariant residues, Asp166 and Asn171, and 2 highly conserved residues, Arg165 and Leu167. While the purpose of the glycine loop is to anchor the phosphate moiety and, in particular, to help position the γ - PO_4 so that it is poised for transfer, it is the catalytic loop that appears to be the central hub that communicates to many different parts of the molecule. This loop not only directs the catalytic event, but also guides the peptide into its proper orientation so that catalysis can occur. The loop itself and, in particular, the residues that are important for catalysis are highly conserved, while the parts of the loop that direct the peptide binding are not.

Asp166 is one of 4 invariant carboxyl groups in the protein kinase family. It is the only one that is oriented towards the Ala side chain at the pseudo-phosphorylation site in the bound inhibitor peptide. Asp166 most likely functions as a catalytic base. Catalysis is thought to occur as a direct in-line transfer without an enzyme bound phospho-intermediate.

INTERDOMAIN CONTACTS:

The triad composed of the side chains of Lys72, Asp184, and Glu91, shown in Figure 7A, is conserved in every protein kinase and is close to the γ - PO_4 of MgATP. Asp184 was a candidate for the catalytic base; however, the structure indicates that a more plausible role is participation in the chelation of Mg^{2+} in the MgATP complex. The side chain of Asp184 also comes within 4-5Å of the side chain of Asn171. This cluster, Asp184, Asn171, and Asp166, thus forms a second triad of invariant amino acids. Asp184, being a component of both triads, has the potential to shuttle between the two conserved loops, the glycine-rich loop in the small lobe and the catalytic loop in the larger lobe. Hence, if the position of Asp184 changes following the binding of MgATP, as it probably will given its location in the structure relative to the MgATP binding site, the consequences will have a direct impact on both conserved loops. If, for example, Asp184 participates in the chelation of Mg^{+2} , its negative charge would be sequestered from the catalytic loop, thus allowing the other

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residues to rearrange in order to maximize the nucleophilicity of the serine hydroxyl moiety that is poised to receive the phosphate from ATP.

This is the first protein kinase structure to be reported. The protein kinases represent a large family of over 100 enzymes that includes growth factor receptors as well as many oncoproteins. In spite of the tremendous diversity of these enzymes, all share a conserved catalytic core that retains the same essential features of secondary and tertiary structure and the same general mechanism of catalysis. The essential hallmarks of this conserved core include: (1) two lobes with a cleft between that is occupied by the substrates, (2) a unique nucleotide binding fold dominated by β -structure, (3) a largely helical domain associated with peptide binding and catalysis, (4) two β -sheets converging at the active site near the domain interface, and (5) two conserved loops, one in each lobe, that converge at the active site. In marked contrast to these conserved features shared by all protein kinases, recognition of the peptide by the catalytic subunit involves non-conserved amino acids, and the peptide binding sites extend over diverse and widely separated regions on the surface of the enzyme. The detailed structure of the bound inhibitor peptide and its specific interactions with the catalytic subunit are described below.

Affector binding site data may incorporate information derived from several experimental avenues. In addition, to crystallographic studies, substrate analogues provide insights into the specific features of a given substrate that are important for recognition. Chemical approaches such as affinity labeling and group specific labeling can identify regions and specific residues that are in close proximity to substrates. Crystallographic studies can include a structural analysis of the apoenzyme, i.e. the structure of the enzyme without other associated molecules. However, more importantly, crystallographic studies of co-crystals of the enzyme with bound substrates or effectors are provided, so that the precise features of the active site can be defined.

Thus, in the model system of the present invention, crystals of the cAMP-dependent protein kinase C-subunit/PKI(5-24) were obtained and structural data derived therefrom. This structure of the catalytic subunit is presented as example 4. The inhibitor peptide PKI(5-24) is a fragment of the heat stable protein kinase inhibitor. Additional information about this inhibitor can be found in a publication by H.-C. Cheng et al. (Biochem J. 231:655-661, 1986). This peptide includes the consensus features common to all peptide substrates and inhibitors of cAMP-dependent protein kinase. In addition, it contains other features that convey unique high affinity binding. The crystals of complexed enzyme and inhibitor

provide insight into the guidelines necessary for designing effector molecules for other protein kinases.

5 The general requirements for peptide recognition by the catalytic subunit were characterized originally using synthetic peptides. These studies found a consensus sequence that includes two basic residues, typically arginines, followed by one intervening small residue preceding the site of phosphotransfer. The phosphate acceptor site, either Ser or Thr, is followed immediately by a large hydrophobic residue. The typical peptide substrate used for this kinase, based on the phosphorylation site in pyruvate kinase, is shown in Table 2. Replacement of the Ser with an Ala generates an inhibitor peptide with poor affinity. 10 In contrast is the inhibitor peptide, PKI(5-24), derived from the thermostable protein kinase inhibitor (PKI). This peptide, seen in 2, encompasses the consensus sequences described above but also contains additional unique features that convey high affinity binding. The particular features that contribute to high affinity binding were defined using peptide analogues, and the most important ones are also indicated in 2. This peptide, PKI(5-24), 15 was co-crystallized with the catalytic subunit, and the structure of that peptide as well as its interaction with the protein are discussed below.

A schematic of substrate and inhibitor peptides of cAMP-dependent protein kinases are provided in Table 2. The nomenclature used for the peptides designates the phosphorylation site or pseudophosphorylation site residue as P. In the case of substrates, 20 P will be Ser or Thr; in the case of PKI(5-24), P is Ala. The residues flanking this site are designated as P+1, P-1, etc. as indicated. This nomenclature provides a common frame of reference for all peptide substrates and inhibitors and can be invoked readily for every protein kinase.

The Ser peptide is based on the *in vivo* phosphorylation site in pyruvate kinase. 25 Residues shown to be important for peptide recognition are shaded and were identified using synthetic peptide analogues of the Ser peptide and of PKI. Procedures for determining which residues are important for peptide recognition using peptide analogues can be found in articles by Glass et al. and Kemp et al. (J. Biol. Chem. 262:8802-8810, 1989 and J. Biol. Chem. 252:4888-4894, 1977 respectively.)

30

Example 5

Conformation Determination of the Bound Inhibitor

The conformation of bound PKI(5-24) is shown in Figure 9. Backbone C and N atoms are shown in bold. Residues particularly important for binding are labelled according to the nomenclature of 2. The amino-terminus extending from the P-16 Thr through the P-8

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Ala forms an amphipathic α -helix. This helix is followed by a turn flanked by glycines at the P-7 and P-4 positions. The glycines may be important for accommodating the turn or for providing flexibility to facilitate binding of the Arg that follows each Gly. The remainder of the peptide is in an extended conformation, and the density corresponding to the region at the C-terminus, the P+2 Asp and the P+3 His, is not well defined.

The catalytic subunit itself consists of 2 lobes - a smaller lobe, associated primarily with MgATP binding, and a larger lobe. Nearly all of the features necessary for peptide recognition are found within the larger lobe, although the specific residues involved are widely dispersed both in the linear sequence and on the surface of the enzyme. The extended portion of the peptide that includes the consensus region for recognition of all substrates and inhibitors lies along the surface of the cleft corresponding to the larger lobe. The helical segment of the peptide is amphipathic, and its hydrophobic side lies in a hydrophobic pocket on the surface of the large lobe. The specific interactions of the peptide with the protein can be described by (i) the interactions that account for the unique highly affinity binding of PKI and (ii) by the features of the protein that are important for recognizing the consensus sequence common to both the inhibitors and substrate.

HIGH AFFINITY BINDING SITE

Based on the crystal structure, the high affinity binding attributed to the N-terminus of PKI(5-24) is dominated by hydrophobic interactions involving primarily the phenylalanine side chain at the P-11 position. Glass et al. showed that a replacement of this Phe with an Ala caused a 100-fold increase in K_i while replacement with 1'-naphthylalanine, a residue that is considerably larger and more hydrophobic than Phe, actually decreased the K_i by 4-fold. Figure 10 illustrates the high affinity binding site interactions. A hydrophobic pocket on the surface of the C-subunit nicely complements the hydrophobic face of the helix in the inhibitor peptide. This hydrophobic pocket is lined by residues 235 through 239, Tyr-Pro-Pro-Phe-Phe, with the phenyl ring in the inhibitor peptide sandwiched between the side chains of Tyr235 and Phe239. In Figure 10A the P-11 Phe ring (numbered 356) is shown in relation to three nearby hydrophobic residues of the C-subunit: Tyr 235, Pro 236, and Phe 239. Figure 10B illustrates the interactions of the P-11 Phe and P-6 Arg sidechains with the C-subunit. Distances between charged-residue sidechain atoms $<3.5\text{\AA}$ apart are indicated by thin connecting lines. Based on the structure, the Tyr at the P-14 position is not essential for this hydrophobic interaction. In addition, to the hydrophobic interactions associated with the helix, the orientation of the high affinity binding region PKI(5-24) is fixed by the ionic

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contacts involving the P-6 Arg. Two nitrogens in the guanidine side chain of this Arg undergo ion-pairing with the two oxygens of the carboxyl group of Glu203.

CONSENSUS RECOGNITION SITE

Figure 11 summarizes the interactions that contribute to recognition of the common consensus region of the inhibitor peptide. The interactions of the P-3 and P-2 Arg residues and the P+1 Ile residue with C-subunit residues are shown. Lines are drawn between charged-residue sidechain atoms <3.5Å apart. The P+1 Ile sidechain projects into the hydrophobic area formed by Leu 198, Pro 202, and Leu 205. Electrostatic interactions dominate the portion of the peptide proximal to the site of phosphotransfer, while hydrophobic interactions dominate the C-terminal region distal to the phosphotransfer site.

Two important requirements for peptide recognition by cAPK are basic residues at the P-3 and P-2 positions. Others have shown that replacing either Arg in the Ser peptide substrate leads to a 16-400-fold increase in K_m , even when the Arg is replaced with a Lys. The environment flanking the P-3 and P-2 arginines explains these results since each Arg interacts with more than one carboxyl side chain.

Table 3 provides a listing of the amino acid residues present at the various points of contact between PKI(5-24) and two protein kinases, cAPK and casein kinase II (CKII). It can be seen from Figure 11 and Table 3 that in the C subunit of cAPK, that those residues lining the p+1 site are very hydrophobic and provide a pocket for the hydrophobic p+1 residues. In CKII, the residues lining this pocket are all basic or positively charged. This basic pocket compliments an acidic residue at the p+1 position and this is consistent with the known specificity of CKII, i.e. CKII prefers acidic groups at the p+1 position.

Figure 12 provides information on the consensus recognition site binding interactions. The electron density corresponding to the anionic P-3 site is shown in Figure 12A. Residue numbers 361, 364, 365, and 368 correspond respectively to PKI(5-24) P-6, P-3, P-2, and P+1 residues. The electron density of the P-3 Arg sidechain tip is shown in proximity to Thr 51 carbonyl in the glycine-rich loop, and Glu 127 and Glu 331 sidechain carboxylates of the domain-linking region. In Figure 12B the 1- σ ($2F_o - F_c$) electron density of the P-2 Arg sidechain is shown in proximity to sidechain carboxylates of Glu 170 of the catalytic loop and Glu 230; the P-6 Arg sidechain is shown near sidechain carboxylate of Glu 203. In Figure 12C the 1.5- σ ($2F_o - F_c$) electron density of the P+1 Ile sidechain is shown projecting into a hydrophobic pocket comprised of residues Leu198, Pro202, and Leu205. The side chain of this P-3 Arg interacts with Glu127. The carboxyl side chain of Glu331 also is approximately 3Å from the guanidinium nitrogens. The tip of Asp329 is approximately

5Å away. Thus, the position of the guanidinium moiety is fixed. In addition, the side chain of the P-3 Arg comes close to the backbone carbonyl of Thr51 in the glycine-rich loop and to the hydroxyls of the ribose ring. The side chain of Glu333 lies close to Lys47 in β -strand 1, and the side chain Glu334 is approximately 3Å from the hydroxyl group of Thr48.

5 The P-2 site is also very anionic, and this Arg, likewise, interacts with more than one carboxyl group. As indicated in Figures 11 and 12B, the ϵ -nitrogen forms an ion-pair with Glu170, while one of the terminal nitrogens interacts with Glu230. Glu 203 also comes close to this guanidinium side chain; however, its interaction with the P-6 Arg is dominant. In the absence of an Arg at the P-6 position, Glu230 may ion-pair with the P-2 Arg. Unlike the
10 P-3 recognition site, all of the carboxyl groups at the P-2 site are an integral part of the large lobe.

DISTAL HYDROPHOBIC SITE (P+1)

 Peptide analogue studies of others predicted a hydrophobic requirement at the P+1 position since replacement of the Leu with Gly in PKI(5-24) caused a 150-fold increase in
15 K_m . The reasons for this requirement are now clear from the structure (Figs. 11 and 12C). Leu198, Pro202, and Leu205 form a hydrophobic groove that surrounds the Ile side chain. This hydrophobic region that constitutes the P+1 site lies at the edge of the cleft and is likely important for proper orientation of the actual site of phosphotransfer at the P position. In the binary complex this region begins to align in an antiparallel β -like
20 configuration with the carbonyl of the P+1 Ile coming less than 4Å from the backbone amide of Gly200 and the carbonyl of Gly200 coming within approximately 3Å of the backbone amide of this Ile at the P+1 position (Fig. 13B). Substitution of a Pro for Leu at the P+1 position in the Ser peptide (2) yields an extremely poor substrate. Nevertheless, a depsipeptide analogue of this peptide lacking an amide proton at this P+1 site is still a
25 good substrate for the catalytic subunit.

 Figure 13 illustrates the catalytic site area. Residue numbers 364 and 367 correspond to the P-3 Arg and the P Ala. 1.5- σ ($2F_o - F_c$) electron density is shown in all cases. Figure 13A provides the site of catalysis together with the possible catalytic base sidechain of Asp 166 near the β -C of the P Ala. Thr 51 of the glycine-rich loop is shown
30 near the P-3 Arg sidechain, and hydrophobic sidechains of residues Phe 54 (at the loop apex) and Phe 187 are shown near the site of phosphotransfer. The addition of a hydroxyl group would place the side chain of the residue at the P position close enough for a direct transfer of the γ -phosphate for MgATP. The side chain of the P-1 Asn also interacts with the glycine-rich loop as shown in Fig. 13A. Figure 13B diagrams the consensus recognition

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site residues Arg-Arg-Asn-Ala-Ile together with the glycine-rich phosphate anchor loop to the left and residues 198-202 to the right. The term "residue" is here used interchangeably with amino acid. The carbonyl of Gly 200 can be seen pointing to the amide N of the P+1 Ile. Figure 13C shows the phosphate of Thr 197 buried among sidechains of cationic and H-bonding residues (His 87, Arg 165, Lys 189, Thr 195). Cys 199 is also shown nearby.

Thr197, one of the two stable phosphorylation sites in this enzyme, also flanks the P+1 site. Multiple electrostatic interactions, seen in Figure 13C, hold this PO₄ in place and account for its resistance to removal by phosphatases. Fixing this phosphate moiety contributes conformational stability, not only to Thr197 but also to the adjacent hydrophobic residues important for recognition at the P+1 site and for the proper orientation of the site of phosphotransfer. Based on the crystal structure, this anionic group appears to be important for the final correct assembly of the structure.

CORRELATION WITH EXPERIMENTAL PREDICTIONS

Several chemical approaches identified amino acid side chains that contribute to peptide recognition. Differential labeling with a water soluble carbodiimide, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC), targeted solvent-accessible carboxyl groups that were accessible in the free C-subunit but protected in the presence of substrate (Buechler et al. Biochemistry 29:1937-1943, 1990). Two regions were identified using this approach. Glu170 was very reactive in the absence of peptide, but fully protected in the presence of peptide. The other region was the cluster of carboxyl groups near the C-terminus, Asp³²⁸. Asp-Tyr-Glu-Glu-Glu-Glu³³⁴. As indicated in Figure 12, Glu170 interacts with the P-2 Arg while the cluster of carboxyl groups flanks the P-3 site.

The crystal structure localized Cys199 close to the peptide recognition site and to the γ -PO₄ subsite of ATP. In the binary complex, Cys 199 does not appear to participate in peptide binding other than to contribute to the general hydrophobic environment around the P+1 site.

CONFORMATIONAL CHANGES ASSOCIATED WITH PEPTIDE BINDING

Substrate-induced conformational changes are associated with peptide binding to the catalytic subunit. Global changes in conformation, first observed using circular dichroism, showed both a loss of alpha-helical content and an increase in beta structure following peptide binding. A global change in shape also was observed using low angle neutron scattering. These results demonstrated a reduction of the radius of gyration (Rg) following substrate binding and furthermore established that the inhibitor peptide alone, but not MgATP, was sufficient to cause the reduction in Rg. The substrate-induced reduction in Rg

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indicates that the apoenzyme corresponds to an open configuration of the protein while the binary and ternary complex represent a closed configuration.

The recognition of the peptide by the catalytic subunit is believed to be a multistep process. The initial step, associated with a loss in α -helical structure, was induced by both the substrate and inhibitor heptapeptides shown in 2. The second step, presumably corresponding to the final orientation of the peptide into the correct position at the active site, was associated with an increase in β -structure and could only be accomplished with the substrate peptide, not by the Ala peptide inhibitor. This increase in β -structure is probably due, in part, to the P+1 region of the peptide interacting the protein. Understanding these substrate-induced conformational changes will eventually require a detailed comparison of the apoenzyme structure with binary and ternary complexes containing inhibitors and substrate peptides both in the presence and absence of MgATP.

The peptide-induced conformational changes in catalytic subunit may reflect a closing of the cleft and probably involve the region linking the small and large lobes as well. This linker region consists of two chains: residues 123 through 127 and a highly acidic segment, residues 328-334. The P-3 peptide binding site is the only region of the inhibitor peptide that interacts directly with both of these extended chains that link the two lobes. One anionic group at the P-3 site is Glu127 and the other is Glu331. Since several of the carboxy groups in the C-terminal linking chain also interact with portions of the nucleotide binding site, even in the binary complex, this P-3 residue may contribute to the substrate-induced conformational changes.

CONSERVED AND VARIABLE SITES IN PROTEIN KINASES

The recognition of a protein substrate by the catalytic subunit is not unlike the recognition of a protein antigen by the variable domain of an immunoglobulin. The binding sites of both structures are dominated by interfacing β -sheets surrounded by loops that participate in recognition of the protein. The catalytic subunit also has helical regions, but it is the β -sheets that converge at the active site and it is the loops that play the dominant role in peptide recognition and catalysis. One β -sheet comes from the small lobe and the other from the large lobe. These two sheets are sandwiched together at the cleft. In the case of protein kinases, two of the loops are essential for catalysis and are highly conserved, unlike the immunoglobulins, whose function is only to bind antigens.

The two essential conserved loops that assemble at the site of catalysis in the catalytic subunit, seen in Figures 14 and 15, are the glycine-rich loop in the small lobe and the catalytic loop in the large lobe. Both lie on the surface that lines the cleft between the

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two lobes. The glycine-rich loop serves as an anchor for the phosphates of MgATP, whereas the catalytic loop is essential for peptide binding and catalysis. Key features of the active site of the catalytic subunit are shown in Figure 14. Nine of the amino acids that are nearly invariant in all protein kinases are indicated. Gly186, another invariant residue, is not shown. The alpha carbons are in black, oxygens dotted, and nitrogens in horizontal hatching. The position of the phosphorylation site at Thr197 is indicated by vertical hatching. The portion of the active site associated with the small lobe is shaded and includes three of the invariant amino acids, Gly 52, Lys72, and Glu91. The remaining six are located in the large lobe. Residues close enough for hydrogen bonding or ion pairing are indicated by a dashed line while residues within 4-5 Å of one another are connected by a dotted line. As seen in Figure 14, seven of the nine invariant amino acids conserved in all protein kinases are located here, either in the loops themselves or connecting directly with loop residues. The single invariant glycine, Gly52, lies in the phosphate anchoring loop. The proposed catalytic base, Asp166, as well as Asn171, are in the catalytic loop. It is remarkable how thoroughly interconnected this region is with multiple ion pairs providing a finely tuned scaffolding for communication at the active site.

The three invariant residues in the small lobe all participate in nucleotide binding. Unlike Gly52, which is part of a flexible loop, both Lys72 and Glu91 are anchored to defined parts of the secondary structure - Lys72 to β -strand 3 and Glu91 to the C-helix. The difference Fourier map shows the phosphate density near these residues, with the presumed γ - PO_4 density close enough to the P Ala C_β for phosphotransfer were it a Ser[Thr] and indicates that these residues play a key role in the recognition of the phosphates of MgATP.

In the catalytic loop the two invariant residues, Asp166 and Asn171, interact with each other. Not only are their side chains close, but, more importantly, the nitrogen in the amide side chain of Asn171 is less than 3Å from the backbone carbonyl of Asp166. One additional nearly invariant residue, Asp220, contributes directly to stabilization of the catalytic loop. The two oxygens of this carboxylate come with hydrogen bonding distance of the backbone carbonyl and amide of residue 164 that immediately precedes the loop. The interaction of the catalytic loop with a conserved residue that lies deep within the large lobe fixes the loop from one side while peptide binding and interactions with the small lobe fix it from the opposite direction. As seen in Figure 13A and 13B, the consensus region of the peptide is sandwiched between the P+1 site on one side and the glycine-rich loop on the other side.

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Of all the invariant residues, Asp184 is the only one that appears to communicate with both the small lobe and the large lobe. In the binary complex, it is most closely associated with Lys72, but it is also only 4-5Å from Asn171 and Asp166 in the catalytic loop. Although not shown in Figure 14, Asp184 is itself part of a tight turn with the carboxylate located within hydrogen bonding distance of the backbone amide of Gly186, another invariant residue. This entire segment, Asp166-Phe-Gly, is highly conserved in all protein kinases, and hydrogen bonding to stabilize the turn is probably conserved as well. Asp184 certainly has the potential to shuttle between the two conserved loops, and it is anticipated that the contacts of Asp184 will differ somewhat in both the apoenzyme and in the ternary complex containing bound MgATP as well as peptide. If Asp184 participates in the chelation of Mg^{2+} , as disclosed above, then this charge will be sequestered from the immediate environment of the catalytic loop. Other residues close to the conserved residues in the catalytic loop in the binary complex are Tyr164 and Lys 168. The Tyr 164 side chain is less than 3Å from the side chain nitrogen of Asn171, and the Lys168 side chain comes close to the carboxylate of Asp166. Either Tyr or His, another good hydrogen-bonding residue, is always found at position 164, so this contact can also be conserved. Any significant change in the position of Asp184 will likely change the environment of the catalytic loop. Asp184, as well as Asn171 and Asp166, have also been identified as a sequence motif associated with many phosphotransferases, and this may represent a common mechanism among protein kinases.

The versatility and importance of the catalytic loop is highlighted not only by the conserved networking of essential amino acids at the active site, but also by the special ways in which this conserved network communicates with the variable residues that compose the peptide binding sites. This communication specifically involves loop residues that are not highly conserved. Glu170, for example, contributes directly to the anionic P-2 site. Thr201 in the P+1 site, on the other hand, comes very close to the side chain of Asp166. These two particular regions of contact involving the peptide binding site and the catalytic loop, Lys¹⁶⁸-Pro-Glu and Thr²⁰¹-Pro-Glu-Tyr-Leu-Ala-Pro-Glu, contain sequences that differ characteristically between the kinases that transfer phosphate to Ser/Thr and those that transfer phosphate to tyrosine (Hanks et al., *supra*).

Arg165 is actually highly conserved in most protein kinases, and it connects in a unique way with the P+1 peptide binding site. Specifically, it points towards the phosphothreonine and helps to fix that phosphate so that the hydrophobic groove that follows and provides a pocket for the side chain of the P+1 residue is firmly positioned

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(Figure 13C). This is an autophosphorylation site, and it is the only phosphorylation site in the catalytic subunit that could conceivably result from an intramolecular autophosphorylation. Chemical analysis has shown that this phosphate is very resistant to removal by phosphatases and based on this crystallographic data, appears to contribute to the final conformation stability of the enzyme. It should be emphasized as well that a phosphorylation site in this region of the protein is not a conserved feature of all protein kinases. Some kinases such as pp60^{C-Src}, a protooncogene whose viral counterpart is found in Rous Sarcoma Virus, do have an autophosphorylation site nearby, but many others do not. Whether the catalytic loop communicates in unique ways with other autophosphorylation sites in other protein kinases remains to be established.

The two invariant residues that are most distant from the active site are Asp208 and Arg280. These residues constitute a conserved ion pair that lies just beneath the P+1 site and appears to stabilize a very hydrophobic region that buttresses the P+1 peptide binding site.

Figure 15 is a schematic of the conserved and variable regions of the catalytic subunit. The ribbon diagram depicts the folding of the catalytic subunit. Conserved regions include two loops - the glycine-rich loop and the catalytic loop - and are indicated. The variable peptide binding sites are shown as solid areas. Invariant amino acids Gly52, Lys72, Glu91, Asp166, Asn171, Asp184, Glu208, Asp220, and Arg280 are indicated by a large dot and are numbered. Dashed lines indicate residues that are close enough to pair, while the dotted line extends from Arg165 to the Thr197. Several points should be emphasized regarding the recognition of a peptide or protein substrate by the catalytic subunit. First is the number of sites and their diversity. Some of these peripheral peptide recognition sites are hydrophilic and highly charged; others are hydrophobic. As seen in Figure 15, most are found within the large lobe of the catalytic core shared by all protein kinases, but some also lie outside of this boundary. A second observation is that the requirements for recognition at the consensus site are not absolute. A comparison of *in vivo* phosphorylation sites reveals that the actual residues at each site vary somewhat as does the spacing between the positively charge side chains and the site of phosphotransfer. Thus, even in the consensus region, some variability can be tolerated. A third point is the potential for variability in recognition of different inhibitor, and presumably substrate, proteins that bind with a high affinity to the catalytic subunit. Most of the features essential for the high affinity recognition of PKI are apparent from this structure of the binary complex. The regulatory subunit, however, also binds to the C-subunit with a subnanomolar affinity in the absence

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of cAMP. The consensus region, P-3 through P+1, is shared by both molecules. However, the R-subunit, cleaved at the P-5 position, still retains its high affinity binding for the C-subunit. In addition, the P-16 to P+1 region of the R^I-subunit is Pro-Pro-Pro-Pro-Asn-Pro-Val-Val-Lys-Gly-Arg-Arg-Arg-Arg-Gly-Ala-Ile, and this certainly cannot conform to the helical motif that dominates the corresponding region of PKI(5-24). Hence, an amphipathic helix is not required for the high affinity binding of the regulatory subunit. Instead, the residues that contribute to the high affinity binding of the regulatory subunit, specifically, must lie beyond the P+3 position and may complement a different portion of the surface of the C-subunit. This variability presumably can also extend to protein substrates where the catalytic subunit may recognize unique sequences that lie outside the consensus site.

Unlike the conserved residues that are invariant in all protein kinases, the sites involved in peptide recognition differ for each kinase. About 30% have some general similarities to cAPK. Others are quite different. However the template allows us to predict the specificity of each contact point. Figure 16 provides the sequence of PKI(5-24) and illustrates the distances between the points of contact and the catalytic site in three-dimensional space as measured from the template. The P site or site of catalysis is denoted by an arrow. Asterisks designate sites particularly important for the high affinity binding of PKI(5-24). Recognition sites essential for PKI binding to other substrates are denoted as labelled archways p+1, p-2, p-3, p-6, and p-11. All of the distances, with the exception of the p+1 site, are greater than 5 Å. That positions 5 Å or greater from the site of catalysis are important for inhibitor specificity have heretofore been undisclosed.

The identification of the subsites that are important to maintaining the specificity of the effector molecule interaction and provide K_d less than 100nM facilitates the design of other inhibitors. PKI(5-24) can be used as a scaffold for molding new inhibitors, and in addition once the electrochemical interactions are understood from an analysis of the three dimensional template, other effectors that are not peptides can additionally be identified. Thus, effectors could come from a group including but not necessarily limited to peptides, polypeptides, unmodified molecules existing in nature, synthetic molecules, nucleic acids, polymers, organics, or hydrocarbons. Molecules that exist in nature and that are known to interact with enzymes could be modified to produce effector molecules. Examples from this group include antibodies, antibiotics, protein, other enzymes, lipids, polysaccharides, saccharides and vitamins. Thus, inhibitors can be designed that utilize both conserved and nonconserved points of contact.

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The invariant residues within the protein kinase family and specifically, cAPK, are used to apply the template and its coordinates to other protein kinases. There are 8-9 invariant residues for the protein kinase family. Other families may have differing numbers of invariant residues. Table 4 list the invariant residues and the distances between these
5 residues. The distances are calculated between α -carbons. The distances between residues 52, 72 and 91 are expected to remain close to constant since these residues are all in the amino-terminal domain of the protein. Similarly, the distances between residues 166, 171, 184, 186, 208 and 280 would be expected to remain constant due to their being in the carboxy-terminal domain. Motion of the amino-terminal domain relative to the carboxy
10 terminal domain is expected to change the distances between residues in different domains.

The distances calculated in Table 4 help form the model template since these three-dimensional positions are taken from the crystal diffraction patterns and help to define a conserved shape for the protein kinase family catalytic core. A point of contact is defined herein to occur at the invariant residues and is additionally defined as a point of close
15 spatial approximation between the atoms of the residues within or around the catalytic core and the atoms of the effector. These points of contact affect the specificity and the Kd of the enzyme/effector interaction.

The template is best described by Figure 11. The coordinates for the template listed in Table 4 and Figure 17 provide the spatial characteristics that permit one of skill in the
20 art to input the template structure into a computer program and perform the invention disclosed herein. While the coordinates together define a three-dimensional surface that permits visualization of the catalytic site, there are invariant residues that establish important foci within the structure.

Lys 72 is invariable within the catalytic site for the protein kinase family and is an
25 anchor for superimposing other protein kinases onto the template. Asp 166 can additionally be a second important anchor. Similarly the other invariant positions likewise have importance for fitting other kinases. A combination of the coordinates with the invariant residue positions allows important regions within and around the catalytic site to be visualized. From a study of the interaction of cAPK with PKI(5-24), important hydrophobic and ionic interactions can be analyzed. When a new enzyme is superimposed onto these
30 coordinates these hydrophobic and ionic interactions are assessed with PKI(5-24). It is then possible to study what changes can be made to PKI(5-24) to model a new effector. A study of the residue sidechains and the charge distribution within the site is used to fine tune the new effector.

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Any protein kinase having homology in and around the catalytic site with cAPK can be used to design specific effector molecules. Hanks et al. provides a list with homologous residues highlighted. Many growth factor receptors have protein kinase activities. These include but are not limited to platelet-derived growth factor, colony stimulating factor, the insulin receptor family and epidermal growth factor. Protein kinases are involved in hematopoiesis and lymphopoiesis. Some, like myosin light chain kinase, are calcium-calmodulin dependent, and further, a variety of protein kinases are oncogenic products. These include but are not limited to viral and cellular homologues of src, mos, abl, Neu, Fgr, and Yes. Any of these kinases as well as others fitting the characteristics disclosed herein could be used in this invention to produce specific effector molecules.

The phosphorylation target sequences are available for a variety of protein kinases. These include phosphorylatable amino acids with their surrounding residues. For some kinases this will provide a good starting point for inhibitor design. Other protein kinases have a regulatory subunit associated with the catalytic subunit in the inactive form. The binding sequences with the regulatory subunits are other starting points for effector molecule design. Additionally, there are a group of protein kinases that have a regulatory domain. This domain binds the catalytic site when the enzyme is inactive. Binding of an exogenous molecule changes the kinase conformation such that the regulatory domain no longer binds. A review by Pearson et al. provides a table of protein kinase phosphorylation site sequences (Methods in Enzymology Vol. 200, 1991 in press).

Once a template is created there are several options available for designing an effector molecule and these were outlined in the section above entitled "Brief Description of Effector Design."

EXAMPLE 6

Inhibitor design for pp60^{c-src} without pp60^{c-src} purification

pp60^{c-src} is the proto-oncogene homologue of the src protein kinase from Rous Sarcoma Virus. The protein causes unrestrained cell proliferation. In this example, the invariant residues for pp60^{c-src} are identified with a star and in bold below:

ESLRLEVKLGQGCFGEVWMGTWNGTTRVAIKTLKPGTMSPEAFLQEAQVMKKL
RHEKLV

OLYAVVSEEPITYVTEYMSKGSLLDFLKGETGKYLRPLQVDMAAQIASGMAYBE
RMNY

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VHRDLRAANILVGENLVCKVADFGRLIEDNEYTARQGAKFPIKWTAPEAALY
GRFTI

5 KSDVWSFGILLTELTTKGRVPYPGMVNREVLQDQVERGYRMPCPPECPSLHDLH
CQCWR

*
KEPEERPTFEYLQAFLEDYFTST

10 These residues are incorporated into the appropriate position from the invariant
residues listed in Table 4 using the coordinate set provided in Figure 17. Once the template
is in place and the catalytic site from pp60^{c-src} has been superimposed onto the template,
it is possible to visualize the catalytic site. The site can additionally be refined using the
complementary target phosphorylation site for pp60^{c-src}.

RLIEDNEY*TARQGAK

15 * denotes the site of phosphorylation.

Residues are altered using computer modelling until a fit is achieved for pp60^{c-src}
on the template. Thus, residues 184, 166, 172, 220, 208, and 280 from the pp60 c-src
sequence have positions in space that maintain those distances disclosed in Table 4. Ionic
and hydrophobic amino acid side chains are matched within the catalytic core with
20 complementary residues to create a new inhibitor molecule. Recombinant cAPK is then
mutated to duplicate the three-dimensional structure within the core. Crystals of mutated
cAPK are analyzed alone or together with a proposed inhibitor. The structure is again
analyzed in the context of the invariant residues listed in Table 4.

25 Positions 52, 72, and 91 are mobile invariant residues whose positions will vary
depending on the quality of inhibitor. The distances of these residues are listed in Table
4 for PKI(5-24) and cAPK. It is anticipated that peptide inhibitors of equal affinity for
cAPK will have similar distances. Non-peptide inhibitors can be designed that do not
produce a rotation, or fraction of fit, exactly in the same direction as peptide inhibitors, such
as PKI(5-24). A comparison of the crystal structure of cAPK and cAPK with PKI(5-24)
30 indicate that positions 52, 72 and 91 rotate 12° toward residues 184, 166, 172, 220, 208 and
280. This rotation defines a range of peptide inhibitors. Another strong peptide inhibitor
will similarly produce a 12° rotation toward the six residues listed above while residues 52,
72 and 91 may have a smaller angle of rotation for weaker peptide inhibitors.

In addition, there are important points of contact between cAPK and PKI(5-24).
35 The specific contact amino acids on PKI(5-24) are starred and the corresponding points of
contact within the catalytic core of cAPK are identified as positions p+1, p-2, p-3, p-6 and
p-11. These points of contact are conserved within the catalytic core of all protein kinases

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and similar points of contact will be readily identifiable to those of skill in the art for other protein kinases. A sphere of influence having a radius of 11Å or less, more preferably 6Å or less, and extending from the inhibitor around the points of contact at positions p+1, p-2, p-3, p-6, and p-11 can be used to define regions that are critical for inhibitor specificity.

5 As described above, in connection with Figure 11, the points of contact can be used to identify the replacements necessary to design appropriate inhibitors or other effectors for a new enzyme. Thus, amino acid replacements are used which form appropriate ionic and hydrophobic interactions at these points of contact. Hydrogen bonding interactions are also preferably used to identify replacements. Of course, the modelling can extend beyond the
10 identified points of contacts in order to provide still further specificity

 This same type of analysis can be performed with the mutated cAPK that mimics the catalytic core of pp60^{c-src}. Thus, an analysis of the mutant crystals permits one to predict the affinity of a given inhibitor. The inhibitor can be further modified to improve the ionic and hydrophobic interactions surrounding the points of contact using the spheres of
15 influence described above. The angle of rotation of the mobile invariant residues can be used to predict whether or not a given peptide inhibitor will be useful. These changes are all performed within the constraints of the coordinates of Figure 17.

 A peptide inhibitor that, once modelled has distances similar to Table 4 and meets the design criteria described above can be synthesized and tested for function *in vitro* or *in*
20 *vivo*.

 The coordinates obtained from the binary complex and the resulting template allow us for the first time to fully appreciate the complexity and sophistication of the process by which a protein kinase recognizes its protein substrate. While peptide analogues provide important clues, the diversity of the peptide binding sites and their dispersion over such a
25 wide area on the enzyme surface makes it imperative to have structural data on complexes of the enzyme with effector molecules. The structure of the binary complex of cAPK with PKI(5-24) provides, for the first time, a molecular basis for the rational design of effector molecules, both peptide and nonpeptide, that can target specific protein kinases. Furthermore, because the basic catalytic core of this enzyme is so conserved in all protein
30 kinases, a template based on the crystal structure can also serve as a mold for modelling for other protein kinases.

 Although this invention has been described using protein kinases as a model system, with cAPK being shown as a specific example of an enzyme for determining the template, the present invention is not intended to be limited to this model. Other changes to the

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methods described herein will suggest themselves to those of ordinary skill in the art. Accordingly, the spirit and scope of the present invention is to be determined with reference to the appendant claims.

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WE CLAIM:

1. A method of designing a highly specific effector which exerts an effect on the activity of a first enzyme, said first enzyme being a member of a class of enzymes having a conserved catalytic core, comprising:

5 identifying a second enzyme that is a member of said class in which a first effector can affect the activity of said second enzyme;

forming a first complex of said first effector and said second enzyme;

obtaining data regarding the conformation of said second enzyme at sites greater than 5 Å from the site of catalysis of said second enzyme in said first complex;

10 designing an effector which induces a conformation on said first enzyme at sites greater than 5 Å from the site of catalysis thereof which is homologous to the conformation of the conformation of said second enzyme at homologous sites in said first complex, when said effector is formed as a second complex with said first enzyme; and

15 producing said effector.

2. The method of Claim 1, additionally comprising crystallizing said first complex and obtaining X-ray crystallography data therefrom.

3. The method of Claim 1, wherein the designing step comprises:

20 identifying a potential effector likely to induce a conformation on the catalytic core of said first enzyme which is homologous to the conformation of the catalytic core of said second enzyme in said first complex, when said effector is formed as a second complex with said first enzyme; and

determining whether said potential effector induces said conformation

25 through the use of a technique selected from the group consisting of computer modelling, small angle neutron scattering, and circular dichroism.

4. The method of Claim 3, wherein said potential effector comprises a peptide.

5. The method of Claim 3, wherein said potential effector comprises a molecule selected from the group consisting of nucleotides, polynucleotides, alcohols, polymers, lipids carbohydrates, sugars, native or synthetic molecules, protein and organic compounds and combinations thereof.

30 6. The method of Claim 1, wherein all of the members of said class have related functions.

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7. The method of Claim 1, wherein the catalytic cores of all of the members of said class have conserved amino acid residues.

8. The method of Claim 7, wherein the designing step comprises designing an affector having homologous topography and charge fields that complement the catalytic core of said first enzyme and capable of inducing a conformation wherein the conserved amino acid residues of said first enzyme are in homologous locations to said second enzyme in said first complex.

9. The method of Claim 1, wherein each of the effectors is an inhibitor.

10. The method of Claims 1, wherein each of the effectors is an activator.

11. The method of Claim 1, wherein said first affector comprises all or a portion of said first enzyme.

12. The method of Claim 1, wherein said first complex is a holoenzyme.

13. The affector produced by the method of Claim 1.

14. The method of Claim 7, wherein said class of enzymes comprises protein kinases.

15. The method of Claim 14, wherein said second enzyme is a viral oncogene product or a cellular homologue thereof.

16. The method of Claim 15, wherein said second enzyme is p60 v-Src from RSV or its cellular homologue, pp60 c-src.

17. The method of Claim 16, wherein said second enzyme comprises cAMP-dependent protein kinase.

18. The method of Claim 2, wherein said second enzyme comprises a native mammalian protein kinase.

19. The method of Claim 2, wherein said second enzyme comprises recombinant protein kinase.

20. A method of designing a highly specific affector which exerts an effect on the activity of a first enzyme, said first enzyme being a member of a class of enzymes having conserved residues at an affector binding site, comprising:

identifying a second enzyme that is a member of said class in which a first affector can affect the activity of said second enzyme, said first affector having a dissociation constant with said second enzyme of less than 1 μ M;

forming a first complex of said first affector and said second enzyme;

obtaining data regarding the conformation of the affector binding site of said second enzyme in said first complex;

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designing an effector which induces a conformation on the effector binding site of said first enzyme which is homologous to the conformation of the effector binding site of said second enzyme in said first complex, when said effector is formed as a second complex with said first enzyme; and

5 producing said effector.

21. The method of Claim 20, wherein said class of enzymes have a nucleotide binding site and each of said effectors is capable of binding to said nucleotide binding site.

22. A method of designing a highly specific effector which exerts an effect on the activity of a first enzyme, said first enzyme being a member of a class of enzymes having a conserved catalytic core, comprising:

10

 identifying a second enzyme that is a member of said class in which a first effector can affect the activity of said second enzyme;

 forming a first complex of said first effector and said second enzyme, said first complex having at least three points of contact between said first effector and second enzyme;

15

 obtaining data regarding the conformation of the catalytic core of said second enzyme in said first complex;

 designing an effector which induces a conformation on the catalytic core of said first enzyme which is homologous to the conformation of the catalytic core of said second enzyme in said first complex, when said effector is formed as a second complex with said first enzyme; and

20

 producing said effector

23. A crystallized protein kinase/effector complex having stable decay characteristics over 15 minutes.

24. A crystallized protein kinase/effector complex having a Bragg spacing diffraction limit of less than 4Å.

25

25. The crystallized protein kinase of Claim 24 having stable decay characteristics over 15 minutes.

26. A crystallized complex of a cAMP-dependent protein kinase and an inhibitor thereof.

30

27. Use of the crystallized complex of Claim 26 in an X-ray crystallography procedure to produce data regarding the three dimensional structure of said cAMP-dependent protein kinase in said complex.

28. Use of the data produced by Claim 27 for designing an inhibitor of a second protein kinase which will induce a similar three dimensional structure on the active site of said second protein kinase as the three dimensional structure of said cAMP-dependent protein kinase in said complex.

5 29. An inhibitor designed by Claim 28.

30. A method of preparing a highly specific effector of a first enzyme, said first enzyme being a member of a class of enzymes having a conserved catalytic core with a plurality of invariant residues, comprising:

- 10 a. identifying a second enzyme that is a member of said class and having a known effector thereof;
- b. forming a first complex of said second enzyme and said known effector;
- c. obtaining data regarding the three dimensional coordinates of said invariant residues in said first complex, said coordinates forming a template;
- 15 d. generating a model wherein said first enzyme is in a conformation in which said invariant residues are in substantially the same conformation as in said template;
- e. identifying a change in the variable residues in the catalytic core of said first enzyme in the conformation of step (d) when compared to the variable residues in the catalytic core of said second enzyme in the conformation of step (b);
- 20 f. preparing a modified form of said second enzyme, wherein the modified second enzyme includes the non-conserved change identified in step (e);
- g. designing an effector which computer modelling indicates will induce a conformation on the catalytic core of said first enzyme in which said invariant residues are in substantially the same coordinates as the coordinates of said template, when said first enzyme is formed as a second complex with the effector designed in this step; and
- 25 h. producing said effector.

31. The method of Claim 30 wherein said change is a non-conserved change in the variable residues.

32. The method of Claim 30, additionally comprising:

- i. forming a third complex of said modified second enzyme and an effector capable of binding thereto;

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j. obtaining data regarding the three dimensional coordinates of the invariant residues in said third complex; and

k. using the data obtained in step (i) to design an affector which computer modelling indicates will induce a conformation on the catalytic core of said first enzyme in which said invariant residues are closer to the coordinates of said template than the conformation induced by the affector designed in step (g), when said first enzyme is formed as a fourth complex with the affector designed in this step.

33. The method of Claim 32 wherein the affector of step (i) is the known affector of step (a).

34. The method of Claim 32, additionally comprising modifying the computer modelling used in step (g) in light of the data of step (j), prior to performing step (k).

35. The method of Claim 30, additionally comprising obtaining amino acid sequence data relating to the catalytic cores of the first and second enzymes.

36. The method of Claim 30 wherein step (f) comprises site directed mutagenesis of a recombinantly produced second enzyme.

37. The method of Claim 30, wherein the coordinates of said template are substantially as shown in Figure 17.

38. The method of Claim 30, wherein each of the effectors is an inhibitor.

39. The method of Claim 30, wherein said template includes coordinates separated by the distances substantially as shown in Table 4.

40. An affector prepared by the method of Claim 39.

41. A pharmaceutical composition comprising the affector of Claim 40.

42. A method of designing a specific inhibitor for a protein kinase, comprising: obtaining data regarding the three-dimensional structure of a first protein kinase;

using said data in the design of an inhibitor for a second, different, protein kinase; and

producing said inhibitor.

43. The method of Claim 42 wherein said first protein kinase is cAMP dependent protein kinase or an analogue thereof.

44. The method of Claim 43, wherein the obtaining step comprises crystallizing a complex of cAMP dependent protein kinase and an inhibitor thereof.

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45. The method of Claim 44, wherein the obtaining step additionally comprises obtaining X-ray crystallography data on the crystallized complex obtained from the crystallizing step.

5 46. The method of Claim 44, additionally comprising obtaining information regarding the structure of the crystallized complex obtained in the crystallizing step through circular dichroism, small angle neutron scattering, or other chemical procedures.

47. Use of the data of Figure 17 or of Table 4 in the design of an affector for a protein kinase.

10 48. A method of preparing a highly specific inhibitor of a first enzyme, said first enzyme being a member of a class of enzymes having a conserved catalytic core with a plurality of invariant residues, comprising:

a. identifying a second enzyme that is a member of said class and having a known first inhibitor thereof;

b. forming a first complex of said second enzyme and said first inhibitor;

15 c. obtaining data regarding the three dimensional coordinates of said invariant residues in said first complex;

d. designing a second inhibitor which computer modelling indicates will induce a conformation on the catalytic core of said first enzyme in which said invariant residues are in substantially the same coordinates as the coordinates
20 obtained in step (c), when said first enzyme is formed as a second complex with said second inhibitor;

e. preparing said second inhibitor;

f. forming a third complex of said second inhibitor and a third enzyme complexable therewith, said third enzyme having a plurality of said invariant
25 residues;

g. obtaining data regarding the three dimensional coordinates of said invariant residues in said third complex;

h. using the data obtained from step (g) to design a third inhibitor which computer modelling indicates will induce a conformation on the catalytic core of said
30 first enzyme closer to that in which said invariant residues are in substantially the same coordinates as the coordinates obtained in step (c) than induced by the second inhibitor, when said first enzyme is formed as a fourth complex with said third inhibitor; and

i. producing said third inhibitor.

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49. The method of Claim 48, wherein said first inhibitor is an inhibitory domain of said second enzyme.

50. The method of Claim 48, wherein said third enzyme comprises at least 5 invariant residues.

5 51. The method of Claim 48, wherein said third enzyme is a naturally occurring enzyme.

52. The method of Claim 48, wherein said third enzyme is a mutant enzyme.

53. A method of determining the efficacy of a first effector in affecting a first enzyme that is a member of a class of enzymes having a plurality of invariant residues
10 among the members of said class, comprising:

determining the three dimensional coordinates of the invariant residues of a second enzyme in a first conformation wherein said second enzyme is in a complex with a second effector that is a strong effector of said enzyme;

15 determining the three dimensional coordinates of the invariant residues of said second enzyme in a second conformation wherein said enzyme is in a conformation other than said first conformation;

identifying the mobile invariant residues of said enzyme, said mobile invariant residues being those invariant residues at coordinates substantially different in said first conformation than in said second conformation;

20 determining the three dimensional coordinates of the mobile invariant residues of said first enzyme when said first enzyme is in a conformation wherein said first enzyme is in a complex with said first effector;

25 comparing the three dimensional coordinates of the mobile invariant residues of said first enzyme in said conformation with the coordinates of the mobile invariant residues of said enzyme in said first conformation, wherein relatively small differences in the coordinates indicate relatively high efficacy of said first effector.

54. The method of Claim 53, wherein the step of determining the coordinates of said first enzyme in said conformation is performed using computer modelling of said conformation.

30 55. The method of Claim 53, wherein the steps of determining the first and second conformations comprise obtaining X-ray crystallographic data of said enzyme.

56. The method of Claim 53, wherein said second conformation is a conformation produced by a ternary complex.

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57. The method of Claim 56, wherein said ternary complex comprises a protein kinase, a nucleotide and an effector.

58. The method of Claim 53, wherein said second conformation is a conformation produced by said second enzyme not complexed with a ligand.

5 59. The method of Claim 53, wherein said second enzyme is the same enzyme as said first enzyme.

60. A method of designing specific inhibitors of a first protein kinase having a template defined by a plurality of invariant residues present in substantially all protein kinases, comprising:

10 obtaining data regarding the three dimensional coordinates of the invariant residues of a second protein kinase and of the points of contact between said second protein kinase and a known inhibitor thereof, said coordinates being obtained when said second protein kinase is formed as a complex with said known inhibitor;

15 generating a model of said first protein complex wherein said template is defined by the positions of said invariant residues in said complex;

examining the amino acid residues present in said first protein kinase at positions corresponding to the points of contact in said complex;

designing an inhibitor of said first protein kinase capable of forming ionic and hydrophobic interactions with said amino acid residues; and

20 producing said inhibitor of said first protein kinase.

61. The method of Claim 60, wherein said second protein kinase is cAMP dependent protein kinase.

62. The method of Claim 61, wherein said known inhibitor is PKI(5-24).

25 63. The method of Claim 62, wherein the points of contact in said complex comprise positions corresponding to the positions selected from the group consisting of P+1, P-2, P-3, P-6 and P-11 along said known inhibitor.

64. The method of Claim 60, wherein the positions corresponding to the points of contact in the examining step comprise positions within a sphere having a radius of 11 Å from the coordinates of the point of contact obtained in the obtaining step.

30 65. The method of Claim 64, wherein the positions corresponding to the points of contact in the examining step comprise positions within a sphere having a radius of 6 Å from the coordinates of the point of contact obtained in the obtaining step.

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66. The method of Claim 60, wherein the designing step additionally comprises designing said inhibitor to form appropriate hydrogen bonding with said amino acid residues.

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FIGURE 1

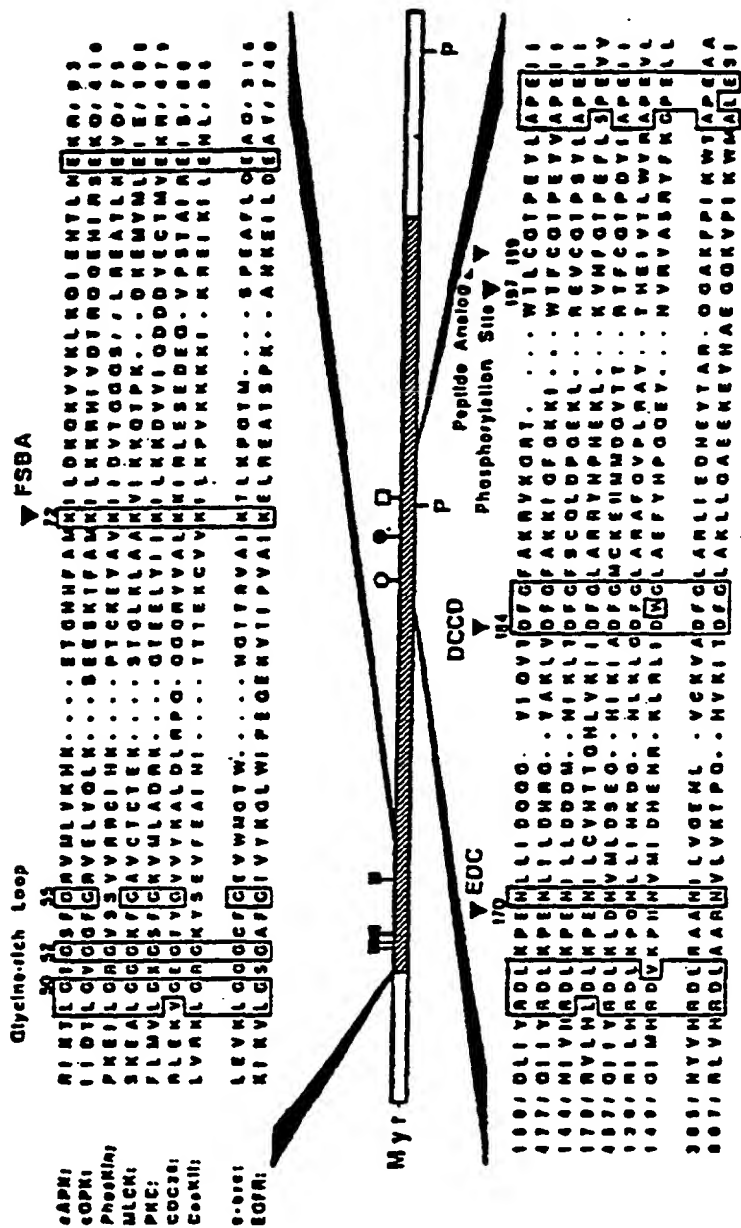
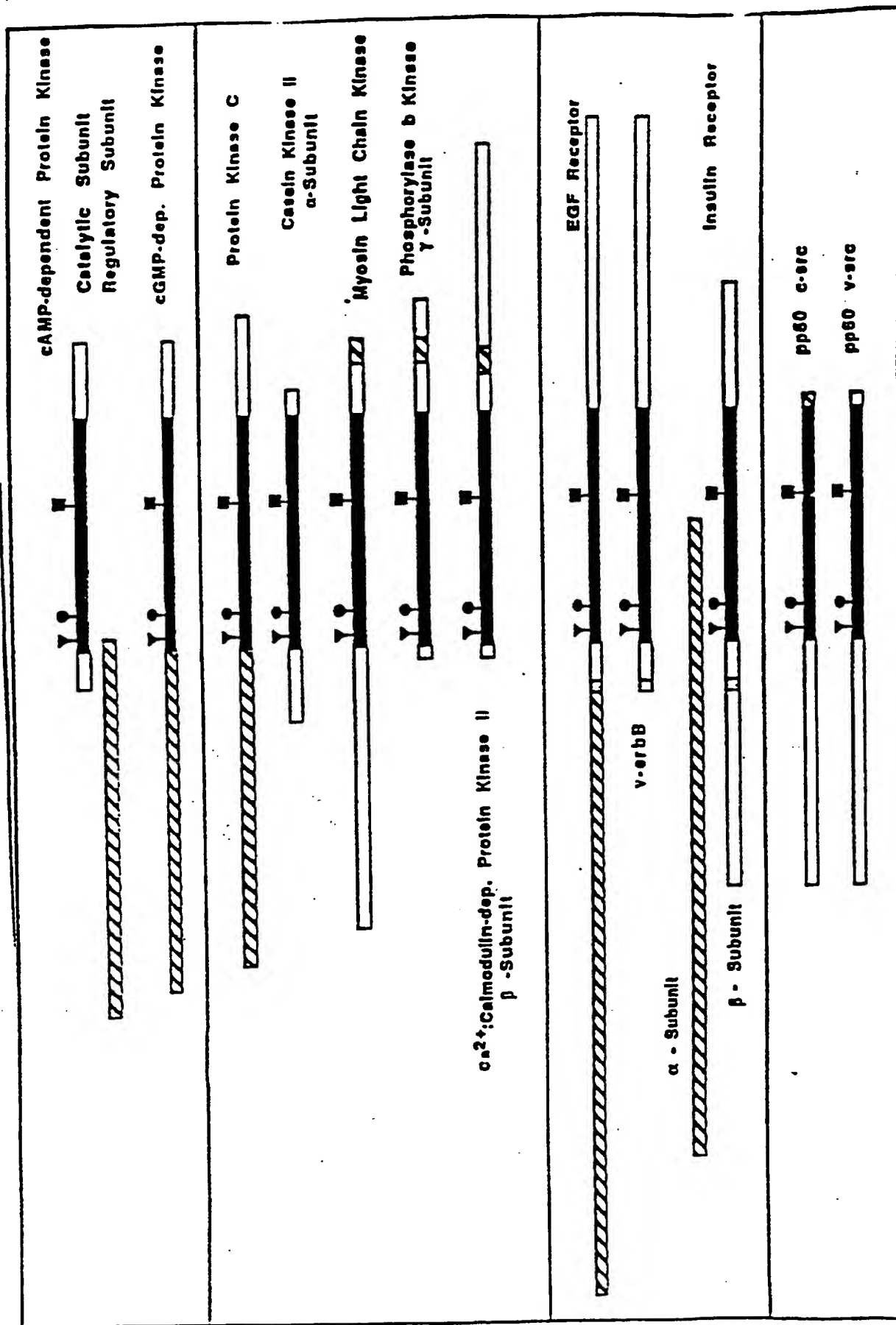


FIGURE 2



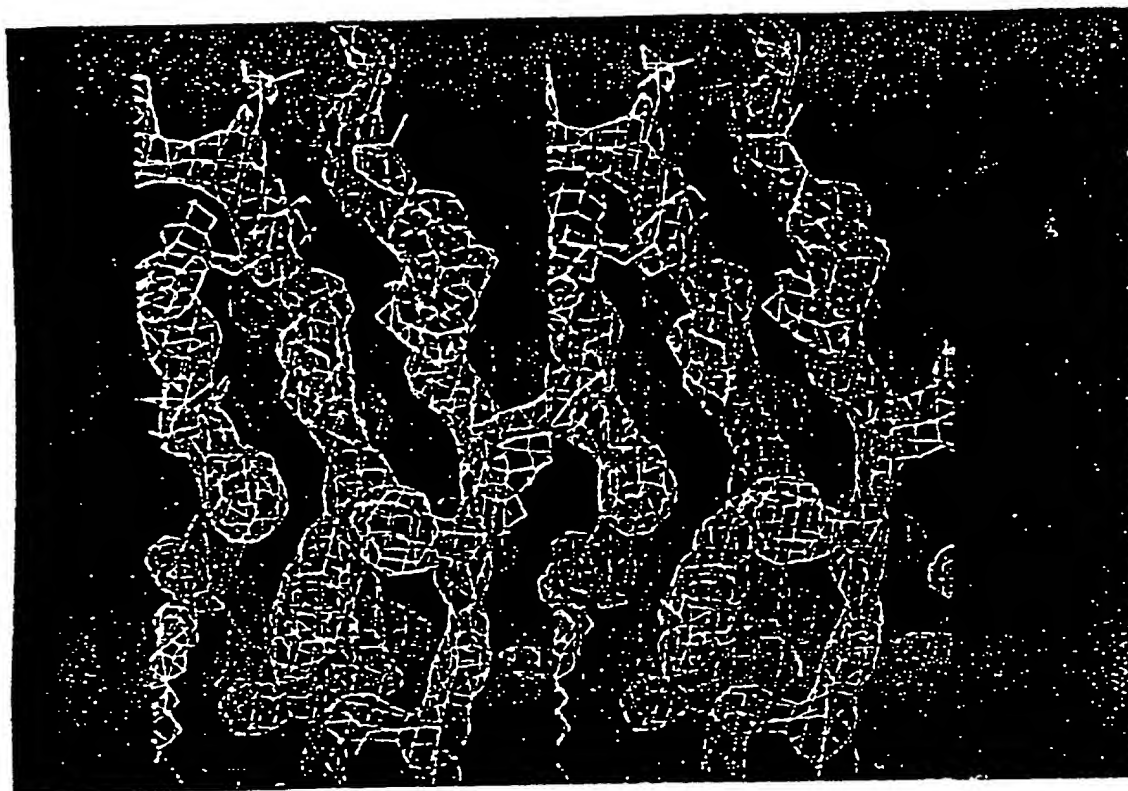


FIGURE 3A

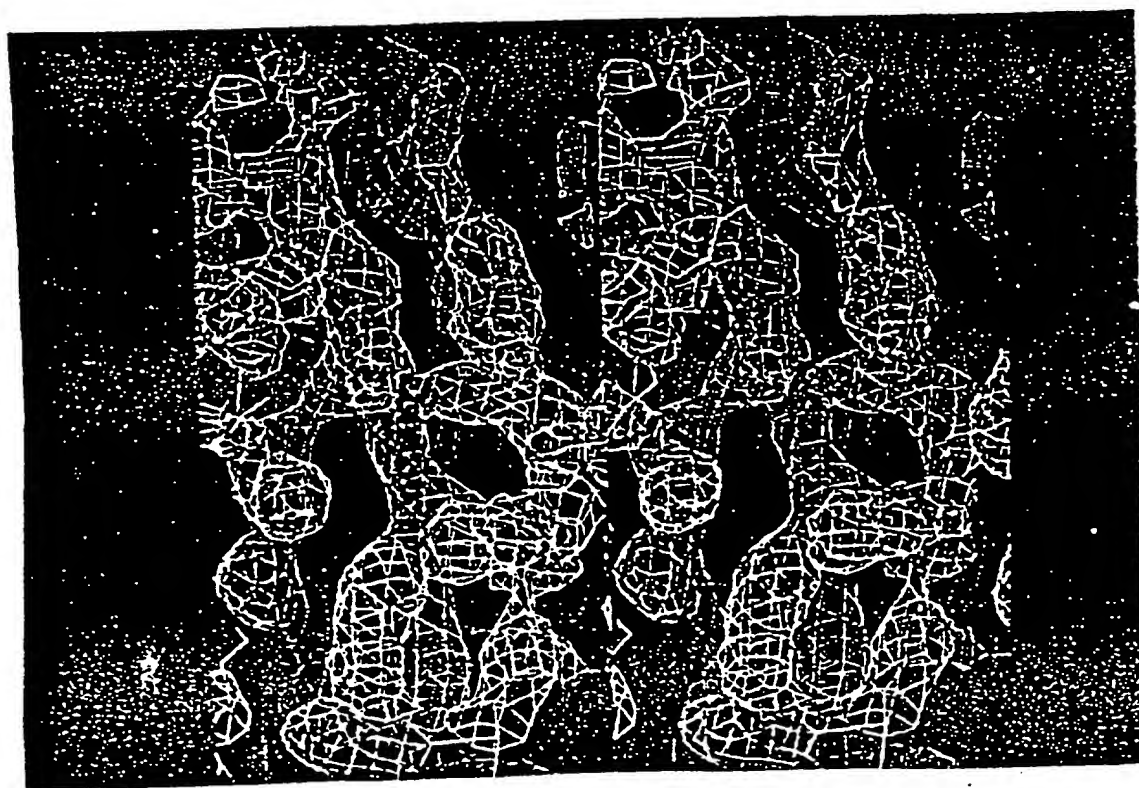
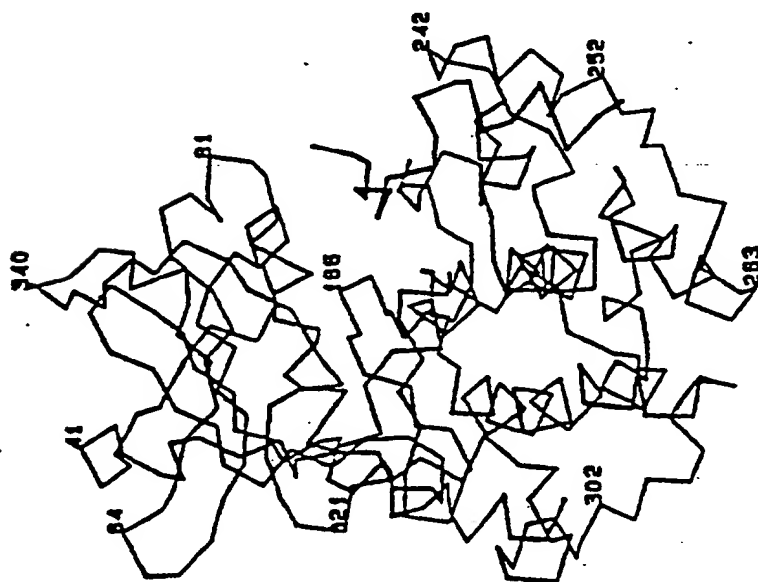
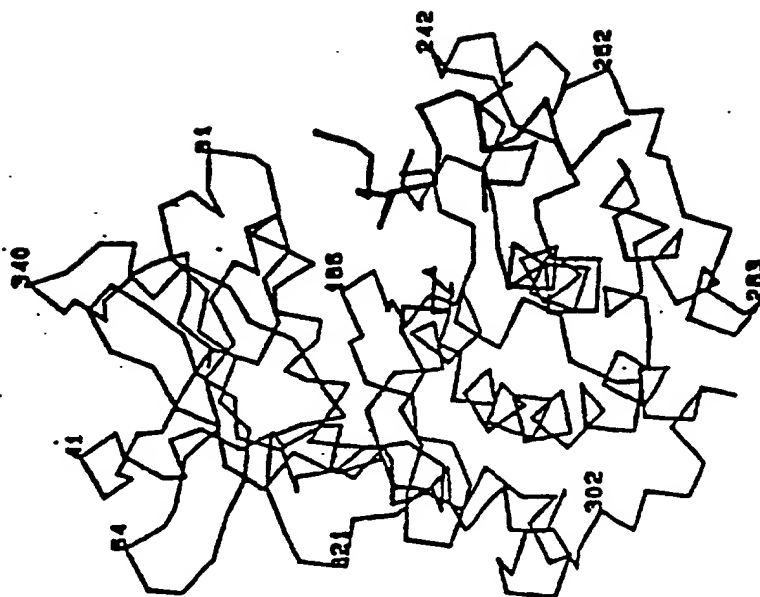


FIGURE 3B

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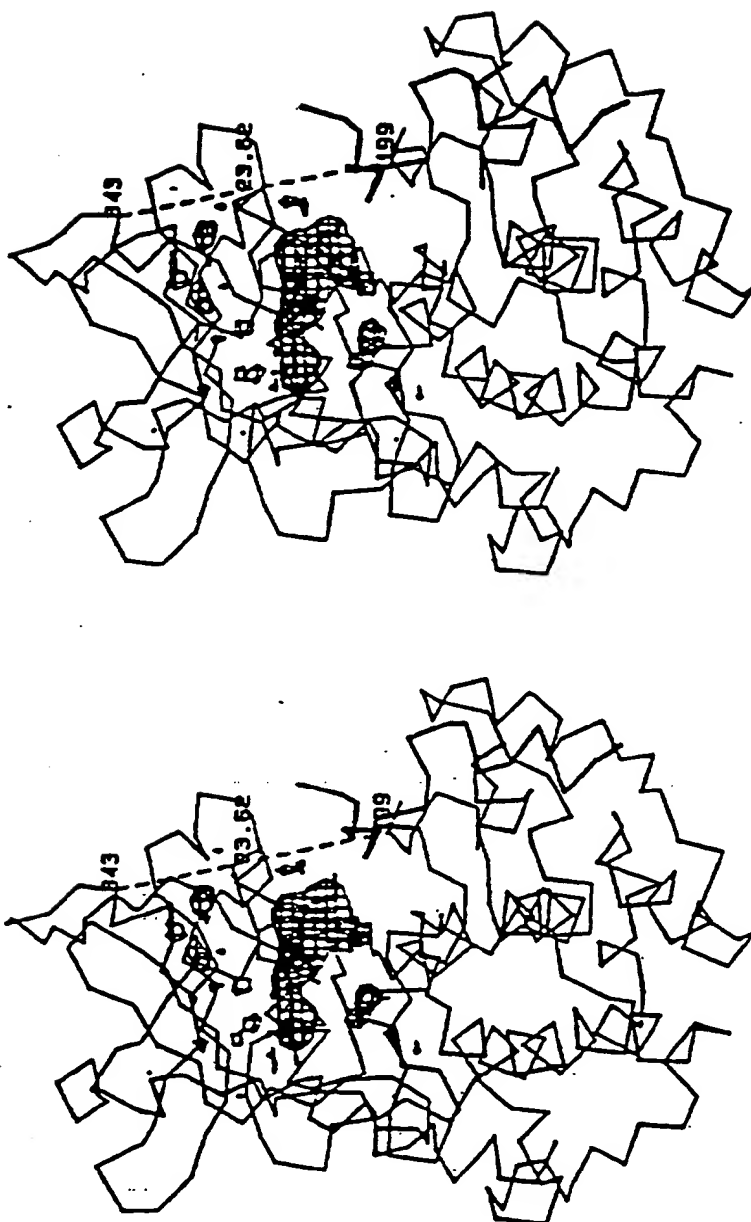
4

FIGURE



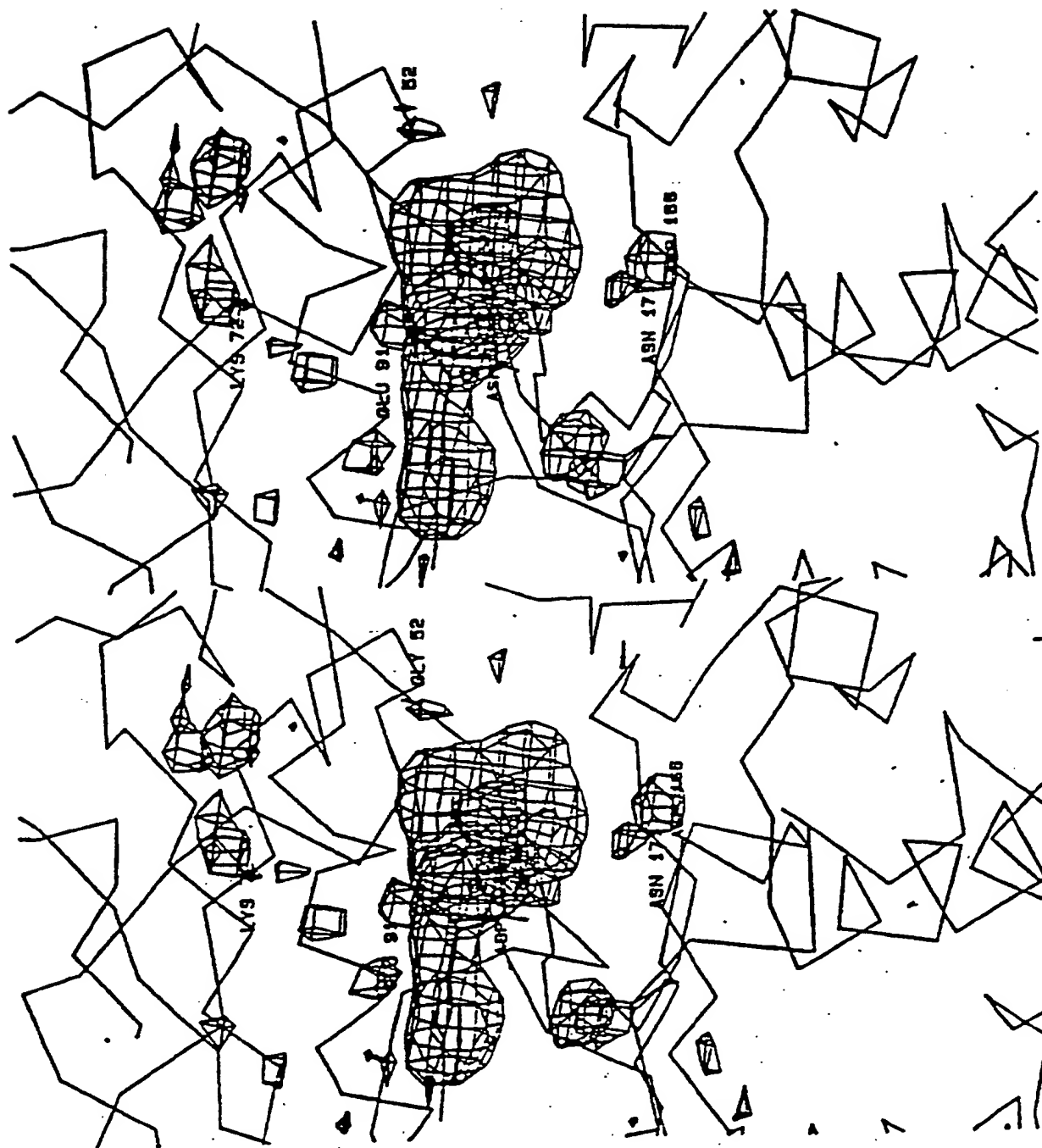
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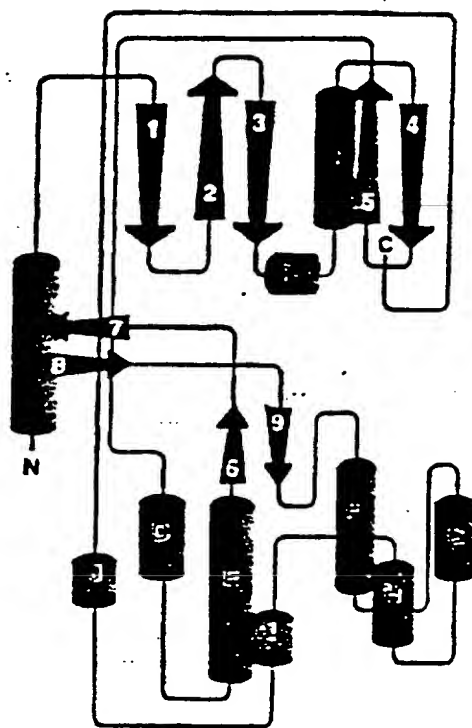
FIGURE 5A



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FIGURE 5B





FIGURE

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FIGURE 7B

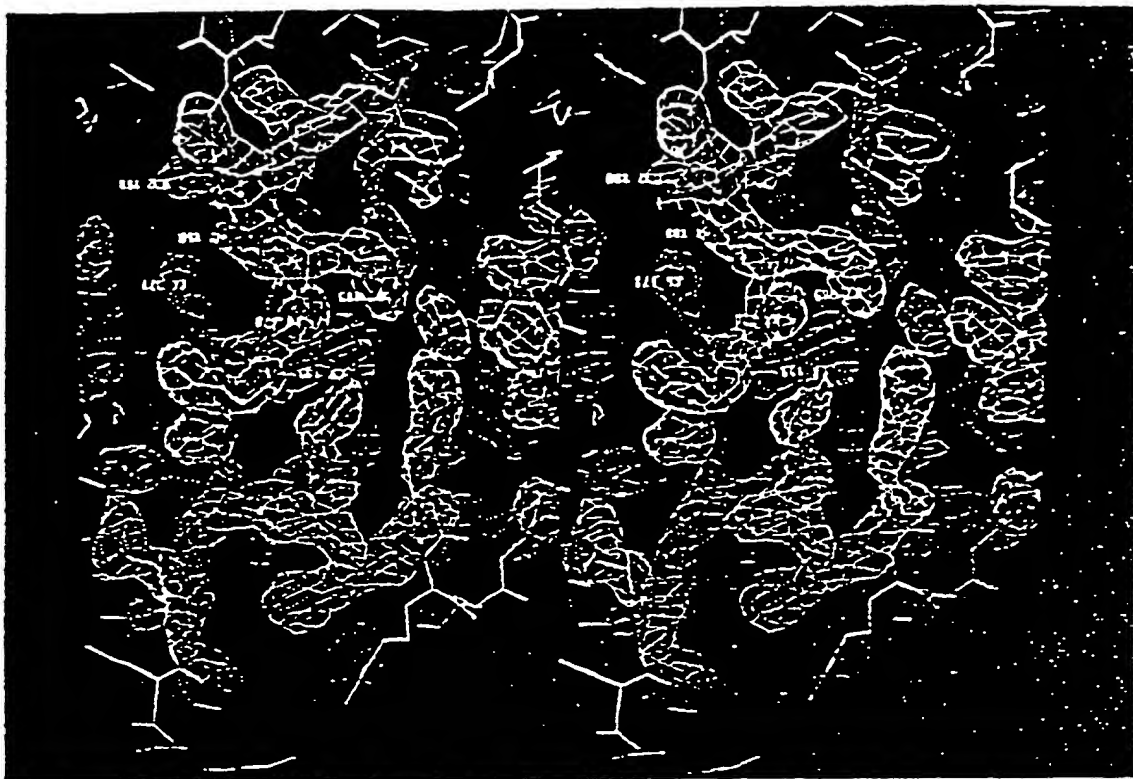
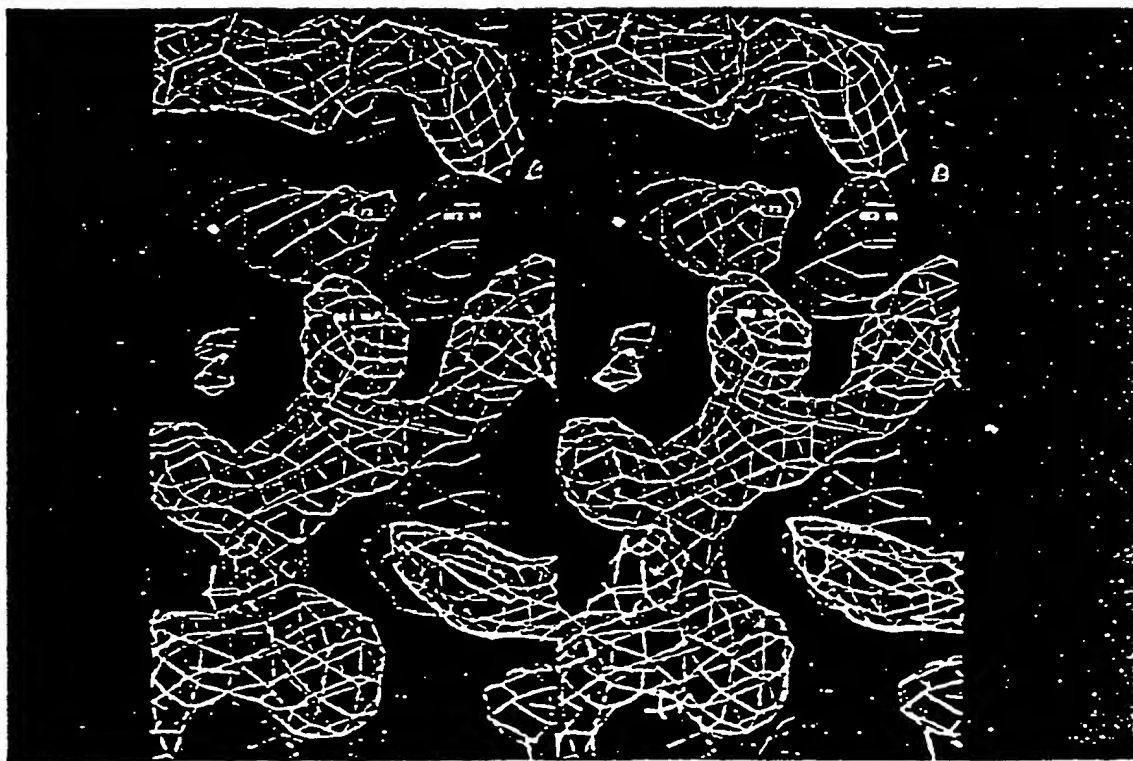
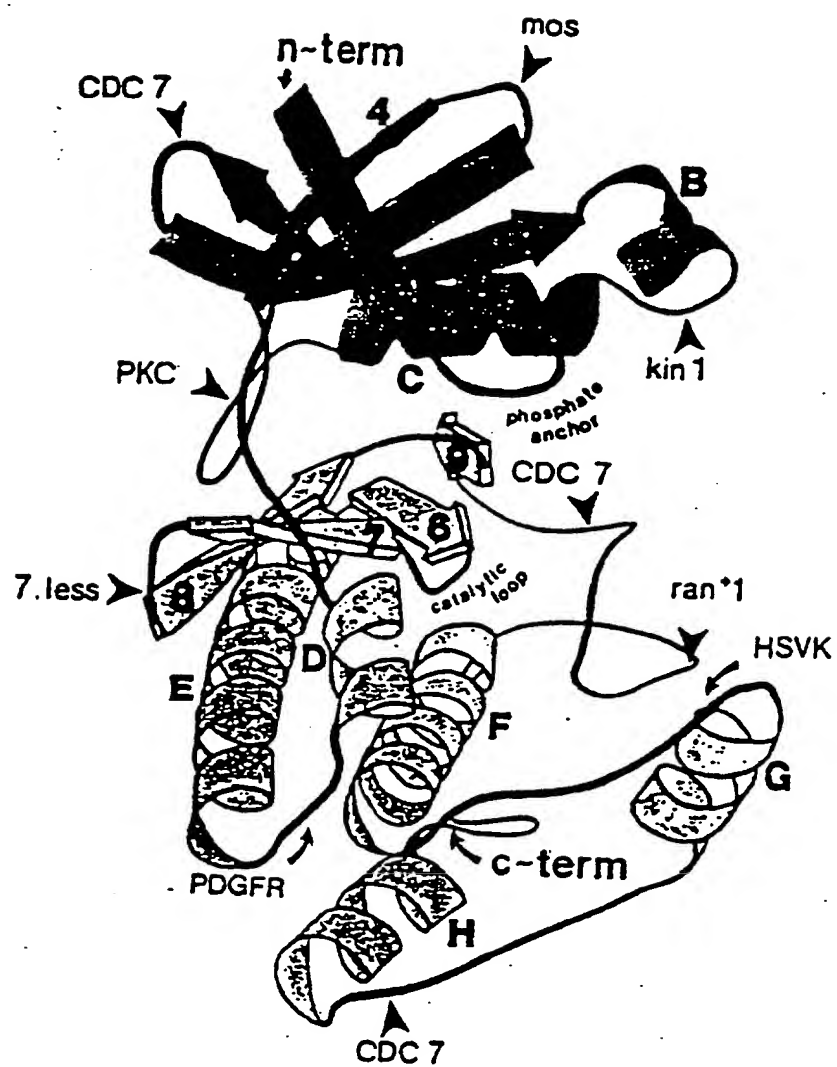


FIGURE 7A



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FIGURE

8B

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FIGURE 8A

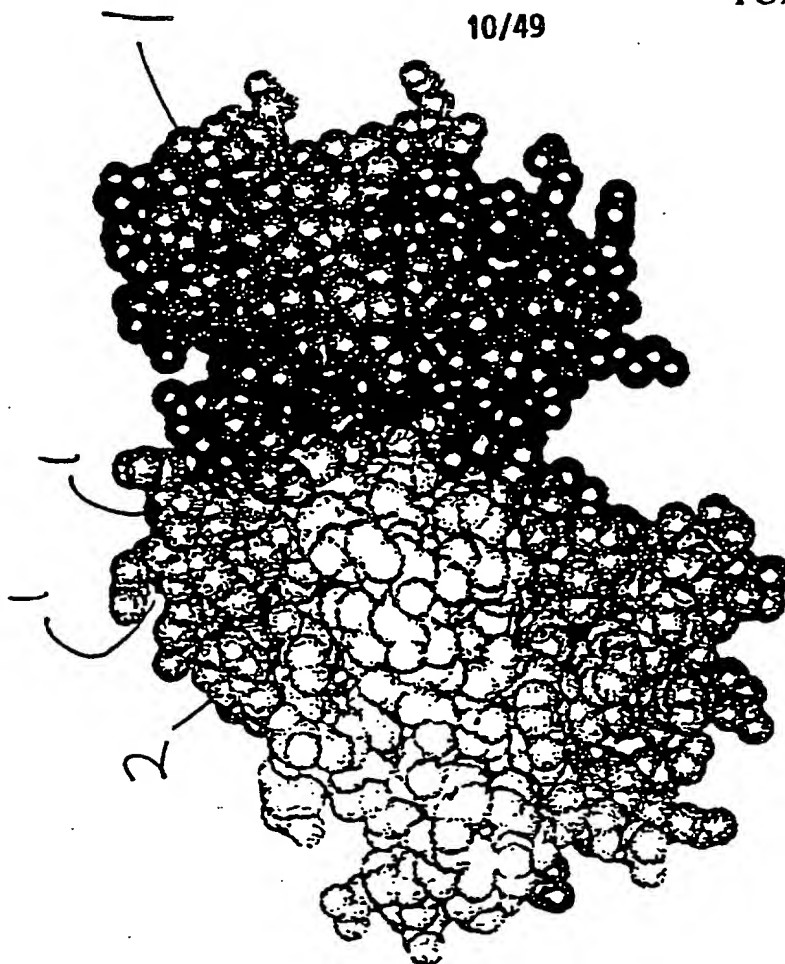
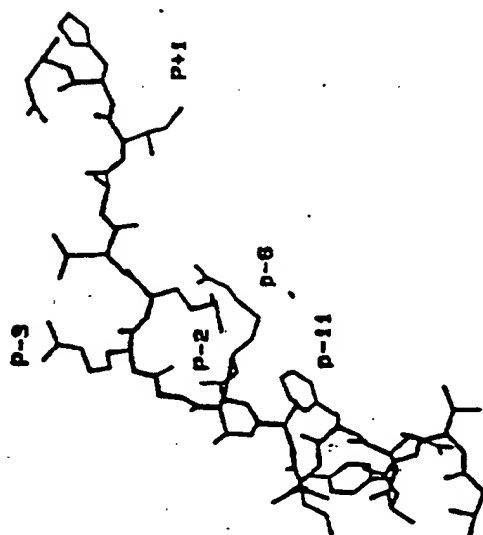
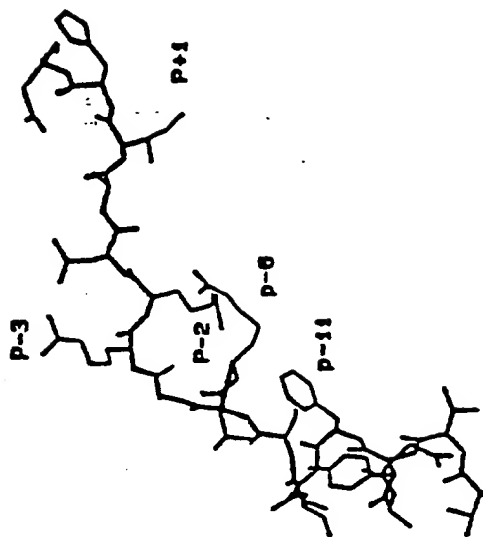


FIGURE 8C



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FIGURE 9



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FIGURE 10A

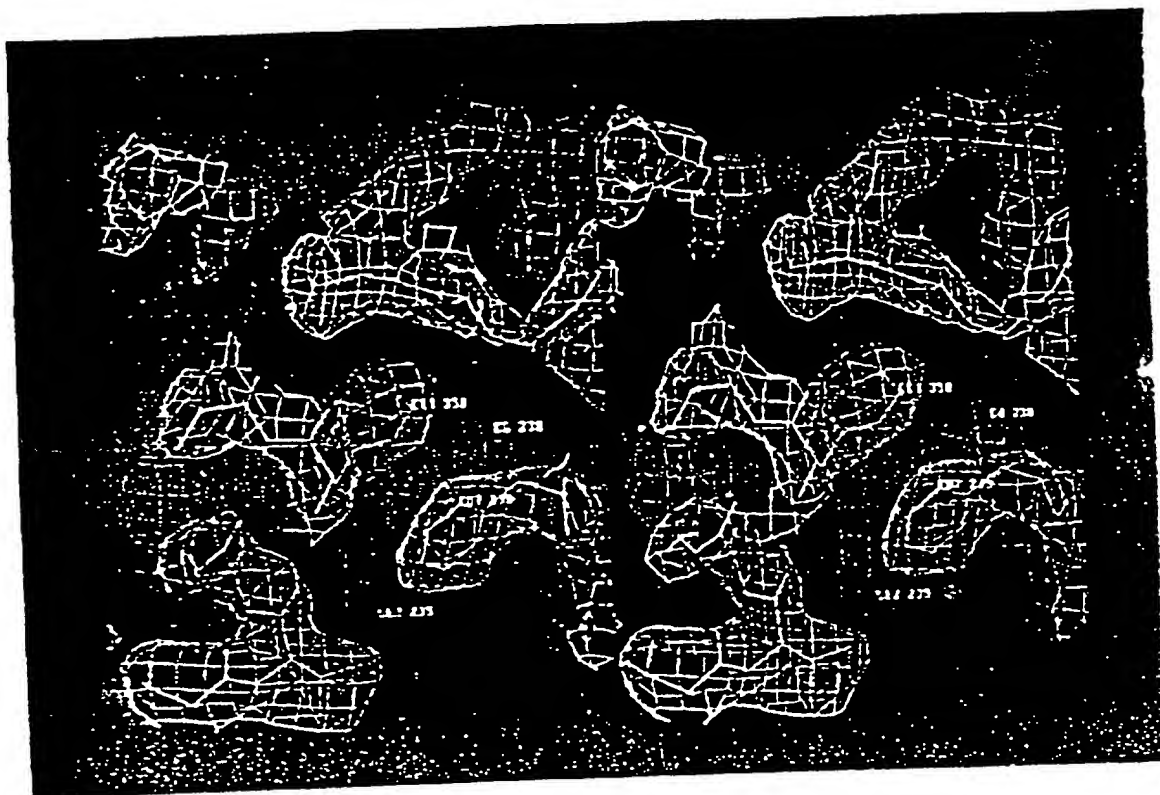
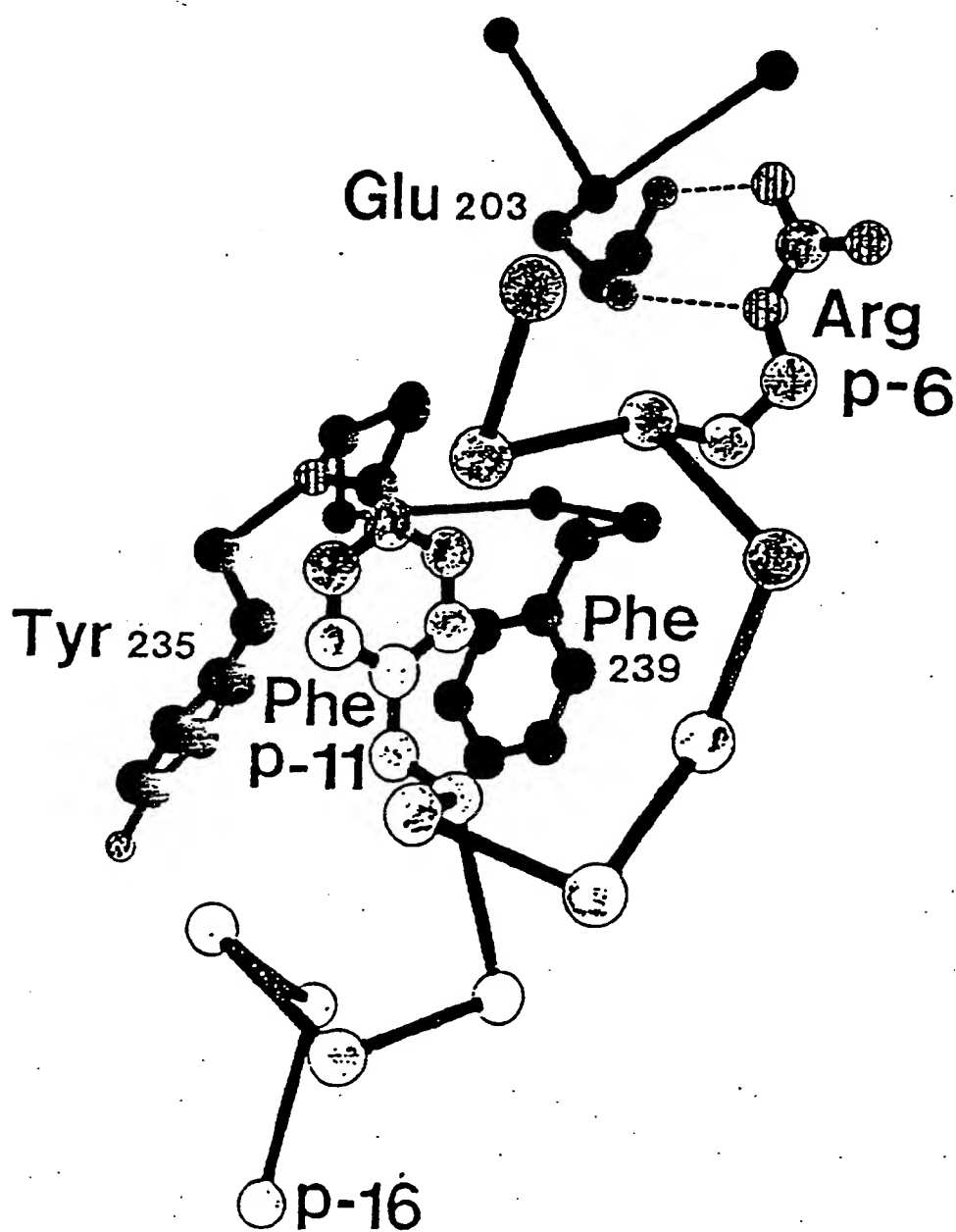


FIGURE 10B



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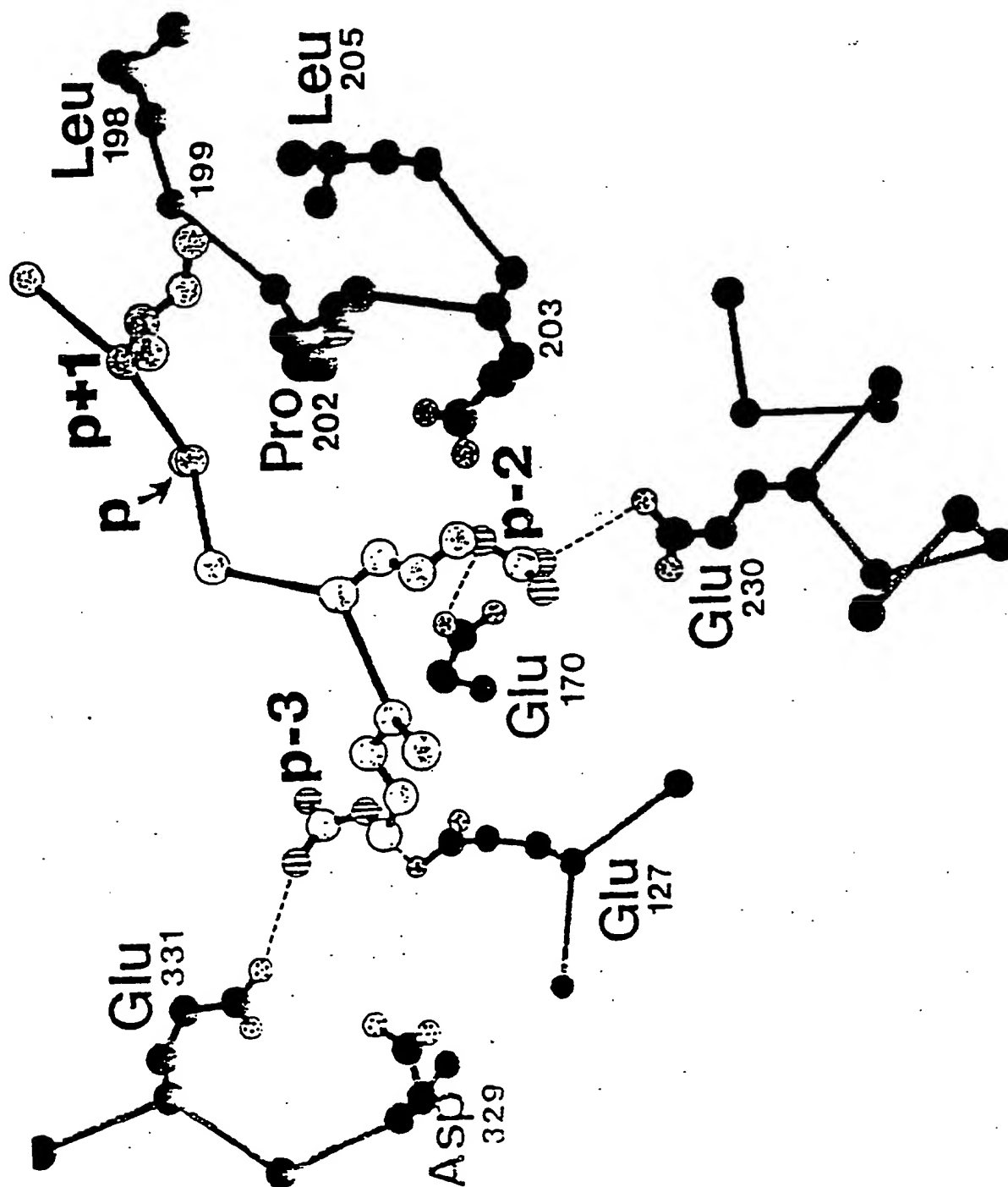


FIGURE 11

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FIGURE 12B

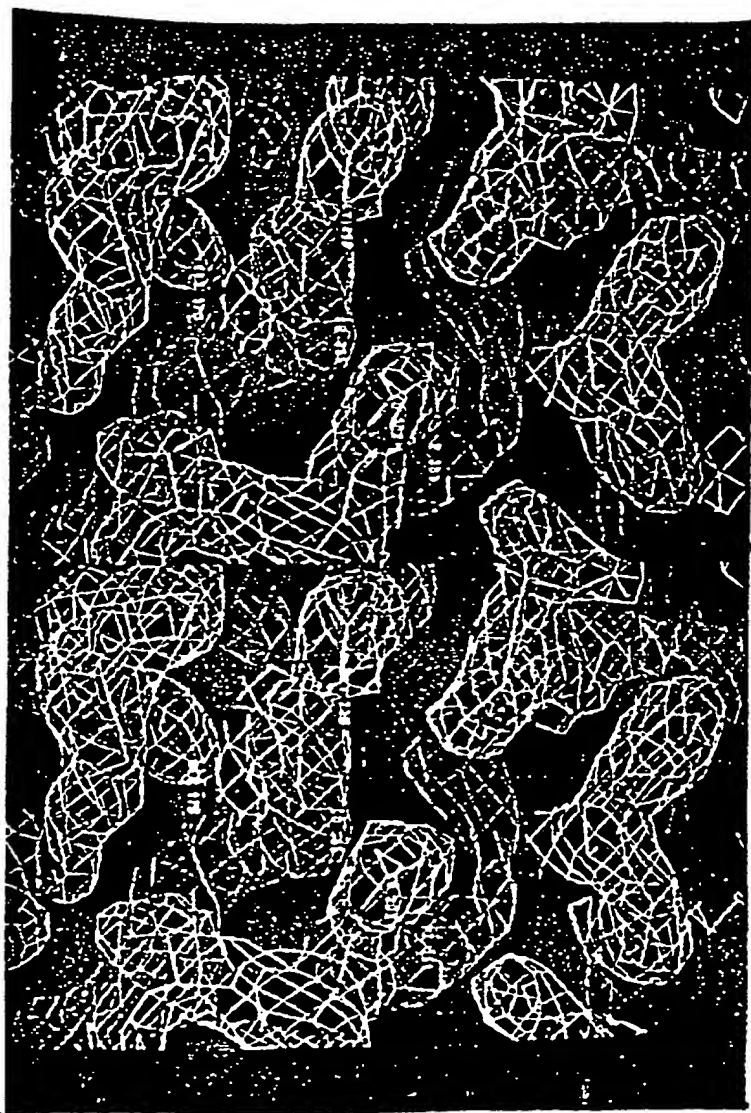
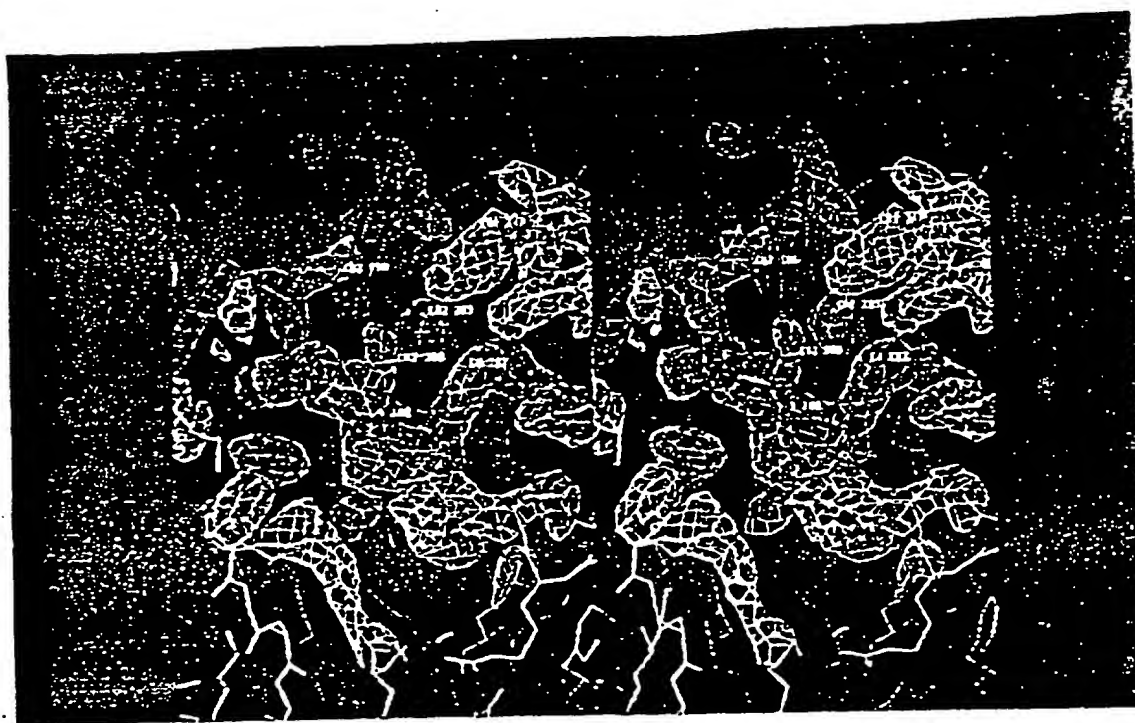


FIGURE 12A



FIGURE 12C



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FIGURE 13A

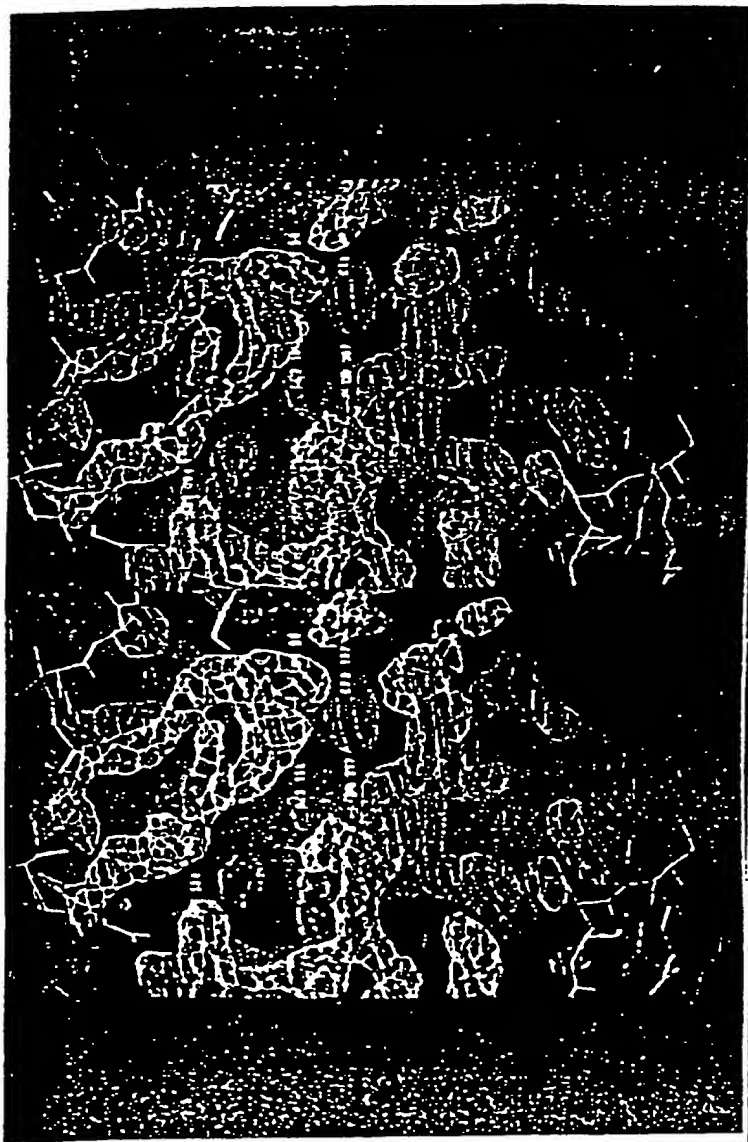
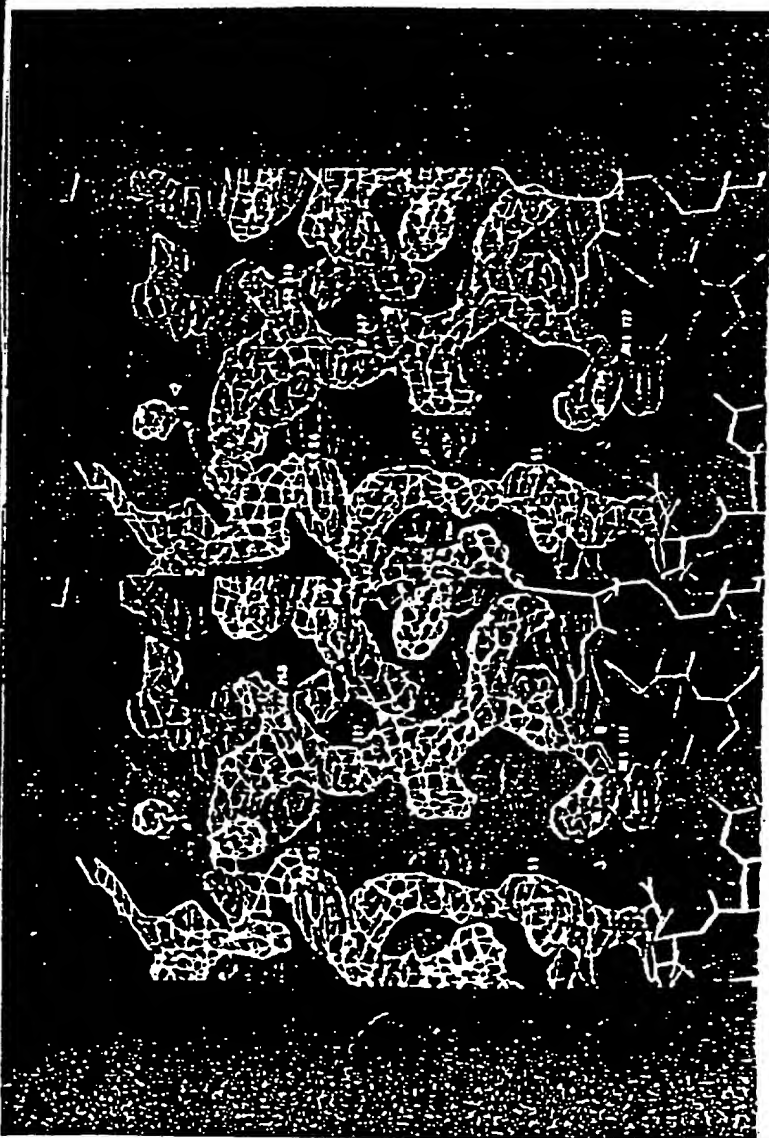


FIGURE 13B



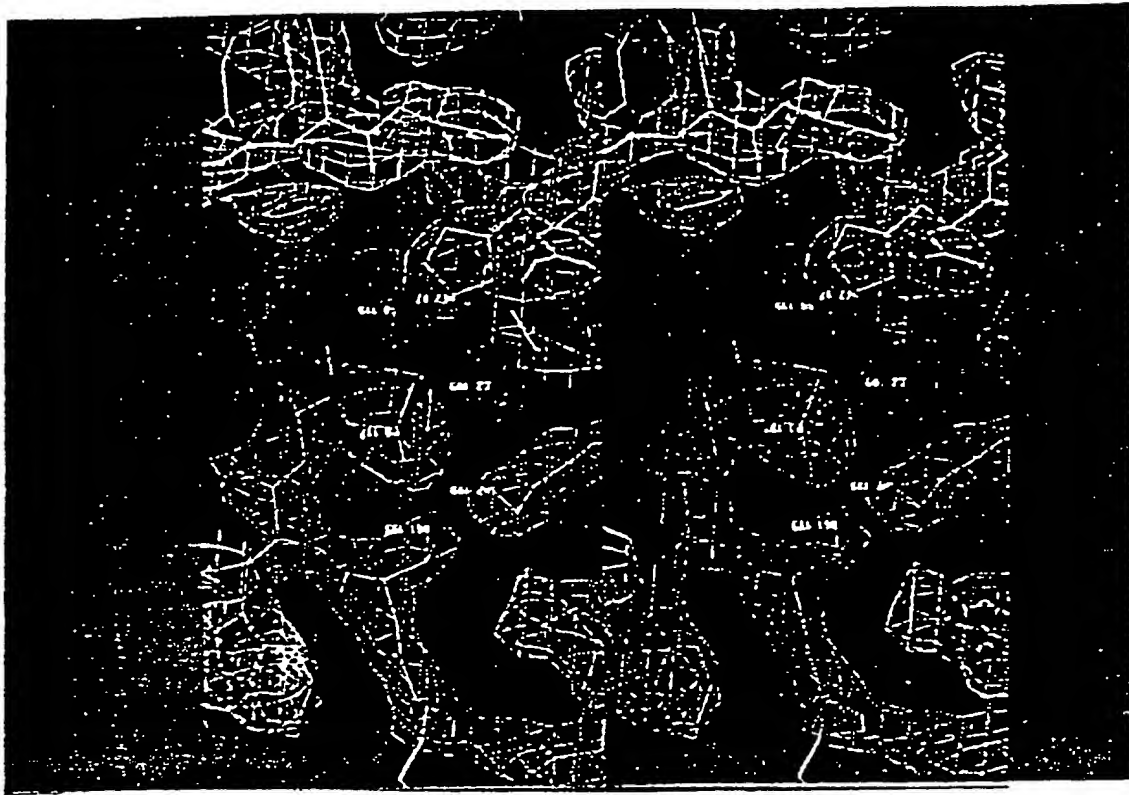
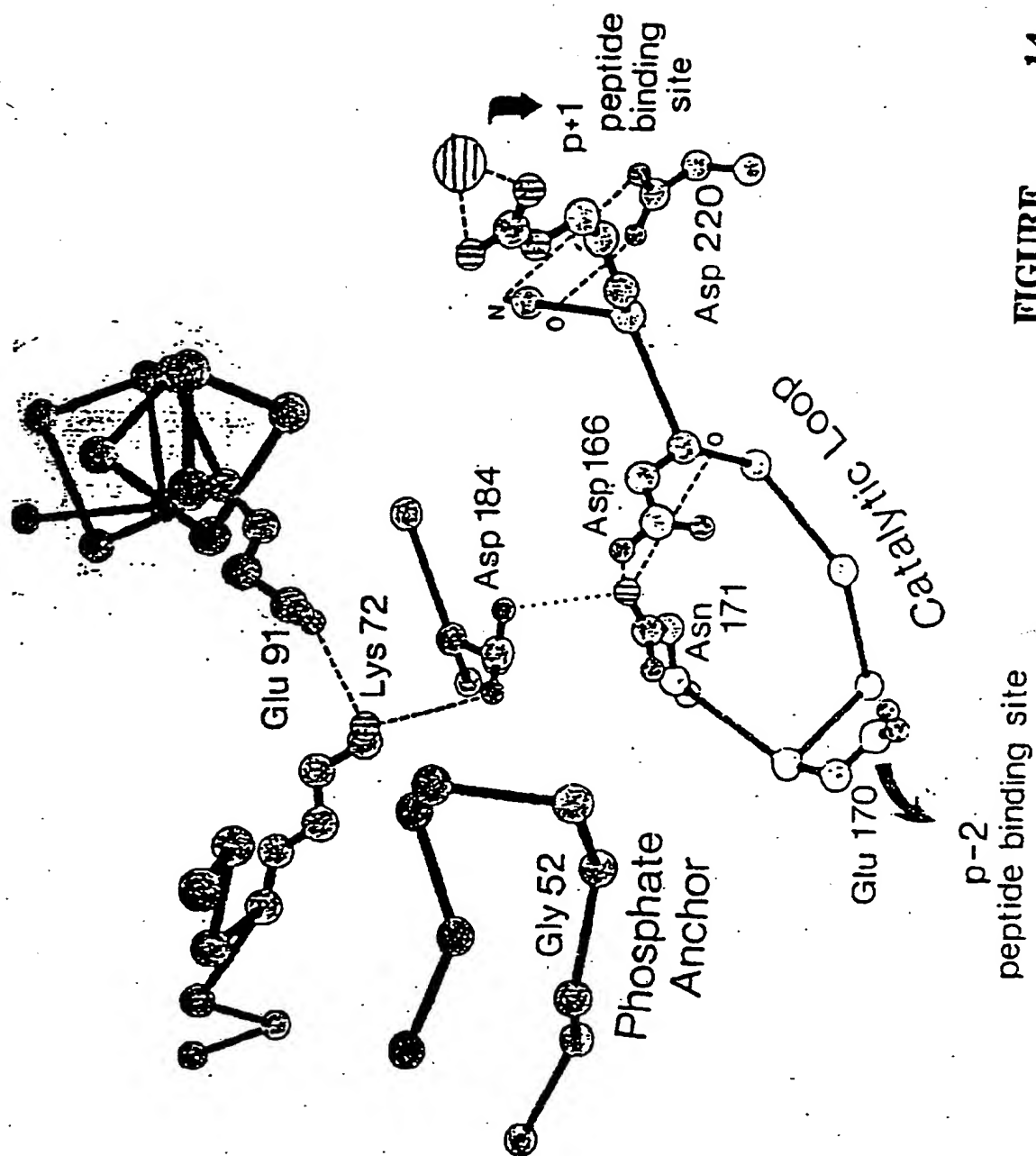


FIGURE 13C



FIGURE

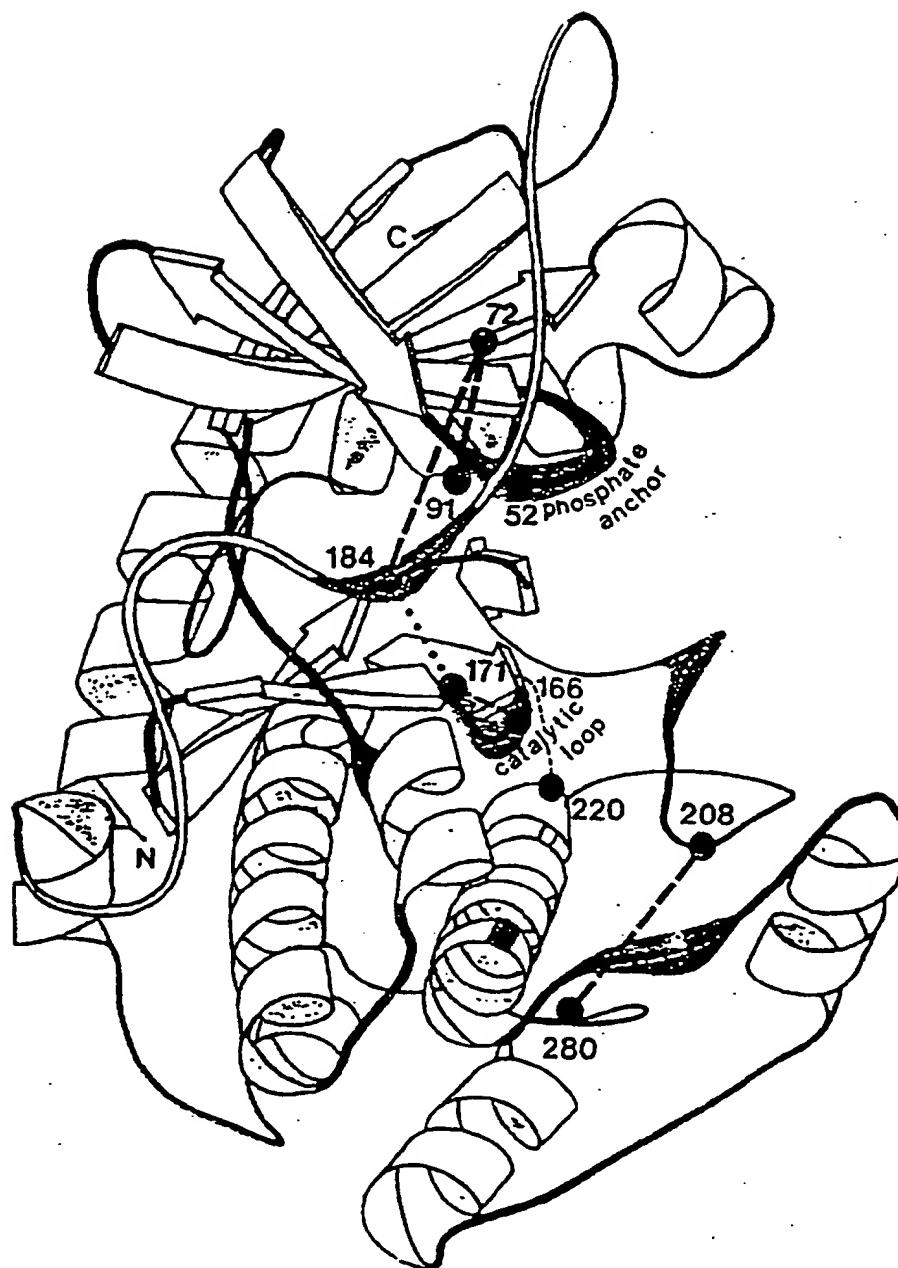
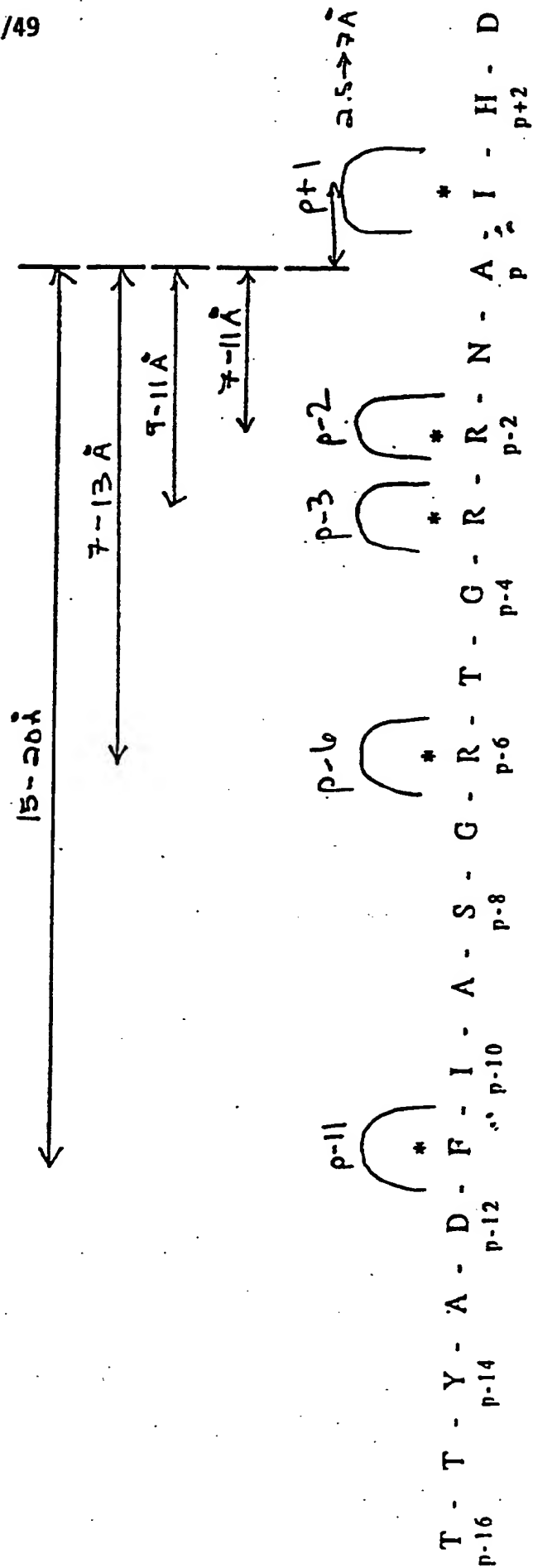


FIGURE 15

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FIGURE 16

PKI(5-24)



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17

FIGURE

diastere of Catalytic Subunit of cAMP-dependent Protein Kinase (Residues 35-370) Completed with PKI(5-24) (Residues 351-370)					
1	N	VAL	15	1	ALA
1	CA	VAL	15	22	22
2	CA	VAL	15	23	23
3	CB	VAL	15	23	23
4	CG1	VAL	15	23	23
5	CG2	VAL	15	23	23
6	C	VAL	15	23	23
7	VAL	15	15	23	23
8	N	LYS	16	24	24
9	CA	LYS	16	24	24
10	CB	LYS	16	24	24
11	CG	LYS	16	24	24
12	CD	LYS	16	24	24
13	CE	LYS	16	24	24
14	N2	LYS	16	24	24
15	C	LYS	16	24	24
16	O	LYS	16	24	24
17	N	GLU	17	25	25
18	CA	GLU	17	25	25
19	CB	GLU	17	25	25
20	CG	GLU	17	25	25
21	CD	GLU	17	25	25
22	CE	GLU	17	25	25
23	O2	GLU	17	25	25
24	C	GLU	17	25	25
25	N	ASP	18	26	26
26	H	PHE	18	26	26
27	CA	PHE	18	26	26
28	CB	PHE	18	26	26
29	CG	PHE	18	26	26
30	CD	PHE	18	26	26
31	CE	PHE	18	26	26
32	C	PHE	18	26	26
33	O	PHE	18	26	26
34	N	LEU	19	27	27
35	CA	LEU	19	27	27
36	CB	LEU	19	27	27
37	CG	LEU	19	27	27
38	CD	LEU	19	27	27
39	CE	LEU	19	27	27
40	C	LEU	19	27	27
41	CD	LEU	19	27	27
42	CD	LEU	19	27	27
43	C	LEU	19	27	27
44	N	LYS	20	28	28
45	H	LYS	20	28	28
46	CA	LYS	20	28	28
47	CB	LYS	20	28	28
48	C	LYS	20	28	28
49	O	LYS	20	28	28
50	N	LYS	21	29	29
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52	CB	LYS	21	29	29
53	CG	LYS	21	29	29
54	CD	LYS	21	29	29
55	CE	LYS	21	29	29
56	N2	LYS	21	29	29
57	C	LYS	21	29	29
58	O	LYS	21	29	29
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60	CA	LYS	22	30	30
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63	O	LYS	22	30	30
64	N	LYS	23	31	31
65	CA	LYS	23	31	31
66	CB	LYS	23	31	31
67	CG	LYS	23	31	31
68	CD	LYS	23	31	31
69	CE	LYS	23	31	31
70	N2	LYS	23	31	31
71	C	LYS	23	31	31
72	O	LYS	23	31	31
73	N	GLU	24	32	32
74	CA	GLU	24	32	32
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76	CG	GLU	24	32	32
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84	CB	ASP	25	33	33
85	CG	ASP	25	33	33
86	CD	ASP	25	33	33
87	CE	ASP	25	33	33
88	C	ASP	25	33	33
89	O	ASP	25	33	33
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107	C	LEU	27	35	35
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110	CA	LYS	28	36	36
111	CB	LYS	28	36	36
112	CG	LYS	28	36	36
113	CD	LYS	28	36	36
114	CE	LYS	28	36	36
115	N2	LYS	28	36	36
116	C	LYS	28	36	36
117	O	LYS	28	36	36
118	N	LYS	29	37	37
119	CA	LYS	29	37	37
120	CB	LYS	29	37	37
121	CG	LYS	29	37	37
122	CD	LYS	29	37	37
123	CE	LYS	29	37	37
124	N2	LYS	29	37	37
125	C	LYS	29	37	37
126	O	LYS	29	37	37
127	N	LYS	30	38	38
128	CA	LYS	30	38	38
129	CB	LYS	30	38	38
130	C	LYS	30	38	38
131	O	LYS	30	38	38
132	N	LYS	31	39	39
133	CA	LYS	31	39	39
134	CB	LYS	31	39	39
135	CG	LYS	31	39	39
136	CD	LYS	31	39	39
137	CE	LYS	31	39	39
138	C	LYS	31	39	39
139	O	LYS	31	39	39
140	N	LYS	32	40	40
141	CA	LYS	32	40	40
142	CB	LYS	32	40	40
143	CG	LYS	32	40	40
144	CD	LYS	32	40	40
145	CE	LYS	32	40	40
146	C	LYS	32	40	40
147	O	LYS	32	40	40
148	N	LYS	33	41	41
149	CA	LYS	33	41	41
150	CB	LYS	33	41	41
151	CG	LYS	33	41	41
152	CD	LYS	33	41	41
153	CE	LYS	33	41	41
154	C	LYS	33	41	41
155	O	LYS	33	41	41
156	N	LYS	34	42	42
157	CA	LYS	34	42	42
158	CB	LYS	34	42	42
159	CG	LYS	34	42	42
160	CD	LYS	34	42	42
161	CE	LYS	34	42	42
162	C	LYS	34	42	42
163	O	LYS	34	42	42
164	N	LYS	35	43	43
165	CA	LYS	35	43	43
166	CB	LYS	35	43	43
167	CG	LYS	35	43	43
168	CD	LYS	35	43	43
169	CE	LYS	35	43	43
170	C	LYS	35	43	43
171	O	LYS	35	43	43
172	N	LYS	36	44	44
173	CA	LYS	36	44	44
174	CB	LYS	36	44	44
175	CG	LYS	36	44	44
176	CD	LYS	36	44	44
177	CE	LYS	36	44	44
178	C	LYS	36	44	44
179	O	LYS	36	44	44
180	N	LYS	37	45	45
181	CA	LYS	37	45	45
182	CB	LYS	37	45	45
183	CG	LYS	37	45	45
184	CD	LYS	37	45	45
185	CE	LYS	37	45	45
186	C	LYS	37	45	45
187	O	LYS	37	45	45
188	N	LYS	38	46	46
189	CA	LYS	38	46	46
190	CB	LYS	38	46	46
191	CG	LYS	38	46	46
192	CD	LYS	38	46	46
193	CE	LYS	38	46	46
194	C	LYS	38	46	46
195	O	LYS	38	46	46
196	N	LYS	39	47	47
197	CA	LYS	39	47	47
198	CB	LYS	39	47	47
199	CG	LYS	39	47	47
200	CD	LYS	39	47	47
201	CE	LYS	39	47	47
202	C	LYS	39	47	47
203	O	LYS	39	47	47
204	N	LYS	40	48	48
205	CA	LYS	40	48	48
206	CB	LYS	40	48	48
207	CG	LYS	40	48	48
208	CD	LYS	40	48	48
209	CE	LYS	40	48	48
210	C	LYS	40	48	48
211	O	LYS	40	48	48
212	N	LYS	41	49	49
213	CA	LYS	41	49	49
214	CB	LYS	41	49	49
215	CG	LYS	41	49	49
216	CD	LYS	41	49	49
217	CE	LYS	41	49	49
218	C	LYS	41	49	49
219	O	LYS	41	49	49
220	N	LYS	42	50	50
221	CA	LYS	42	50	50
222	CB	LYS	42	50	50
223	CG	LYS	42	50	50
224	CD	LYS	42	50	50
225	CE	LYS	42	50	50
226	C	LYS	42	50	50
227	O	LYS	42	50	50
228	N	LYS	43	51	51
229	CA	LYS	43	51	51
230	CB	LYS	43	51	51
231	CG	LYS	43	51	51
232	CD	LYS	43	51	51
233	CE	LYS	43	51	51
234	C	LYS	43	51	51
235	O	LYS	43	51	51
236	N	LYS	44	52	52
237	CA	LYS	44	52	52
238	CB	LYS	44	52	52
239	CG	LYS	44	52	52
240	CD	LYS	44	52	52
241	CE	LYS	44	52	52
242	C	LYS	44	52	52
243	O	LYS	44	52	52
244	N	LYS	45	53	53
245	CA	LYS	45	53	53
246	CB	LYS	45	53	53
247	CG	LYS	45	53	53
248	CD	LYS	45	53	53
249	CE	LYS	45	53	53
250	C	LYS	45	53	53
251	O	LYS	45	53	53
252	N	LYS	46	54	54
253	CA	LYS	46	54	54
254	CB	LYS	46	54	54
255	CG	LYS	46	54	54
256	CD	LYS	46	54	54
257	CE	LYS	46	54	54
258	C	LYS	46	54	54
259	O	LYS	46	54	54
260					

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128	CA	TRP	3.608	8.431	-19.342	3.00	17.60	ATOM	193	C	THR	37	-3.725	22.256	-9.221	1.00	17.60
129	CB	TRP	3.945	7.029	-19.784	3.00	17.60	ATOM	194	O	THR	37	-2.481	23.203	-9.315	1.00	17.60
130	CD	TRP	3.397	6.884	-20.180	3.00	17.60	ATOM	195	N	ALA	38	-1.009	22.080	-8.107	1.00	17.60
131	CE	TRP	4.475	6.972	-19.343	3.00	17.60	ATOM	196	CA	ALA	38	-1.278	22.965	-6.989	1.00	17.60
132	CF	TRP	5.509	6.989	-20.757	3.00	17.60	ATOM	197	CB	ALA	38	-1.037	22.231	-5.708	1.00	17.60
133	CG	TRP	4.737	7.062	-18.000	3.00	17.60	ATOM	198	C	ALA	38	-0.460	24.243	-6.977	1.00	17.60
134	CH	TRP	3.726	6.803	-21.197	3.00	17.60	ATOM	199	O	ALA	38	0.331	24.564	-7.997	1.00	17.60
135	CI	TRP	3.023	6.895	-21.197	3.00	17.60	ATOM	200	N	GLN	39	-0.399	25.033	-5.897	1.00	17.60
136	CJ	TRP	6.828	7.111	-19.965	3.00	17.60	ATOM	201	CA	GLN	39	0.532	26.157	-5.813	1.00	17.60
137	CK	TRP	6.056	7.182	-17.587	3.00	17.60	ATOM	202	CB	GLN	39	-0.371	27.351	-6.332	1.00	17.60
138	CL	TRP	7.089	7.232	-18.303	3.00	17.60	ATOM	203	CD	GLN	39	0.431	28.749	-6.308	1.00	17.60
139	CM	TRP	3.993	9.388	-20.420	3.00	17.60	ATOM	204	CE	GLN	39	-6.491	1.00	17.60		
140	CN	TRP	3.107	9.048	-20.501	3.00	17.60	ATOM	205	CE	GLN	39	-1.061	30.597	-5.654	1.00	17.60
141	CO	TRP	1.056	8.587	-21.311	3.00	17.60	ATOM	206	CE	GLN	39	-3.418	29.563	-7.636	1.00	17.60
142	CP	TRP	1.262	10.417	-22.451	3.00	17.60	ATOM	207	C	GLN	39	0.904	26.284	-4.331	1.00	17.60
143	CQ	TRP	0.404	9.816	-23.698	3.00	17.60	ATOM	208	O	GLN	39	0.064	26.148	-3.429	1.00	17.60
144	CR	TRP	3.180	8.804	-24.289	3.00	17.60	ATOM	209	N	LEU	40	2.184	26.560	-4.057	1.00	17.60
145	CS	TRP	0.420	7.977	-25.323	3.00	17.60	ATOM	210	CA	LEU	40	2.791	26.451	-2.926	1.00	17.60
146	CT	TRP	0.071	8.531	-26.309	3.00	17.60	ATOM	211	CB	LEU	40	4.200	27.082	-2.926	1.00	17.60
147	CU	TRP	0.234	6.767	-25.038	3.00	17.60	ATOM	212	CG	LEU	40	5.404	26.933	-2.007	1.00	17.60
148	CV	TRP	0.977	11.856	-22.225	3.00	17.60	ATOM	213	CD	LEU	40	5.689	28.318	-1.462	1.00	17.60
149	CW	TRP	1.617	12.709	-22.862	3.00	17.60	ATOM	214	CE	LEU	40	5.196	25.882	-0.934	1.00	17.60
150	CX	TRP	-0.020	12.135	-23.374	3.00	17.60	ATOM	215	C	LEU	40	2.027	26.986	-3.454	1.00	17.60
151	CA	ASP	-0.380	13.519	-23.130	3.00	17.60	ATOM	216	O	LEU	40	1.587	26.781	-0.549	1.00	17.60
152	CB	ASP	-2.383	13.211	-22.630	3.00	17.60	ATOM	217	N	ASP	41	1.719	28.259	-1.672	1.00	17.60
153	CC	ASP	-1.715	13.229	-23.685	3.00	17.60	ATOM	218	CA	ASP	41	1.042	29.206	-0.839	1.00	17.60
154	CD	ASP	-3.513	12.686	-22.528	3.00	17.60	ATOM	219	CB	ASP	41	0.821	30.636	-1.716	1.00	17.60
155	CE	ASP	-0.224	13.515	-19.619	3.00	17.60	ATOM	220	CD	ASP	41	-0.459	30.564	-2.545	1.00	17.60
156	C	ASP	-1.090	13.026	-18.936	3.00	17.60	ATOM	221	OD	ASP	41	-0.940	31.725	-2.738	1.00	17.60
157	O	ASP	0.963	13.902	-19.215	3.00	17.60	ATOM	222	OD	ASP	41	-3.104	29.553	-2.975	1.00	17.60
158	N	PRO	3.250	13.686	-19.880	3.00	17.60	ATOM	223	C	ASP	41	-0.253	28.734	-0.247	1.00	17.60
159	CD	PRO	3.194	14.234	-17.839	3.00	17.60	ATOM	224	O	ASP	41	-0.808	29.339	0.671	1.00	17.60
160	CA	PRO	2.739	14.398	-17.767	3.00	17.60	ATOM	225	N	GLN	42	-0.807	27.741	-0.880	1.00	17.60
161	CB	PRO	3.186	14.611	-19.154	3.00	17.60	ATOM	226	CA	GLN	42	-2.056	27.326	-0.374	1.00	17.60
162	CC	PRO	0.521	15.528	-17.360	3.00	17.60	ATOM	227	CB	GLN	42	-2.919	27.129	-1.577	1.00	17.60
163	C	PRO	0.012	16.322	-18.173	3.00	17.60	ATOM	228	CD	GLN	42	-2.650	25.992	-2.532	1.00	17.60
164	O	PRO	0.535	15.659	-16.013	3.00	17.60	ATOM	229	CE	GLN	42	-3.803	26.370	-3.970	1.00	17.60
165	N	SCR	-0.072	16.432	-15.310	3.00	17.60	ATOM	230	OD	GLN	42	-2.448	27.439	-4.418	1.00	17.60
166	CA	SCR	-1.502	15.451	-14.263	3.00	17.60	ATOM	231	NE	GLN	42	-3.557	25.530	-6.782	1.00	17.60
167	CB	SCR	1.234	17.806	-15.065	3.00	17.60	ATOM	232	C	GLN	42	-2.081	26.115	0.529	1.00	17.60
168	CG	SCR	2.317	17.445	-14.841	3.00	17.60	ATOM	233	O	GLN	42	-3.041	25.511	0.623	1.00	17.60
169	C	SCR	0.863	19.075	-15.089	3.00	17.60	ATOM	234	N	PHE	43	-0.796	25.784	0.981	1.00	17.60
170	O	SCR	1.832	20.127	-14.999	3.00	17.60	ATOM	235	CA	PHE	43	-0.521	24.696	1.081	1.00	17.60
171	N	GLN	1.510	21.138	-16.155	3.00	17.60	ATOM	236	CB	PHE	43	0.476	23.683	1.316	1.00	17.60
172	CA	GLN	2.298	21.021	-17.486	3.00	17.60	ATOM	237	CD	PHE	43	-0.091	22.086	0.147	1.00	17.60
173	CB	GLN	3.828	21.101	-17.327	3.00	17.60	ATOM	238	CD	PHE	43	-0.618	21.783	0.189	1.00	17.60
174	CG	GLN	4.370	20.435	-18.064	3.00	17.60	ATOM	239	CD	PHE	43	0.010	23.337	-1.126	1.00	17.60
175	CD	GLN	4.389	21.893	-16.433	3.00	17.60	ATOM	240	CE	PHE	43	-1.438	21.143	-0.646	1.00	17.60
176	CE	GLN	1.712	20.757	-13.592	3.00	17.60	ATOM	241	CE	PHE	43	-0.508	22.676	-2.157	1.00	17.60
177	CC	GLN	2.032	20.065	-12.593	3.00	17.60	ATOM	242	C	PHE	43	-1.271	21.585	-1.922	1.00	17.60
178	C	GLN	1.362	22.041	-13.470	3.00	17.60	ATOM	243	C	PHE	43	-0.358	25.400	2.984	1.00	17.60
179	O	GLN	1.316	22.717	-12.188	3.00	17.60	ATOM	244	O	PHE	43	1.018	26.258	2.767	1.00	17.60
180	N	ASN	1.963	24.116	-12.270	3.00	17.60	ATOM	245	N	ASP	44	-0.285	25.003	4.166	1.00	17.60
181	CA	ASN	6.395	23.032	-10.844	3.00	17.60	ATOM	246	CA	ASP	44	-0.297	25.522	5.195	1.00	17.60
182	CB	ASN	4.322	23.217	-12.005	3.00	17.60	ATOM	247	CB	ASP	44	-0.777	25.931	6.353	1.00	17.60
183	CG	ASN	-0.121	22.852	-11.741	3.00	17.60	ATOM	248	CD	ASP	44	-0.387	27.243	7.069	1.00	17.60
184	C	ASN	-0.050	23.796	-12.035	3.00	17.60	ATOM	249	OD	ASP	44	-0.497	27.262	8.108	1.00	17.60
185	OD	ASN	-0.429	21.720	-13.128	3.00	17.60	ATOM	250	OD	ASP	44	0.018	28.246	6.033	1.00	17.60
186	C	ASN	-1.650	21.381	-10.443	3.00	17.60	ATOM	251	C	ASP	44	1.042	24.327	5.901	1.00	17.60
187	O	ASN	-1.625	19.938	-9.989	3.00	17.60	ATOM	252	O	ASP	44	0.446	23.234	5.854	1.00	17.60
188	N	THR	-1.226	19.085	-9.071	3.00	17.60	ATOM	253	N	ARG	45	2.309	24.588	6.329	1.00	17.60
189	CA	THR	-2.393	19.553	-9.497	3.00	17.60	ATOM	254	CA	ARG	45	3.337	23.576	6.654	1.00	17.60
190	CB	THR						ATOM	255	CB	ARG	45	4.671	23.942	5.952	1.00	17.60
191	CD	THR						ATOM	256	CD	ARG	45	5.807	24.822	6.586	1.00	17.60
192	CE	THR						ATOM	257	CD	ARG	45	6.831	24.063	7.474	1.00	17.60

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OH	258	HE	ARG	45	0.189	24.601	7.476	1.00	17.60	19.985	11.398	-4.683	1.00	17.60
OH	259	CE	ARG	45	0.709	25.370	9.477	1.00	17.60	17.650	11.003	-4.910	1.00	17.60
OH	260	HE	ARG	45	0.021	25.738	9.585	1.00	17.60	18.941	10.502	-4.827	1.00	17.60
OH	261	HE	ARG	45	0.007	25.735	0.423	1.00	17.60	16.767	15.469	-2.728	1.00	17.60
OH	262	C	ARG	45	0.007	25.735	0.423	1.00	17.60	16.282	15.396	-2.728	1.00	17.60
OH	263	N	ARG	45	0.105	26.057	0.889	1.00	17.60	16.100	14.368	-2.499	1.00	17.60
OH	264	N	ARG	45	0.105	26.057	0.889	1.00	17.60	14.764	14.257	-1.950	1.00	17.60
OH	265	CA	ARG	46	0.597	21.083	0.434	1.00	17.60	14.644	15.193	-0.761	1.00	17.60
OH	266	CA	ARG	46	0.597	21.083	0.434	1.00	17.60	13.736	15.032	-0.631	1.00	17.60
OH	267	CA	ARG	46	0.821	20.312	10.090	1.00	17.60	15.627	15.015	0.099	1.00	17.60
OH	268	CG	ARG	46	0.073	20.139	11.594	1.00	17.60	15.776	15.968	1.146	1.00	17.60
OH	269	CG	ARG	46	0.690	21.965	9.979	1.00	17.60	16.515	17.149	0.499	1.00	17.60
OH	270	C	ARG	46	0.211	20.974	10.073	1.00	17.60	18.035	16.736	0.270	1.00	17.60
OH	271	N	ARG	46	0.586	21.775	10.597	1.00	17.60	18.907	16.107	1.434	1.00	17.60
OH	272	N	ARG	46	0.014	19.387	10.146	1.00	17.60	18.543	16.550	2.742	1.00	17.60
OH	273	CA	ARG	47	0.072	18.778	11.591	1.00	17.60	18.964	16.231	3.935	1.00	17.60
OH	274	CB	ARG	47	0.566	17.394	12.064	1.00	17.60	19.004	15.207	4.065	1.00	17.60
OH	275	CB	ARG	47	0.572	16.324	12.313	1.00	17.60	18.545	16.918	5.021	1.00	17.60
OH	276	CD	ARG	47	0.829	16.390	13.239	1.00	17.60	14.535	16.347	1.886	1.00	17.60
OH	277	CE	ARG	47	0.005	16.859	12.546	1.00	17.60	13.033	15.387	2.189	1.00	17.60
OH	278	CE	ARG	47	0.571	18.370	9.177	1.00	17.60	14.052	17.360	2.121	1.00	17.60
OH	279	C	ARG	47	0.837	17.729	9.425	1.00	17.60	13.050	17.719	3.182	1.00	17.60
OH	280	N	ARG	47	0.880	18.210	9.284	1.00	17.60	13.014	19.327	3.584	1.00	17.60
OH	281	N	ARG	48	0.671	17.302	8.501	1.00	17.60	12.188	20.277	2.663	1.00	17.60
OH	282	CA	ARG	48	11.026	17.995	8.610	1.00	17.60	12.022	19.422	4.949	1.00	17.60
OH	283	CB	ARG	48	10.787	19.207	8.034	1.00	17.60	11.713	17.091	2.732	1.00	17.60
OH	284	CG	ARG	48	12.152	17.122	7.984	1.00	17.60	11.420	16.901	1.534	1.00	17.60
OH	285	CG	ARG	48	9.554	15.060	8.940	1.00	17.60	10.950	16.665	3.746	1.00	17.60
OH	286	C	ARG	48	10.119	15.407	9.937	1.00	17.60	9.680	15.995	3.532	1.00	17.60
OH	287	N	ARG	49	0.679	15.197	8.233	1.00	17.60	10.129	14.616	3.236	1.00	17.60
OH	288	N	ARG	49	0.401	13.421	8.449	1.00	17.60	9.488	14.217	2.096	1.00	17.60
OH	289	CA	ARG	49	7.338	13.369	7.512	1.00	17.60	8.148	13.359	2.895	1.00	17.60
OH	290	CB	ARG	49	5.952	13.869	7.786	1.00	17.60	8.092	11.953	3.020	1.00	17.60
OH	291	CG	ARG	49	5.078	13.387	6.667	1.00	17.60	8.759	16.257	4.733	1.00	17.60
OH	292	CG	ARG	49	5.429	13.371	9.097	1.00	17.60	8.716	15.359	5.741	1.00	17.60
OH	293	CD	ARG	49	9.620	12.981	8.216	1.00	17.60	8.076	17.374	4.566	1.00	17.60
OH	294	C	ARG	49	10.776	12.123	9.052	1.00	17.60	7.269	18.051	5.512	1.00	17.60
OH	295	N	ARG	50	10.776	12.123	9.052	1.00	17.60	7.270	19.343	5.146	1.00	17.60
OH	296	N	ARG	50	11.429	12.339	7.060	1.00	17.60	8.353	20.548	4.697	1.00	17.60
OH	297	CA	ARG	50	12.442	13.364	6.695	1.00	17.60	7.624	21.728	4.178	1.00	17.60
OH	298	C	ARG	50	12.205	14.419	5.482	1.00	17.60	9.230	21.089	5.801	1.00	17.60
OH	299	N	ARG	51	13.630	12.749	5.802	1.00	17.60	5.828	17.552	5.497	1.00	17.60
OH	300	N	ARG	51	14.560	13.512	5.000	1.00	17.60	5.327	17.479	4.370	1.00	17.60
OH	301	CA	ARG	51	15.596	14.190	5.938	1.00	17.60	5.054	17.225	6.552	1.00	17.60
OH	302	CB	ARG	51	15.564	13.381	5.265	1.00	17.60	3.596	17.017	6.466	1.00	17.60
OH	303	CG	ARG	51	16.838	13.349	6.254	1.00	17.60	2.873	16.300	7.754	1.00	17.60
OH	304	CG	ARG	51	15.313	12.417	4.123	1.00	17.60	3.013	17.069	9.050	1.00	17.60
OH	305	C	ARG	51	14.728	11.747	6.317	1.00	17.60	1.366	16.300	7.569	1.00	17.60
OH	306	O	ARG	51	15.958	12.742	3.145	1.00	17.60	3.044	18.408	6.375	1.00	17.60
OH	307	N	ARG	52	16.599	11.722	2.295	1.00	17.60	3.602	19.330	6.979	1.00	17.60
OH	308	CA	ARG	52	17.729	12.371	1.492	1.00	17.60	2.001	18.629	5.601	1.00	17.60
OH	309	C	ARG	52	17.903	13.550	1.819	1.00	17.60	1.271	19.886	5.690	1.00	17.60
OH	310	N	ARG	53	18.518	11.607	0.556	1.00	17.60	1.586	20.885	4.567	1.00	17.60
OH	311	CA	ARG	53	19.369	12.712	-0.211	1.00	17.60	2.742	20.749	3.692	1.00	17.60
OH	312	CB	ARG	53	20.556	12.112	-0.777	1.00	17.60	2.378	19.569	2.828	1.00	17.60
OH	313	CG	ARG	53	21.209	13.380	-0.990	1.00	17.60	1.016	19.519	2.112	1.00	17.60
OH	314	CG	ARG	53	17.719	12.495	-1.401	1.00	17.60	-0.137	19.701	2.962	1.00	17.60
OH	315	C	ARG	53	18.599	13.241	-1.808	1.00	17.60	-0.253	19.682	5.695	1.00	17.60
OH	316	O	ARG	54	18.919	14.449	-1.964	1.00	17.60	-0.782	18.584	5.959	1.00	17.60
OH	317	N	ARG	54	18.231	15.267	-1.033	1.00	17.60	-0.993	20.743	5.432	1.00	17.60
OH	318	CA	ARG	54	18.194	14.724	-4.518	1.00	17.60	-2.415	20.832	5.610	1.00	17.60
OH	319	CB	ARG	54	18.455	13.239	-4.695	1.00	17.60	-2.705	21.558	6.769	1.00	17.60
OH	320	CG	ARG	54	19.751	12.365	-4.616	1.00	17.60	-6.039	21.721	7.497	1.00	17.60
OH	321	CG	ARG	54	17.397	12.367	-4.847	1.00	17.60	-6.267	22.732	8.413	1.00	17.60
OH	322	CD	ARG	54						-5.125	20.315	7.574	1.00	17.60

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388	CD1 H18	62	-5.937	21.388	0.527	1.00 17.60	ATOM	453	C	ALA	70	3.783	15.795	1.533	1.00 17.60
389	ND2 H18	62	-5.421	22.491	0.044	1.00 17.60	ATOM	454	O	ALA	70	3.437	15.320	0.450	1.00 17.60
390	C H18	67	-2.388	21.690	6.437	1.00 17.60	ATOM	455	N	ME1	71	4.548	16.909	1.586	1.00 17.60
391	C H18	62	-1.861	22.676	6.233	1.00 17.60	ATOM	456	CA	ME1	71	5.139	17.483	0.365	1.00 17.60
392	N LYS	63	-3.452	21.370	3.615	1.00 17.60	ATOM	457	CB	ME1	71	5.282	18.987	0.401	1.00 17.60
393	CA LYS	63	-4.021	21.905	2.508	1.00 17.60	ATOM	458	CG	ME1	71	4.212	19.775	-0.254	1.00 17.60
394	CB LYS	63	-4.988	21.035	3.707	1.00 17.60	ATOM	459	SD	ME1	71	4.873	21.444	-0.425	1.00 17.60
395	CD LYS	63	-5.893	21.054	0.716	1.00 17.60	ATOM	460	CE	ME1	71	3.859	22.475	0.584	1.00 17.60
396	CD LYS	63	-5.749	21.202	-0.539	1.00 17.60	ATOM	461	C	ME1	71	6.565	16.969	0.125	1.00 17.60
397	CE LYS	63	-5.618	22.261	-1.616	1.00 17.60	ATOM	462	O	ME1	71	7.301	16.966	1.121	1.00 17.60
398	N LYS	63	-4.432	23.198	3.235	1.00 17.60	ATOM	463	N	LYS	72	7.069	16.516	-1.031	1.00 17.60
399	C LYS	63	-4.836	22.931	3.236	1.00 17.60	ATOM	464	CA	LYS	72	8.474	16.136	-1.230	1.00 17.60
400	O LYS	63	-5.808	22.468	4.093	1.00 17.60	ATOM	465	CB	LYS	72	8.639	14.971	-2.168	1.00 17.60
401	N GLU	64	-4.609	24.235	2.912	1.00 17.60	ATOM	466	CC	LYS	72	9.921	14.141	-2.247	1.00 17.60
402	CA GLU	64	-5.653	25.218	3.629	1.00 17.60	ATOM	467	CC	LYS	72	9.686	12.126	-3.350	1.00 17.60
403	CB GLU	64	-5.235	26.666	3.337	1.00 17.60	ATOM	468	CE	LYS	72	10.493	12.015	-3.314	1.00 17.60
404	CD GLU	64	-3.724	27.045	3.392	1.00 17.60	ATOM	469	HE	LYS	72	11.931	12.403	-3.932	1.00 17.60
405	CD GLU	64	-3.314	28.509	3.676	1.00 17.60	ATOM	470	C	LYS	72	9.038	17.379	-1.067	1.00 17.60
406	EL GLU	64	-4.059	29.162	4.418	1.00 17.60	ATOM	471	O	LYS	72	8.510	17.864	-2.872	1.00 17.60
407	OF2 GLU	64	-2.265	29.002	3.198	1.00 17.60	ATOM	472	N	ILE	73	10.064	17.973	-1.325	1.00 17.60
408	C GLU	64	-7.175	24.955	3.454	1.00 17.60	ATOM	473	CA	ILE	73	10.516	19.214	-1.051	1.00 17.60
409	O	64	-7.960	25.886	3.613	1.00 17.60	ATOM	474	CB	ILE	73	10.532	20.295	-0.782	1.00 17.60
410	N SER	65	-7.464	23.748	3.079	1.00 17.60	ATOM	475	CG2	ILE	73	10.810	21.941	-1.462	1.00 17.60
411	CA SER	65	-8.057	23.326	3.010	1.00 17.60	ATOM	476	CG1	ILE	73	9.201	20.263	0.036	1.00 17.60
412	CD SER	65	-9.613	23.537	1.638	1.00 17.60	ATOM	477	CD1	ILE	73	9.227	21.445	0.988	1.00 17.60
413	O SER	65	-9.657	24.945	3.394	1.00 17.60	ATOM	478	C	ILE	73	11.921	18.970	-2.232	1.00 17.60
414	C SER	65	-9.351	21.876	3.406	1.00 17.60	ATOM	479	O	ILE	73	12.721	18.747	-3.393	1.00 17.60
415	SER	65	-9.235	20.999	2.697	1.00 17.60	ATOM	480	K	LEU	74	12.139	18.696	-3.561	1.00 17.60
416	K GLY	66	-8.724	21.668	4.599	1.00 17.60	ATOM	481	CA	LEU	74	13.427	18.498	-4.269	1.00 17.60
417	CA GLY	66	-8.009	20.469	5.313	1.00 17.60	ATOM	482	CB	LEU	74	13.316	17.022	-5.623	1.00 17.60
418	C GLY	66	-7.881	19.560	5.353	1.00 17.60	ATOM	483	CG	LEU	74	13.044	16.409	-5.844	1.00 17.60
419	O GLY	66	-7.280	19.461	6.670	1.00 17.60	ATOM	484	CD1	LEU	74	14.354	15.222	-5.168	1.00 17.60
420	N ASN	67	-7.413	18.993	4.465	1.00 17.60	ATOM	485	CD2	LEU	74	11.732	15.967	-5.291	1.00 17.60
421	CA ASN	67	-6.574	17.841	4.513	1.00 17.60	ATOM	486	C	LEU	74	14.024	19.054	-4.616	1.00 17.60
422	CB ASN	67	-6.451	17.021	3.235	1.00 17.60	ATOM	487	O	LEU	74	13.371	20.722	-5.203	1.00 17.60
423	CG ASN	67	-6.024	16.605	2.884	1.00 17.60	ATOM	488	N	ASP	75	15.294	20.063	-4.358	1.00 17.60
424	OD1 ASN	67	-8.214	15.593	3.340	1.00 17.60	ATOM	489	CA	ASP	75	15.864	21.384	-4.577	1.00 17.60
425	ND2 ASN	67	-8.632	17.407	2.022	1.00 17.60	ATOM	490	CB	ASP	75	17.023	21.597	-3.561	1.00 17.60
426	C ASN	67	-5.102	17.987	4.757	1.00 17.60	ATOM	491	CG2	ASP	75	17.817	22.892	-3.653	1.00 17.60
427	O ASN	67	-4.524	19.068	4.657	1.00 17.60	ATOM	492	OD1	ASP	75	18.303	23.194	-4.748	1.00 17.60
428	N HIS	68	-4.325	16.841	5.115	1.00 17.60	ATOM	493	OD2	ASP	75	18.046	23.592	-2.647	1.00 17.60
429	CA HIS	68	-3.105	16.720	5.349	1.00 17.60	ATOM	494	C	ASP	75	16.319	21.356	-5.996	1.00 17.60
430	CB HIS	68	-2.853	16.115	6.720	1.00 17.60	ATOM	495	O	ASP	75	17.334	20.626	-6.189	1.00 17.60
431	CG HIS	68	-3.156	17.072	7.885	1.00 17.60	ATOM	496	N	LYE	76	15.812	22.105	-6.962	1.00 17.60
432	CD2 HIS	68	-4.294	17.032	8.606	1.00 17.60	ATOM	497	CA	LYE	76	16.296	21.980	-8.344	1.00 17.60
433	ND1 HIS	68	-2.371	18.021	8.350	1.00 17.60	ATOM	498	CB	LYE	76	15.825	23.151	-9.221	1.00 17.60
434	CE1 HIS	68	-2.962	18.580	9.355	1.00 17.60	ATOM	499	CG	LYE	76	14.593	22.951	-9.711	1.00 17.60
435	N2 HIS	68	-6.306	17.987	9.486	1.00 17.60	ATOM	500	CD	LYE	76	12.857	23.988	-10.680	1.00 17.60
436	C HIS	68	-2.549	15.824	4.236	1.00 17.60	ATOM	501	CE	LYE	76	16.099	25.407	-10.127	1.00 17.60
437	O HIS	68	-3.278	15.041	3.615	1.00 17.60	ATOM	502	N2	LYE	76	13.055	26.340	-10.483	1.00 17.60
438	N TYR	69	-1.265	15.957	3.933	1.00 17.60	ATOM	503	C	LYE	76	17.870	21.864	-8.522	1.00 17.60
439	CA TYR	69	-0.812	15.375	2.813	1.00 17.60	ATOM	504	O	LYE	76	18.241	20.886	-9.139	1.00 17.60
440	CB TYR	69	-0.815	16.715	1.583	1.00 17.60	ATOM	505	N	GLN	77	18.665	22.716	-7.856	1.00 17.60
441	CG TYR	69	-2.254	16.372	0.908	1.00 17.60	ATOM	506	CA	GLN	77	20.154	22.762	-7.919	1.00 17.60
442	CD1 TYR	69	-3.013	15.273	0.628	1.00 17.60	ATOM	507	CB	GLN	77	20.883	23.889	-7.223	1.00 17.60
443	CE2 TYR	69	-4.287	15.413	0.077	1.00 17.60	ATOM	508	CG	GLN	77	20.007	25.074	-6.942	1.00 17.60
444	CD2 TYR	69	-2.765	17.629	0.405	1.00 17.60	ATOM	509	CD	GLN	77	20.651	26.417	-7.186	1.00 17.60
445	CE2 TYR	69	-4.018	17.783	0.036	1.00 17.60	ATOM	510	CE1	GLN	77	21.571	26.605	-7.972	1.00 17.60
446	CZ TYR	69	-6.779	16.657	-0.201	1.00 17.60	ATOM	511	NE2	GLN	77	20.096	27.405	-6.518	1.00 17.60
447	CH TYR	69	-6.104	16.750	-0.623	1.00 17.60	ATOM	512	C	GLN	77	20.803	21.534	-7.396	1.00 17.60
448	O TYR	69	-0.862	15.452	3.207	1.00 17.60	ATOM	513	O	GLN	77	21.516	20.883	-8.161	1.00 17.60
449	O TYR	69	1.211	15.956	4.268	1.00 17.60	ATOM	514	N	LYE	78	20.549	21.139	-6.137	1.00 17.60
450	N ALA	70	1.793	16.986	4.268	1.00 17.60	ATOM	515	CA	LYS	78	21.231	19.971	-5.634	1.00 17.60
451	CA ALA	70	3.197	15.038	2.711	1.00 17.60	ATOM	516	CB	LYS	78	20.955	19.832	-4.154	1.00 17.60
452	CB ALA	70	3.730	13.630	2.746	1.00 17.60	ATOM	517	CG	LYS	78	19.878	19.027	-3.541	1.00 17.60

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1	16.061	30.985	-17.407	1.00	17.60	ATOM	533	CO	GLU	86
2	15.074	21.724	-18.374	1.00	17.60	ATOM	534	CD	GLU	86
3	13.851	11.484	-18.384	1.00	17.60	ATOM	535	OE1	GLU	86
4	12.552	12.552	-19.136	1.00	17.60	ATOM	536	OE2	GLU	86
5	13.517	11.847	-14.413	1.00	17.60	ATOM	537	C	GLU	86
6	14.288	11.783	-14.597	1.00	17.60	ATOM	538	N	GLU	86
7	16.075	11.259	-15.338	1.00	17.60	ATOM	539	K	HIS	87
8	15.113	10.446	-12.532	1.00	17.60	ATOM	540	CA	HIS	87
9	15.727	9.448	-11.537	1.00	17.60	ATOM	541	CD	HIS	87
10	17.003	8.601	-11.858	1.00	17.60	ATOM	542	CG	HIS	87
11	16.119	8.826	-11.108	1.00	17.60	ATOM	543	CD2	HIS	87
12	17.327	7.679	-12.711	1.00	17.60	ATOM	544	ND1	HIS	87
13	18.551	7.308	-12.496	1.00	17.60	ATOM	545	CE1	HIS	87
14	19.012	6.005	-11.528	1.00	17.60	ATOM	546	NE2	HIS	87
15	14.252	11.361	-11.680	1.00	17.60	ATOM	547	C	HIS	87
16	13.380	10.833	-11.038	1.00	17.60	ATOM	548	O	HIS	87
17	14.357	12.674	-13.612	1.00	17.60	ATOM	549	N	THR	88
18	13.349	12.498	-10.934	1.00	17.60	ATOM	600	CA	THR	88
19	13.979	14.911	-10.816	1.00	17.60	ATOM	601	CB	THR	88
20	14.995	14.694	-9.818	1.00	17.60	ATOM	602	CO1	THR	88
21	12.968	15.930	-10.465	1.00	17.60	ATOM	603	CG2	THR	88
22	12.065	13.494	-11.832	1.00	17.60	ATOM	604	C	THR	88
23	10.895	13.407	-13.455	1.00	17.60	ATOM	605	O	THR	88
24	12.306	13.487	-13.118	1.00	17.60	ATOM	606	K	LEU	89
25	11.311	13.632	-14.352	1.00	17.60	ATOM	607	CA	LEU	89
26	12.123	14.039	-15.377	1.00	17.60	ATOM	608	CB	LEU	89
27	12.826	15.444	-15.260	1.00	17.60	ATOM	609	CO	LEU	89
28	13.869	15.545	-16.378	1.00	17.60	ATOM	610	CD1	LEU	89
29	11.816	16.591	-13.333	1.00	17.60	ATOM	611	CD2	LEU	89
30	10.482	12.615	-14.309	1.00	17.60	ATOM	612	C	LEU	89
31	9.375	12.529	-14.212	1.00	17.60	ATOM	613	O	LEU	89
32	11.141	11.270	-14.212	1.00	17.60	ATOM	614	N	ASN	90
33	10.507	9.954	-14.227	1.00	17.60	ATOM	615	CA	ASN	90
34	11.395	8.734	-14.201	1.00	17.60	ATOM	616	CB	ASN	90
35	12.357	8.498	-15.338	1.00	17.60	ATOM	617	CG	ASN	90
36	12.012	8.535	-16.514	1.00	17.60	ATOM	618	CO1	ASN	90
37	13.613	8.175	-15.045	1.00	17.60	ATOM	619	ND2	ASN	90
38	9.723	9.713	-12.976	1.00	17.60	ATOM	620	C	ASN	90
39	8.758	8.918	-13.048	1.00	17.60	ATOM	621	O	ASN	90
40	10.056	10.294	-11.819	1.00	17.60	ATOM	622	H	GLU	91
41	9.221	10.261	-9.359	1.00	17.60	ATOM	623	CA	GLU	91
42	10.200	11.651	-8.889	1.00	17.60	ATOM	624	CB	GLU	91
43	10.923	11.516	-7.524	1.00	17.60	ATOM	625	CO	GLU	91
44	10.343	10.981	-6.599	1.00	17.60	ATOM	626	OE1	GLU	91
45	12.051	12.054	-7.464	1.00	17.60	ATOM	627	OE2	GLU	91
46	7.919	10.767	-10.643	1.00	17.60	ATOM	628	O	GLU	91
47	6.875	10.159	-10.290	1.00	17.60	ATOM	629	C	GLU	91
48	7.832	12.046	-11.035	1.00	17.60	ATOM	630	O	GLU	91
49	6.681	12.753	-11.163	1.00	17.60	ATOM	631	N	LYS	92
50	6.950	14.192	-11.402	1.00	17.60	ATOM	632	CA	LYS	92
51	5.691	14.995	-11.282	1.00	17.60	ATOM	633	CB	LYS	92
52	4.914	15.064	-12.603	1.00	17.60	ATOM	634	CG	LYS	92
53	5.262	16.238	-13.569	1.00	17.60	ATOM	635	CD	LYS	92
54	4.611	16.062	-14.839	1.00	17.60	ATOM	636	CE	LYS	92
55	5.929	12.119	-12.296	1.00	17.60	ATOM	637	NE	LYS	92
56	4.761	11.807	-12.089	1.00	17.60	ATOM	638	O	LYS	92
57	6.462	11.660	-13.493	1.00	17.60	ATOM	639	C	ARG	93
58	5.668	11.249	-14.566	1.00	17.60	ATOM	640	N	ARG	93
59	6.568	11.160	-15.753	1.00	17.60	ATOM	641	CA	ARG	93
60	5.876	11.035	-17.112	1.00	17.60	ATOM	642	CB	ARG	93
61	6.844	11.306	-18.281	1.00	17.60	ATOM	643	CG	ARG	93
62	6.085	11.237	-19.523	1.00	17.60	ATOM	644	CD	ARG	93
63	6.594	10.762	-20.661	1.00	17.60	ATOM	645	CE	ARG	93
64	7.054	10.330	-20.727	1.00	17.60	ATOM	646	C	ARG	93
65						ATOM	647	HN1	ARG	93

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668	KH2	ARG	93	5.798	10.727	-21.775	1.00	17.60	ATOK	713	O	PRO	101	-3.173	0.442	-1.542	1.00	17.60
669	C	ARG	93	5.023	9.866	-14.274	1.00	17.60	ATOK	714	N	PHE	102	-1.549	-0.293	-3.016	1.00	17.60
670	C	ARG	93	3.834	9.591	-14.540	1.00	17.60	ATOK	715	CA	PHE	102	-0.506	-0.267	-2.125	1.00	17.60
671	N	ILE	94	5.830	9.043	-13.661	1.00	17.60	ATOK	716	CB	PHE	102	0.054	-1.735	-1.937	1.00	17.60
672	CA	ILE	94	5.297	7.759	-13.306	1.00	17.60	ATOK	717	CG	PHE	102	-0.930	-2.916	-1.742	1.00	17.60
673	CB	ILE	94	6.456	6.769	-12.958	1.00	17.60	ATOK	718	CD1	PHE	102	-0.087	-1.019	-2.503	1.00	17.60
674	CD1	ILE	94	5.805	5.467	-12.409	1.00	17.60	ATOK	719	CD2	PHE	102	-1.881	-2.933	-0.712	1.00	17.60
675	CD2	ILE	94	7.373	6.549	-14.191	1.00	17.60	ATOK	720	CD3	PHE	102	-1.748	-3.093	-2.462	1.00	17.60
676	CD3	ILE	94	8.580	5.756	-13.815	1.00	17.60	ATOK	721	CD4	PHE	102	-2.742	-1.022	-0.576	1.00	17.60
677	C	ILE	94	4.368	7.963	-12.117	1.00	17.60	ATOK	722	C	PHE	102	-2.612	-5.092	-1.453	1.00	17.60
678	O	ILE	94	3.251	7.448	-12.183	1.00	17.60	ATOK	723	C	PHE	102	0.543	0.693	-2.742	1.00	17.60
679	N	LEU	95	4.605	8.709	-11.052	1.00	17.60	ATOK	724	N	LEU	103	1.742	0.472	-2.577	1.00	17.60
680	CA	LEU	95	3.739	8.765	-9.957	1.00	17.60	ATOK	725	CA	LEU	103	0.247	1.750	-3.509	1.00	17.60
681	CB	LEU	95	4.372	9.502	-8.745	1.00	17.60	ATOK	726	CB	LEU	103	1.263	2.698	-3.944	1.00	17.60
682	CD1	LEU	95	5.654	8.546	-7.715	1.00	17.60	ATOK	727	CD1	LEU	103	1.315	2.916	-5.357	1.00	17.60
683	CD2	LEU	95	5.654	9.364	-6.612	1.00	17.60	ATOK	728	CD2	LEU	103	1.974	1.981	-6.197	1.00	17.60
684	CD2	LEU	95	4.088	7.682	-6.922	1.00	17.60	ATOK	729	CD3	LEU	103	1.542	0.638	-6.143	1.00	17.60
685	C	LEU	95	2.405	9.443	-10.333	1.00	17.60	ATOK	730	CD4	LEU	103	1.852	2.566	-7.571	1.00	17.60
686	O	LEU	95	3.377	9.036	-9.792	1.00	17.60	ATOK	731	C	LEU	103	0.808	4.024	-3.418	1.00	17.60
687	N	GLN	96	2.282	10.384	-11.261	1.00	17.60	ATOK	732	O	LEU	103	-0.401	4.127	-3.231	1.00	17.60
688	CA	GLN	96	0.971	10.898	-11.565	1.00	17.60	ATOK	733	N	VAL	104	1.574	5.080	-3.196	1.00	17.60
689	CB	GLN	96	1.064	12.220	-12.306	1.00	17.60	ATOK	734	CA	VAL	104	0.967	6.234	-2.575	1.00	17.60
690	CD	GLN	96	1.986	12.334	-13.505	1.00	17.60	ATOK	735	CB	VAL	104	1.834	6.750	-1.428	1.00	17.60
691	CD1	GLN	96	1.285	12.173	-14.819	1.00	17.60	ATOK	736	CD1	VAL	104	3.194	7.227	-1.906	1.00	17.60
692	CD2	GLN	96	0.470	12.994	-15.297	1.00	17.60	ATOK	737	CD2	VAL	104	1.060	7.855	-0.716	1.00	17.60
693	CD3	GLN	96	1.586	11.117	-15.571	1.00	17.60	ATOK	738	C	VAL	104	0.786	7.262	-3.510	1.00	17.60
694	C	GLN	96	0.266	9.882	-12.401	1.00	17.60	ATOK	739	O	VAL	104	1.716	7.596	-4.357	1.00	17.60
695	N	ALA	97	-0.945	9.794	-12.303	1.00	17.60	ATOK	740	N	LYS	105	-0.449	7.803	-4.569	1.00	17.60
696	CA	ALA	97	0.236	9.068	-13.208	1.00	17.60	ATOK	741	CA	LYS	105	-0.979	8.736	-4.569	1.00	17.60
697	CB	ALA	97	0.120	8.133	-14.059	1.00	17.60	ATOK	742	CB	LYS	105	-2.470	8.934	-4.328	1.00	17.60
698	CD	ALA	97	-0.067	7.900	-15.213	1.00	17.60	ATOK	743	CD	LYS	105	-3.467	7.803	-4.596	1.00	17.60
699	C	ALA	97	-1.052	6.090	-13.891	1.00	17.60	ATOK	744	CD	LYS	105	-4.642	7.737	-3.554	1.00	17.60
700	N	VAL	98	0.532	6.797	-13.436	1.00	17.60	ATOK	745	CD	LYS	105	-4.355	6.777	-2.332	1.00	17.60
701	CA	VAL	98	0.224	6.343	-12.413	1.00	17.60	ATOK	746	N2	LYS	105	-4.578	5.325	-2.373	1.00	17.60
702	CB	VAL	98	1.547	5.084	-11.792	1.00	17.60	ATOK	747	C	LYS	105	-0.301	10.093	-4.564	1.00	17.60
703	CD1	VAL	98	1.770	5.965	-9.013	1.00	17.60	ATOK	748	O	LYS	105	-0.028	10.626	-3.509	1.00	17.60
704	CD2	VAL	98	1.505	3.320	-10.879	1.00	17.60	ATOK	749	N	LEU	106	-0.045	10.700	-5.701	1.00	17.60
705	C	VAL	98	-1.052	3.130	-10.808	1.00	17.60	ATOK	750	CA	LEU	106	0.580	11.991	-5.866	1.00	17.60
706	N	ASN	99	-1.586	6.199	-10.521	1.00	17.60	ATOK	751	CB	LEU	106	1.469	12.029	-7.113	1.00	17.60
707	CA	ASN	99	-1.825	3.977	-10.516	1.00	17.60	ATOK	752	CD	LEU	106	2.062	13.324	-7.608	1.00	17.60
708	CB	ASN	99	-2.740	3.837	-9.555	1.00	17.60	ATOK	753	CD1	LEU	106	3.021	13.315	-6.500	1.00	17.60
709	CD	ASN	99	-4.052	4.307	-10.136	1.00	17.60	ATOK	754	CD2	LEU	106	3.021	13.324	-6.500	1.00	17.60
710	C	ASN	99	-3.295	4.391	-9.222	1.00	17.60	ATOK	755	C	LEU	106	-0.674	12.746	-6.113	1.00	17.60
711	N	ASN	99	-6.191	5.325	-9.398	1.00	17.60	ATOK	756	O	LEU	106	-1.284	12.451	-7.124	1.00	17.60
712	CA	ASN	99	-5.983	3.508	-8.266	1.00	17.60	ATOK	757	N	GLU	107	-1.331	13.585	-5.196	1.00	17.60
713	CB	ASN	99	-2.924	2.343	-9.207	1.00	17.60	ATOK	758	CA	GLU	107	-2.315	14.385	-5.404	1.00	17.60
714	CD1	ASN	99	-3.281	1.531	-10.063	1.00	17.60	ATOK	759	CB	GLU	107	-3.055	14.665	-4.107	1.00	17.60
715	CD2	ASN	99	-2.679	1.959	-7.911	1.00	17.60	ATOK	760	CD	GLU	107	-4.575	14.367	-4.166	1.00	17.60
716	CD3	ASN	99	-2.796	0.599	-7.412	1.00	17.60	ATOK	761	CD1	GLU	107	-4.910	13.097	-3.311	1.00	17.60
717	C	ASN	99	-1.481	-0.145	-7.771	1.00	17.60	ATOK	762	CD2	GLU	107	-4.594	13.105	-2.110	1.00	17.60
718	N	PHE	100	-1.617	-1.662	-7.671	1.00	17.60	ATOK	763	CD3	GLU	107	-5.485	12.118	-3.844	1.00	17.60
719	CA	PHE	100	-2.400	-1.350	-8.582	1.00	17.60	ATOK	764	C	GLU	107	-2.032	15.721	-6.031	1.00	17.60
720	CB	PHE	100	-1.075	-2.337	-6.632	1.00	17.60	ATOK	765	O	GLU	107	-2.976	16.285	-5.926	1.00	17.60
721	CD1	PHE	100	-2.665	-3.687	-8.458	1.00	17.60	ATOK	766	N	PHE	108	-0.861	16.330	-5.911	1.00	17.60
722	CD2	PHE	100	-1.364	-3.687	-6.439	1.00	17.60	ATOK	767	CA	PHE	108	-0.535	17.542	-6.663	1.00	17.60
723	CD3	PHE	100	-2.133	-4.261	-7.403	1.00	17.60	ATOK	768	CB	PHE	108	-0.806	18.045	-5.139	1.00	17.60
724	C	PHE	100	-3.038	0.719	-5.909	1.00	17.60	ATOK	769	CD	PHE	108	-2.106	18.954	-5.926	1.00	17.60
725	N	PRO	101	-3.457	-0.211	-5.119	1.00	17.60	ATOK	770	CD1	PHE	108	-3.253	19.454	-5.737	1.00	17.60
726	CA	PRO	101	-4.307	-1.418	-5.637	1.00	17.60	ATOK	771	CD2	PHE	108	-2.153	18.516	-3.831	1.00	17.60
727	CB	PRO	101	-3.995	-0.087	-3.716	1.00	17.60	ATOK	772	CD3	PHE	108	-4.432	19.506	-5.030	1.00	17.60
728	C	PRO	101	-4.813	-1.332	-3.411	1.00	17.60	ATOK	773	CD4	PHE	108	-3.343	18.571	-3.130	1.00	17.60
729	N	PRO	101	-5.468	-1.633	-4.722	1.00	17.60	ATOK	774	C	PHE	108	-4.509	19.058	-3.715	1.00	17.60
730	CA	PRO	101	-2.890	0.068	-2.677	1.00	17.60	ATOK	775	C	PHE	108	0.961	17.592	-7.003	1.00	17.60
731	CB	PRO	101						ATOK	776	O	PHE	108	1.819	17.092	-6.772	1.00	17.60
732	C	PRO	101						ATOK	777	N	SER	109	1.297	18.190	-8.129	1.00	17.60

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178	CA	SER	109	2.655	18.365	-8.589	1.00	17.60	ATOM	863	CB	TIR	117	6.306	20.817	-4.462	1.00	17.60	ATOM
179	CB	SER	109	2.662	17.597	-9.865	1.00	17.60	ATOM	864	CB	TIR	117	7.033	22.102	-4.125	1.00	17.60	ATOM
180	CB	SER	109	4.217	17.409	-10.103	1.00	17.60	ATOM	865	CO1	TIR	117	7.271	27.393	-2.802	1.00	17.60	ATOM
181	CB	SER	109	7.706	19.657	-8.855	1.00	17.60	ATOM	866	CE1	TIR	117	7.946	23.503	-2.440	1.00	17.60	ATOM
182	CB	SER	109	1.634	20.476	-8.994	1.00	17.60	ATOM	867	CD2	TIR	117	7.549	22.933	-3.106	1.00	17.60	ATOM
183	CB	SER	110	3.076	20.486	-8.827	1.00	17.60	ATOM	868	CE2	TIR	117	8.270	24.046	-4.756	1.00	17.60	ATOM
184	CB	SER	110	4.055	21.089	-9.142	1.00	17.60	ATOM	869	CA	TIR	117	9.401	24.335	-3.409	1.00	17.60	ATOM
185	CB	SER	110	3.241	22.837	-8.287	1.00	17.60	ATOM	870	OH	TIR	117	9.204	25.417	-3.010	1.00	17.60	ATOM
186	CB	SER	110	3.348	22.840	-6.779	1.00	17.60	ATOM	871	C	TIR	117	6.231	18.602	-5.630	1.00	17.60	ATOM
187	CB	SER	110	2.493	22.060	-6.009	1.00	17.60	ATOM	872	O	TIR	117	5.977	18.601	-6.820	1.00	17.60	ATOM
188	CB	SER	110	4.101	23.712	-6.335	1.00	17.60	ATOM	873	N	VAL	118	5.943	17.644	-4.771	1.00	17.60	ATOM
189	CB	SER	110	2.435	22.149	-4.613	1.00	17.60	ATOM	874	CA	VAL	118	4.982	16.609	-5.121	1.00	17.60	ATOM
190	CB	SER	110	4.116	23.792	-4.750	1.00	17.60	ATOM	875	CB	VAL	118	5.721	15.356	-5.428	1.00	17.60	ATOM
191	CB	SER	110	3.253	23.029	-3.970	1.00	17.60	ATOM	876	CC	VAL	118	6.495	15.405	-6.683	1.00	17.60	ATOM
192	CB	SER	110	5.488	22.211	-8.891	1.00	17.60	ATOM	877	SD	VAL	118	7.404	13.875	-6.645	1.00	17.60	ATOM
193	CB	SER	110	6.172	21.324	-8.370	1.00	17.60	ATOM	878	CB	VAL	118	9.614	14.507	-7.767	1.00	17.60	ATOM
194	CB	SER	110	5.074	23.420	-9.222	1.00	17.60	ATOM	879	C	VAL	118	4.069	16.390	-3.929	1.00	17.60	ATOM
195	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	880	O	VAL	118	6.621	16.216	-2.862	1.00	17.60	ATOM
196	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	881	N	VAL	119	2.755	16.344	-3.930	1.00	17.60	ATOM
197	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	882	CA	VAL	119	2.041	16.318	-2.692	1.00	17.60	ATOM
198	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	883	CB	VAL	119	1.091	17.472	-2.663	1.00	17.60	ATOM
199	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	884	CO1	VAL	119	0.564	17.509	-1.263	1.00	17.60	ATOM
200	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	885	CO2	VAL	119	1.740	16.803	-2.575	1.00	17.60	ATOM
201	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	886	C	VAL	119	1.319	15.004	-2.667	1.00	17.60	ATOM
202	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	887	O	VAL	119	0.445	14.820	-3.486	1.00	17.60	ATOM
203	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	888	N	VAL	120	1.616	14.075	-3.778	1.00	17.60	ATOM
204	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	889	CA	VAL	120	1.057	12.730	-1.700	1.00	17.60	ATOM
205	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	890	CB	VAL	120	2.087	11.658	-1.304	1.00	17.60	ATOM
206	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	891	CO	VAL	120	2.961	10.918	-2.339	1.00	17.60	ATOM
207	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	892	SD	VAL	120	4.298	11.751	-3.236	1.00	17.60	ATOM
208	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	893	CB	VAL	120	5.646	11.781	-2.077	1.00	17.60	ATOM
209	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	894	C	VAL	120	0.017	12.456	-0.637	1.00	17.60	ATOM
210	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	895	O	VAL	120	0.024	13.337	-0.210	1.00	17.60	ATOM
211	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	896	N	VAL	121	-0.036	11.663	-0.562	1.00	17.60	ATOM
212	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	897	CA	VAL	121	-1.762	11.401	0.547	1.00	17.60	ATOM
213	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	898	CB	VAL	121	-2.822	10.132	0.175	1.00	17.60	ATOM
214	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	899	CO	VAL	121	-4.029	9.811	0.752	1.00	17.60	ATOM
215	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	900	CD	VAL	121	-5.018	11.008	0.925	1.00	17.60	ATOM
216	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	901	CE1	VAL	121	-5.961	11.178	0.107	1.00	17.60	ATOM
217	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	902	CE2	VAL	121	-4.842	11.781	1.904	1.00	17.60	ATOM
218	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	903	C	VAL	121	-0.031	11.183	1.782	1.00	17.60	ATOM
219	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	904	O	VAL	121	-0.339	10.775	1.615	1.00	17.60	ATOM
220	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	905	N	VAL	122	-1.229	11.496	3.036	1.00	17.60	ATOM
221	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	906	CA	VAL	122	-0.382	11.200	4.215	1.00	17.60	ATOM
222	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	907	CB	VAL	122	-0.347	12.358	5.297	1.00	17.60	ATOM
223	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	908	CO	VAL	122	-0.007	12.736	6.722	1.00	17.60	ATOM
224	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	909	CD1	VAL	122	1.326	12.114	7.020	1.00	17.60	ATOM
225	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	910	CD2	VAL	122	1.753	11.928	8.337	1.00	17.60	ATOM
226	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	911	CO1	VAL	122	-0.914	12.189	7.756	1.00	17.60	ATOM
227	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	912	CO2	VAL	122	-0.515	12.010	9.069	1.00	17.60	ATOM
228	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	913	C	VAL	122	0.873	11.881	9.354	1.00	17.60	ATOM
229	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	914	OK	VAL	122	1.395	11.740	10.677	1.00	17.60	ATOM
230	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	915	C	VAL	122	-0.782	9.825	9.773	1.00	17.60	ATOM
231	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	916	N	VAL	123	-1.932	9.490	5.034	1.00	17.60	ATOM
232	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	917	N	VAL	123	0.223	8.961	4.912	1.00	17.60	ATOM
233	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	918	CA	VAL	123	0.075	7.622	5.498	1.00	17.60	ATOM
234	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	919	CB	VAL	123	0.811	6.558	4.595	1.00	17.60	ATOM
235	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	920	CO1	VAL	123	0.558	5.138	5.048	1.00	17.60	ATOM
236	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	921	CO2	VAL	123	0.426	6.540	3.140	1.00	17.60	ATOM
237	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	922	C	VAL	123	0.668	7.786	6.928	1.00	17.60	ATOM
238	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	923	N	VAL	124	-0.195	7.888	7.953	1.00	17.60	ATOM
239	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	924	K	ALA	124	-0.253	8.111	9.336	1.00	17.60	ATOM
240	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	925	CA	ALA	124	-0.856	8.433	10.323	1.00	17.60	ATOM
241	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	926	CB	ALA	124	-0.942	6.966	10.002	1.00	17.60	ATOM
242	CB	SER	110	8.346	23.370	-15.166	1.00	17.60	ATOM	927	ALA	124							ATOM

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OM	908	O	ALA	124	1.619	7.108	11.015	1.00 17.60	ATOM	973	CD	ARG	133	16.509	-1.154	12.712	1.00 17.60
OM	909	N	GLY	125	0.003	5.703	9.411	1.00 17.60	ATOM	974	ME	ARG	133	16.411	-2.149	11.645	1.00 17.60
OM	910	CA	GLY	125	1.321	6.619	10.053	1.00 17.60	ATOM	975	CE	ARG	133	16.906	-1.999	10.468	1.00 17.60
OM	911	C	GLY	125	2.781	4.749	10.287	1.00 17.60	ATOM	976	NH1	ARG	133	17.701	-0.934	10.129	1.00 17.60
OM	912	O	GLY	125	3.783	4.164	11.219	1.00 17.60	ATOM	977	NH2	ARG	133	16.786	-2.931	9.564	1.00 17.60
OM	913	CA	GLY	126	3.168	5.557	9.491	1.00 17.60	ATOM	978	C	ARG	133	11.166	-1.892	15.641	1.00 17.60
OM	914	N	GLY	126	4.895	5.672	9.620	1.00 17.60	ATOM	979	O	ARG	133	13.719	-2.656	16.405	1.00 17.60
OM	915	C	GLY	126	4.501	6.071	8.871	1.00 17.60	ATOM	980	N	ARG	134	12.160	-1.127	16.059	1.00 17.60
OM	916	GLY	GLY	127	6.778	3.599	8.397	1.00 17.60	ATOM	981	CA	ARG	134	11.617	-1.223	17.406	1.00 17.60
OM	917	N	GLY	127	6.778	4.543	8.788	1.00 17.60	ATOM	982	CB	ARG	134	10.571	-0.173	17.527	1.00 17.60
OM	918	CA	GLY	127	7.556	3.574	8.033	1.00 17.60	ATOM	983	CG	ARG	134	8.494	-0.106	18.601	1.00 17.60
OM	919	CG	GLY	127	9.012	4.010	8.130	1.00 17.60	ATOM	984	CD	ARG	134	8.306	0.224	17.697	1.00 17.60
OM	920	CG	GLY	127	9.290	5.212	7.223	1.00 17.60	ATOM	985	CE	ARG	134	7.109	0.812	18.299	1.00 17.60
OM	921	CD	GLY	127	10.662	5.063	7.388	1.00 17.60	ATOM	986	CE	ARG	134	5.833	-0.517	17.943	1.00 17.60
OM	922	CD	GLY	127	11.631	5.105	7.412	1.00 17.60	ATOM	987	NH1	ARG	134	5.584	-0.425	16.990	1.00 17.60
OM	923	CD	GLY	127	10.765	7.301	7.443	1.00 17.60	ATOM	988	NH2	ARG	134	4.804	1.191	18.559	1.00 17.60
OM	924	C	GLY	127	7.371	3.126	6.480	1.00 17.60	ATOM	989	C	ARG	134	11.013	-2.576	17.815	1.00 17.60
OM	925	O	GLY	127	6.912	1.040	9.590	1.00 17.60	ATOM	990	O	ARG	134	11.221	-3.024	18.956	1.00 17.60
OM	926	N	GLY	128	7.713	1.166	7.637	1.00 17.60	ATOM	991	N	ILE	135	10.200	-3.230	16.935	1.00 17.60
OM	927	CA	GLY	128	7.707	-0.327	8.017	1.00 17.60	ATOM	992	CA	ILE	135	9.137	-4.549	17.230	1.00 17.60
OM	928	CB	GLY	128	7.739	-1.085	6.811	1.00 17.60	ATOM	993	CB	ILE	135	8.598	-4.780	16.165	1.00 17.60
OM	929	CG	GLY	128	7.206	-2.477	5.980	1.00 17.60	ATOM	994	CG	ILE	135	8.375	-6.216	16.096	1.00 17.60
OM	930	SD	GLY	128	8.115	-3.435	5.769	1.00 17.60	ATOM	995	CG	ILE	135	6.259	-3.902	16.492	1.00 17.60
OM	931	CE	GLY	128	9.353	-2.652	6.997	1.00 17.60	ATOM	996	CD	ILE	135	10.879	-5.609	17.196	1.00 17.60
OM	932	C	GLY	128	9.009	-0.384	8.758	1.00 17.60	ATOM	997	C	ILE	135	10.879	-5.609	17.196	1.00 17.60
OM	933	O	GLY	128	9.141	-1.290	9.360	1.00 17.60	ATOM	998	O	ILE	135	10.879	-5.609	17.196	1.00 17.60
OM	934	N	GLY	129	10.012	0.450	8.478	1.00 17.60	ATOM	999	N	GLY	136	10.879	-5.609	17.196	1.00 17.60
OM	935	CA	GLY	129	11.313	0.501	9.117	1.00 17.60	ATOM	1000	CA	GLY	136	11.013	-2.576	17.815	1.00 17.60
OM	936	CB	GLY	129	12.147	1.571	8.430	1.00 17.60	ATOM	1001	C	GLY	136	11.013	-2.576	17.815	1.00 17.60
OM	937	CG	GLY	129	13.521	1.700	9.005	1.00 17.60	ATOM	1002	O	GLY	136	11.894	-8.031	14.766	1.00 17.60
OM	938	CD	GLY	129	14.580	1.068	8.491	1.00 17.60	ATOM	1003	N	ARG	137	11.714	-9.123	13.819	1.00 17.60
OM	939	CD	GLY	129	13.725	2.642	10.021	1.00 17.60	ATOM	1004	CA	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	940	CE	GLY	129	15.054	1.210	8.994	1.00 17.60	ATOM	1005	CB	ARG	137	11.714	-9.123	13.819	1.00 17.60
OM	941	CE	GLY	129	14.994	2.768	10.311	1.00 17.60	ATOM	1006	CG	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	942	CE	GLY	129	16.067	2.067	10.016	1.00 17.60	ATOM	1007	CD	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	943	C	GLY	129	11.246	9.706	10.592	1.00 17.60	ATOM	1008	CE	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	944	O	GLY	129	11.741	-0.040	11.339	1.00 17.60	ATOM	1009	CB	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	945	N	GLY	130	10.709	1.918	11.024	1.00 17.60	ATOM	1010	NH1	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	946	CA	GLY	130	10.661	2.290	12.412	1.00 17.60	ATOM	1011	NH2	ARG	137	11.894	-8.031	14.766	1.00 17.60
OM	947	CB	GLY	130	9.782	3.527	12.327	1.00 17.60	ATOM	1012	C	ARG	137	10.194	-15.262	14.101	1.00 17.60
OM	948	CG	GLY	130	9.929	4.482	11.492	1.00 17.60	ATOM	1013	O	ARG	137	10.254	-9.037	13.424	1.00 17.60
OM	949	C	GLY	130	10.080	1.173	13.391	1.00 17.60	ATOM	1014	N	PHE	138	9.444	-8.319	14.005	1.00 17.60
OM	950	O	GLY	130	10.646	0.849	14.325	1.00 17.60	ATOM	1015	CA	PHE	138	9.833	-9.752	12.612	1.00 17.60
OM	951	N	GLY	131	9.034	0.404	12.217	1.00 17.60	ATOM	1016	CB	PHE	138	8.445	-9.905	12.126	1.00 17.60
OM	952	CA	GLY	131	8.451	-0.868	13.671	1.00 17.60	ATOM	1017	CD	PHE	138	8.107	-9.239	10.777	1.00 17.60
OM	953	CB	GLY	131	7.065	-0.414	13.213	1.00 17.60	ATOM	1018	CE	PHE	138	7.903	-7.736	10.926	1.00 17.60
OM	954	CG	GLY	131	5.595	0.806	12.351	1.00 17.60	ATOM	1019	CD	PHE	138	8.339	-6.054	9.935	1.00 17.60
OM	955	CD	GLY	131	6.277	0.414	13.213	1.00 17.60	ATOM	1020	CE	PHE	138	7.325	-2.184	12.059	1.00 17.60
OM	956	ND	GLY	131	6.211	1.349	14.145	1.00 17.60	ATOM	1021	CE	PHE	138	8.204	-5.485	10.125	1.00 17.60
OM	957	CD	GLY	131	5.412	2.255	13.672	1.00 17.60	ATOM	1022	C	PHE	138	7.194	-5.816	12.225	1.00 17.60
OM	958	ND	GLY	131	4.997	1.969	12.477	1.00 17.60	ATOM	1023	C	PHE	138	7.634	-4.960	11.258	1.00 17.60
OM	959	O	GLY	131	9.219	-1.891	13.447	1.00 17.60	ATOM	1024	N	PHE	138	8.268	-11.432	12.126	1.00 17.60
OM	960	C	GLY	131	9.201	-2.756	14.292	1.00 17.60	ATOM	1025	N	PHE	138	9.146	-12.244	11.834	1.00 17.60
OM	961	CA	GLY	132	9.873	-2.149	12.335	1.00 17.60	ATOM	1026	CA	SEP	139	7.120	-11.054	12.467	1.00 17.60
OM	962	CB	GLY	132	10.695	-3.319	12.198	1.00 17.60	ATOM	1027	CB	SEP	139	6.779	-13.266	12.651	1.00 17.60
OM	963	CB	GLY	132	11.209	-3.405	10.776	1.00 17.60	ATOM	1028	CG	SEP	139	5.593	-13.527	13.505	1.00 17.60
OM	964	CD	GLY	132	12.351	-4.312	10.401	1.00 17.60	ATOM	1029	CD	SEP	139	4.707	-12.396	13.489	1.00 17.60
OM	965	CD	GLY	132	11.860	-5.711	10.294	1.00 17.60	ATOM	1030	CE	SEP	139	3.114	-12.809	13.805	1.00 17.60
OM	966	CD	GLY	132	12.926	-3.876	9.118	1.00 17.60	ATOM	1031	CE	SEP	139	2.286	-11.533	13.919	1.00 17.60
OM	967	O	GLY	132	11.834	-3.826	13.174	1.00 17.60	ATOM	1032	CE	SEP	139	2.744	-13.900	12.825	1.00 17.60
OM	968	N	GLY	132	12.091	-4.064	13.923	1.00 17.60	ATOM	1033	C	SEP	139	3.351	-13.492	15.324	1.00 17.60
OM	969	K	ARG	133	12.563	-1.998	13.252	1.00 17.60	ATOM	1034	O	SEP	139	5.433	-13.567	11.232	1.00 17.60
OM	970	CA	ARG	133	13.645	-1.052	14.200	1.00 17.60	ATOM	1035	K	GLU	140	5.822	-12.756	10.532	1.00 17.60
OM	971	CB	ARG	133	14.381	-0.539	13.971	1.00 17.60	ATOM	1036	CA	GLU	140	6.723	-14.802	10.923	1.00 17.60
OM	972	CC	ARG	133	15.341	-0.337	12.722	1.00 17.60	ATOM	1037	CB	GLU	140	6.700	-15.200	9.565	1.00 17.60
OM	973	CC	ARG	133	15.341	-0.337	12.722	1.00 17.60	ATOM	1037	CB	GLU	140	7.060	-16.779	9.650	1.00 17.60

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ATOM	1038	CG	GLU	140	7.591	-17.418	8.358	1.00	17.60
ATOM	1039	CD	GLU	140	8.821	-10.290	8.358	1.00	17.60
ATOM	1040	OEL	GLU	140	8.745	-19.493	8.344	1.00	17.60
ATOM	1041	E2	GLU	140	9.866	-17.767	8.919	1.00	17.63
ATOM	1042	C	GLU	140	5.361	-14.979	8.868	1.00	17.60
ATOM	1043	Q	GLU	140	5.391	-14.429	7.785	1.00	17.60
ATOM	1044	N	PRO	141	4.196	-15.170	9.485	1.00	17.60
ATOM	1045	CD	PRO	141	3.994	-15.981	10.676	1.00	17.60
ATOM	1046	CA	PRO	141	2.893	-14.737	8.978	1.00	17.60
ATOM	1047	CB	PRO	141	3.918	-15.241	9.996	1.00	17.60
ATOM	1048	CG	PRO	141	2.601	-16.490	10.476	1.00	17.60
ATOM	1049	C	PRO	141	2.715	-13.263	8.749	1.00	17.60
ATOM	1050	Q	PRO	141	1.998	-12.776	7.885	1.00	17.60
ATOM	1051	N	HIS	142	9.342	-12.572	9.659	1.00	17.60
ATOM	1052	CA	HIS	142	3.305	-11.166	9.556	1.00	17.60
ATOM	1053	CB	HIS	142	3.719	-10.381	10.911	1.00	17.60
ATOM	1054	CG	HIS	142	3.439	-9.081	11.717	1.00	17.60
ATOM	1055	CD2	HIS	142	2.872	-8.232	10.194	1.00	17.60
ATOM	1056	ND1	HIS	142	3.782	-8.312	12.172	1.00	17.60
ATOM	1057	CE1	HIS	142	3.474	-7.067	11.917	1.00	17.60
ATOM	1058	NE2	HIS	142	2.927	-7.032	10.728	1.00	17.60
ATOM	1059	C	HIS	142	4.272	-10.860	8.384	1.00	17.60
ATOM	1060	O	HIS	142	3.875	-10.053	7.826	1.00	17.60
ATOM	1061	N	ALA	143	5.445	-11.483	8.709	1.00	17.60
ATOM	1062	CA	ALA	143	6.343	-11.106	7.753	1.00	17.60
ATOM	1063	CB	ALA	143	7.597	-11.926	7.753	1.00	17.60
ATOM	1064	C	ALA	143	5.803	-11.411	5.957	1.00	17.60
ATOM	1065	O	ALA	143	5.545	-10.577	4.955	1.00	17.60
ATOM	1066	N	ARG	144	4.876	-12.507	7.737	1.00	17.60
ATOM	1067	CA	ARG	144	4.101	-12.834	4.508	1.00	17.60
ATOM	1068	CB	ARG	144	3.432	-14.131	4.769	1.00	17.60
ATOM	1069	CG	ARG	144	2.859	-14.890	3.614	1.00	17.60
ATOM	1070	CD	ARG	144	2.304	-16.120	4.217	1.00	17.60
ATOM	1071	NE	ARG	144	3.370	-17.120	4.581	1.00	17.60
ATOM	1072	CE	ARG	144	3.352	-17.817	5.710	1.00	17.60
ATOM	1073	NH1	ARG	144	2.368	-17.634	6.573	1.00	17.60
ATOM	1074	NH2	ARG	144	4.387	-18.677	9.543	1.00	17.60
ATOM	1075	C	ARG	144	3.046	-11.815	4.275	1.00	17.60
ATOM	1076	O	ARG	144	2.812	-11.516	3.099	1.00	17.60
ATOM	1077	N	PHE	145	2.390	-13.159	3.829	1.00	17.60
ATOM	1078	CA	PHE	145	1.401	-10.133	4.916	1.00	17.60
ATOM	1079	CB	PHE	145	0.836	-9.382	6.257	1.00	17.60
ATOM	1080	CC	PHE	145	-0.341	-8.612	6.131	1.00	17.60
ATOM	1081	PH1	PHE	145	-1.587	-9.066	5.762	1.00	17.60
ATOM	1082	CD2	PHE	145	-0.172	-7.264	6.366	1.00	17.60
ATOM	1083	CE2	PHE	145	-				

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1168	O	GLU	155	2.807	-6.420	-12.652	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1169	K	TYR	156	1.463	-5.494	-10.823	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1170	CA	TYR	156	1.463	-5.494	-10.823	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1171	CP	TYR	156	0.561	-3.787	-10.654	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1172	CC	TYR	156	0.215	-2.367	-10.974	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1173	CD1	TYR	156	-0.719	-2.079	-11.965	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1174	CE1	TYR	156	-1.057	-0.783	-12.230	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1175	CD2	TYR	156	0.828	-1.364	-10.228	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1176	CE2	TYR	156	0.485	-0.062	-10.480	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1177	CE	TYR	156	-0.450	0.201	-11.463	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1178	OH	TYR	156	-0.811	1.513	-11.605	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1179	C	TYR	156	2.544	-3.525	-12.074	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1180	O	TYR	156	2.595	-3.178	-13.236	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1181	N	LEU	157	3.431	-3.338	-11.195	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1182	CA	LEU	157	4.531	-2.289	-11.581	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1183	CB	LEU	157	5.432	-2.099	-10.411	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1184	CD	LEU	157	5.832	-0.700	-10.023	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1185	CD1	LEU	157	4.709	-0.293	-10.069	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1186	CD2	LEU	157	6.208	-0.791	-8.605	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1187	C	LEU	157	5.349	-2.821	-12.736	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1188	O	LEU	157	5.891	-2.081	-13.540	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1189	N	HIS	158	5.514	-4.109	-12.905	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1190	CA	HIS	158	6.259	-4.639	-14.028	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1191	CB	HIS	158	6.850	-5.970	-13.665	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1192	CD	HIS	158	7.856	-5.334	-12.582	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1193	CD2	HIS	158	8.360	-6.498	-11.767	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1194	ND1	HIS	158	8.395	-4.370	-12.352	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1195	CE1	HIS	158	9.209	-4.812	-11.260	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1196	CE2	HIS	158	9.188	-6.086	-10.974	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1197	C	HIS	158	5.452	-6.981	-15.296	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1198	O	HIS	158	6.057	-4.713	-16.375	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1199	N	SER	159	4.112	-5.008	-15.190	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1200	CA	SER	159	3.238	-5.174	-16.373	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1201	CB	SER	159	1.858	-5.491	-16.005	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1202	CD	SER	159	1.435	-4.464	-15.164	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1203	C	SER	159	3.216	-3.889	-17.170	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1204	N	LEU	160	3.347	-3.912	-16.395	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1205	CA	LEU	160	3.573	-2.776	-15.419	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1206	CB	LEU	160	3.174	-1.504	-17.076	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1207	CD	LEU	160	3.573	-0.323	-16.107	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1208	C	LEU	160	1.931	0.812	-16.139	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1209	CD1	LEU	160	0.754	-0.946	-16.568	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1210	CD2	LEU	160	1.744	0.594	-16.750	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1211	C	LEU	160	5.030	-1.241	-17.538	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1212	O	LEU	160	5.369	-0.109	-17.869	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1213	N	ASP	161	5.885	-2.264	-17.587	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1214	CA	ASP	161	7.321	-2.184	-17.811	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1215	CB	ASP	161	7.314	-1.919	-18.329	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1216	CD	ASP	161	7.428	-3.242	-20.132	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1217	CD1	ASP	161	6.434	-3.353	-20.080	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1218	CD2	ASP	161	8.338	-4.134	-19.996	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1219	C	ASP	161	8.223	-1.257	-17.020	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1220	O	ASP	161	9.382	-0.957	-17.358	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1221	N	LEU	162	7.677	-0.932	-15.837	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1222	CA	LEU	162	8.346	-0.133	-14.865	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1223	CB	LEU	162	7.376	0.501	-13.697	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1224	CC	LEU	162	6.557	1.762	-14.341	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1225	CD1	LEU	162	6.716	2.251	-15.513	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1226	CD2	LEU	162	5.100	1.486	-13.911	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1227	C	LEU	162	9.180	-1.161	-14.146	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1228	O	LEU	162	8.828	-2.378	-16.071	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1229	N	HIS	163	10.254	-0.594	-13.639	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1230	CA	HIS	163	11.292	-1.228	-12.758	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1231	CB	HIS	163	12.160	-1.694	-13.650	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60
1232	CD	HIS	163	12.870	-0.565	-14.619	1.00	17.60	10.750	0.462	-1.099	-12.721	1.00	17.60

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1298	OE2	GIU	170	16.071	1.952	5.052	1.00	17.60	ATOM	1353	N	TYR	179	-1.598	-2.086	6.729	1.00	17.60
1299	C	GIU	170	11.215	3.163	2.791	1.00	17.60	ATOM	1354	CA	TYR	179	-1.216	-2.498	4.866	1.00	17.60
1300	GIU	170	10.130	3.518	4.234	1.00	17.60	ATOM	1355	CB	TYR	179	-2.422	-2.392	3.914	1.00	17.60	
1301	N	ASN	171	11.320	2.788	2.312	1.00	17.60	ATOM	1356	CO	TYR	179	-3.466	-3.507	3.962	1.00	17.60
1302	CA	ASN	171	10.786	3.022	3.499	1.00	17.60	ATOM	1357	CD1	TYR	179	-3.534	-4.387	2.893	1.00	17.60
1303	CB	ASN	171	10.843	2.505	0.192	1.00	17.60	ATOM	1358	CE1	TYR	179	-4.672	-5.397	2.843	1.00	17.60
1304	CO	ASN	171	11.742	3.632	-0.370	1.00	17.60	ATOM	1359	CD2	TYR	179	-4.364	-3.648	5.007	1.00	17.60
1305	OD1	ASN	171	12.265	4.468	0.357	1.00	17.60	ATOM	1360	CE2	TYR	179	-5.315	-4.654	4.966	1.00	17.60
1306	ND2	ASN	171	12.026	3.594	-1.844	1.00	17.60	ATOM	1370	CE7	TYR	179	-5.352	-5.510	3.884	1.00	17.60
1307	C	ASN	171	0.879	2.436	1.628	1.00	17.60	ATOM	1372	OH	TYR	179	-6.287	-6.524	3.807	1.00	17.60
1308	O	ASN	171	0.066	2.585	0.708	1.00	17.60	ATOM	1373	C	TYR	179	-0.259	-1.523	4.377	1.00	17.60
1309	K	LEU	172	6.475	1.826	2.740	1.00	17.60	ATOM	1374	O	TYR	179	-0.275	-0.368	4.854	1.00	17.60
1310	CA	LEU	172	7.169	1.223	2.798	1.00	17.60	ATOM	1375	N	ILE	180	0.615	-1.928	3.447	1.00	17.60
1311	CB	LEU	172	7.362	-0.282	2.998	1.00	17.60	ATOM	1376	CA	ILE	180	1.719	-1.038	3.116	1.00	17.60
1312	CG	LEU	172	6.137	-1.105	2.041	1.00	17.60	ATOM	1377	CB	ILE	180	3.079	-1.813	2.709	1.00	17.60
1313	CD1	LEU	172	6.440	-2.467	2.555	1.00	17.60	ATOM	1378	CG2	ILE	180	3.542	-2.504	4.032	1.00	17.60
1314	CD2	LEU	172	7.342	-1.141	0.851	1.00	17.60	ATOM	1379	CG3	ILE	180	2.964	-2.837	1.754	1.00	17.60
1315	C	LEU	172	6.475	1.878	3.980	1.00	17.60	ATOM	1380	CD1	ILE	180	3.233	-2.787	0.352	1.00	17.60
1316	O	LEU	173	7.055	1.795	5.034	1.00	17.60	ATOM	1381	C	ILE	180	1.328	-0.153	1.931	1.00	17.60
1317	N	LEU	173	5.317	2.530	3.042	1.00	17.60	ATOM	1382	O	ILE	180	0.239	-0.366	1.310	1.00	17.60
1318	CA	LEU	173	4.621	3.203	4.911	1.00	17.60	ATOM	1383	K	GLN	181	2.330	0.920	1.821	1.00	17.60
1319	CB	LEU	173	4.125	4.557	4.496	1.00	17.60	ATOM	1384	CA	GLN	181	2.305	1.933	0.788	1.00	17.60
1320	CD	LEU	173	5.064	5.707	4.199	1.00	17.60	ATOM	1385	CB	GLN	181	1.315	3.078	1.219	1.00	17.60
1321	CD1	LEU	173	4.317	7.005	6.153	1.00	17.60	ATOM	1386	CD	GLN	181	-0.092	2.598	1.296	1.00	17.60
1322	CD2	LEU	173	3.422	5.823	5.316	1.00	17.60	ATOM	1387	CD	GLN	181	-1.036	3.533	0.621	1.00	17.60
1323	C	LEU	173	2.822	1.413	5.337	1.00	17.60	ATOM	1388	OE1	GLN	181	-0.871	4.749	0.672	1.00	17.60
1324	O	LEU	174	3.010	2.806	4.443	1.00	17.60	ATOM	1389	NE2	GLN	181	-2.070	3.051	-0.039	1.00	17.60
1325	N	ILE	174	3.010	2.393	6.625	1.00	17.60	ATOM	1390	C	GLN	181	3.514	2.414	0.471	1.00	17.60
1326	CA	ILE	174	1.925	1.526	7.114	1.00	17.60	ATOM	1391	O	GLN	181	4.204	3.029	1.273	1.00	17.60
1327	CB	ILE	174	2.240	1.013	8.504	1.00	17.60	ATOM	1392	H	VAL	182	3.925	1.970	-0.738	1.00	17.60
1328	CD1	ILE	174	1.087	0.163	8.966	1.00	17.60	ATOM	1393	CA	VAL	182	5.164	2.312	-1.407	1.00	17.60
1329	CD2	ILE	174	3.589	0.266	8.532	1.00	17.60	ATOM	1394	CB	VAL	182	5.381	1.435	-2.594	1.00	17.60
1330	CD1	ILE	174	3.832	-0.952	7.620	1.00	17.60	ATOM	1395	CG1	VAL	182	6.841	1.481	-2.801	1.00	17.60
1331	C	ILE	174	0.687	2.340	7.179	1.00	17.60	ATOM	1396	CG2	VAL	182	4.983	0.018	-2.615	1.00	17.60
1332	O	ILE	175	0.602	3.453	7.668	1.00	17.60	ATOM	1397	C	VAL	182	5.113	3.768	-1.868	1.00	17.60
1333	CA	ASP	175	-0.466	1.893	6.714	1.00	17.60	ATOM	1398	O	VAL	182	4.327	4.292	-2.654	1.00	17.60
1334	CB	ASP	175	-1.617	2.238	6.819	1.00	17.60	ATOM	1399	N	THR	183	5.978	4.464	-1.196	1.00	17.60
1335	CG	ASP	175	-2.360	2.717	5.560	1.00	17.60	ATOM	1400	CA	THR	183	6.230	5.878	-1.350	1.00	17.60
1336	CG1	ASP	175	-2.820	1.380	5.193	1.00	17.60	ATOM	1401	CB	THR	183	6.387	6.401	0.101	1.00	17.60
1337	OD1	ASP	175	-2.679	1.100	4.034	1.00	17.60	ATOM	1402	CG1	THR	183	7.663	5.870	0.743	1.00	17.60
1338	OD2	ASP	175	-3.315	0.648	5.999	1.00	17.60	ATOM	1403	CG2	THR	183	7.499	5.949	-2.214	1.00	17.60
1339	O	ASP	175	-2.594	2.551	7.339	1.00	17.60	ATOM	1404	C	THR	183	8.115	4.907	-2.484	1.00	17.60
1340	N	ASP	175	-2.246	1.740	8.772	1.00	17.60	ATOM	1405	O	THR	183	8.115	4.907	-2.484	1.00	17.60
1341	CA	GLN	176	-3.845	3.062	7.983	1.00	17.60	ATOM	1406	N	ASP	184	7.928	7.121	-2.695	1.00	17.60
1342	CB	GLN	176	-6.714	2.924	9.098	1.00	17.60	ATOM	1407	CA	ASP	184	9.237	7.211	-3.312	1.00	17.60
1343	CG	GLN	176	-5.498	4.137	9.197	1.00	17.60	ATOM	1408	CB	ASP	184	10.358	7.007	-2.797	1.00	17.60
1344	CG1	GLN	176	-6.594	4.105	10.456	1.00	17.60	ATOM	1409	CG	ASP	184	11.496	7.629	-2.733	1.00	17.60
1345	CG2	GLN	176	-7.823	5.030	10.567	1.00	17.60	ATOM	1410	CG1	ASP	184	12.108	8.669	-2.169	1.00	17.60
1346	OD1	GLN	176	-7.785	5.989	11.347	1.00	17.60	ATOM	1411	CG2	ASP	184	12.327	7.064	-3.641	1.00	17.60
1347	NE2	GLN	176	-8.960	4.780	9.885	1.00	17.60	ATOM	1412	C	ASP	184	9.512	6.249	-4.437	1.00	17.60
1348	O	GLN	176	-5.589	1.690	8.895	1.00	17.60	ATOM	1413	O	ASP	184	9.890	5.099	-4.301	1.00	17.60
1349	N	GLN	177	-6.818	1.669	8.819	1.00	17.60	ATOM	1414	N	PHE	185	9.406	6.906	-5.558	1.00	17.60
1350	CA	GLN	177	-5.374	-0.709	8.432	1.00	17.60	ATOM	1415	CA	PHE	185	9.512	6.360	-6.849	1.00	17.60
1351	CB	GLN	177	-6.184	0.823	7.082	1.00	17.60	ATOM	1416	CB	PHE	185	8.399	6.721	-7.705	1.00	17.60
1352	CG	GLN	177	-7.706	-0.598	6.921	1.00	17.60	ATOM	1417	CG	PHE	185	7.425	5.694	-7.618	1.00	17.60
1353	CD	GLN	177	-8.531	-1.250	6.256	1.00	17.60	ATOM	1418	CG1	PHE	185	7.330	4.778	-8.633	1.00	17.60
1354	CD1	GLN	177	-9.253	-1.506	5.275	1.00	17.60	ATOM	1419	CG2	PHE	185	6.823	5.352	-6.419	1.00	17.60
1355	CD2	GLN	177	-8.531	-1.250	6.256	1.00	17.60	ATOM	1420	CE1	PHE	185	6.631	5.635	-8.532	1.00	17.60
1356	NE2	GLN	177	-8.531	-1.506	5.275	1.00	17.60	ATOM	1421	CE2	PHE	185	6.116	4.196	-6.284	1.00	17.60
1357	O	GLN	177	-4.145	-1.631	8.383	1.00	17.60	ATOM	1422	C	PHE	185	6.034	3.166	-7.319	1.00	17.60
1358	C	GLN	177	-4.334	-2.808	8.001	1.00	17.60	ATOM	1423	O	PHE	185	10.888	6.818	-7.409	1.00	17.60
1359	N	GLY	178	-2.891	-1.229	8.688	1.00	17.60	ATOM	1424	C	PHE	185	11.010	6.965	-8.622	1.00	17.60
1360	CA	GLY	178	-1.720	-2.121	8.692	1.00	17.60	ATOM	1425	N	GLY	186	11.794	7.360	-6.533	1.00	17.60
1361	C	GLY	178	-1.259	-2.752	7.366	1.00	17.60	ATOM	1426	CA	GLY	186	13.017	8.013	-6.900	1.00	17.60
1362	O	GLY	178	-0.575	-2.799	7.418	1.00	17.60	ATOM	1427	C	GLY	186	13.804	7.355	-7.955	1.00	17.60

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1428	O	GLY	166	14.364	7.915	-8.807	3.00	17.60	19.008	-0.265	-23.816	1.00	17.60
1429	N	PHE	167	13.097	5.919	-8.007	3.00	17.60	18.297	-2.151	-19.039	1.00	17.60
1430	CA	PHE	167	14.601	5.365	-8.327	3.00	17.60	18.018	-1.255	-20.639	1.00	17.60
1431	CB	PHE	167	15.637	4.456	-8.507	3.00	17.60	18.878	-1.851	-18.683	1.00	17.60
1432	CG	PHE	167	16.700	5.284	-7.954	3.00	17.60	19.282	-0.494	-18.361	1.00	17.60
1433	CDI	PHE	167	17.129	6.419	-8.520	3.00	17.60	18.109	0.233	-17.582	1.00	17.60
1434	CD2	PHE	167	17.191	4.916	-8.761	3.00	17.60	18.490	3.592	-17.454	3.00	17.60
1435	CE1	PHE	167	18.042	7.225	-7.809	3.00	17.60	17.799	-0.356	-16.218	3.00	17.60
1436	CE2	PHE	167	18.101	5.721	-6.110	3.00	17.60	20.561	-0.687	-17.539	3.00	17.60
1437	CL	PHE	167	18.537	6.889	-6.670	3.00	17.60	20.950	-1.823	-17.194	3.00	17.60
1438	C	PHE	167	13.699	4.673	-10.094	3.00	17.60	21.207	0.447	-17.274	3.00	17.60
1439	O	PHE	167	14.208	3.926	-10.910	3.00	17.60	22.534	0.426	-16.720	3.00	17.60
1440	N	ALA	168	12.385	6.917	-10.091	3.00	17.60	23.541	-0.692	-17.876	3.00	17.60
1441	CA	ALA	168	11.455	4.171	-10.917	3.00	17.60	23.765	-0.441	-18.863	3.00	17.60
1442	CB	ALA	168	10.072	4.600	-10.621	3.00	17.60	23.156	-0.589	-20.050	3.00	17.60
1443	C	ALA	168	11.737	4.425	-12.373	3.00	17.60	23.633	-1.810	-20.452	3.00	17.60
1444	ALA	168	12.142	5.570	-12.669	3.00	17.60	22.265	0.119	-20.801	3.00	17.60	
1445	N	LYS	169	11.611	3.537	-13.330	3.00	17.60	24.571	-1.492	-18.571	3.00	17.60
1446	CA	LYS	169	11.986	3.631	-14.684	3.00	17.60	24.439	-2.321	-19.557	3.00	17.60
1447	CB	LYS	169	13.522	3.536	-14.873	3.00	17.60	23.449	-2.394	-21.635	3.00	17.60
1448	CG	LYS	169	14.124	3.832	-16.269	3.00	17.60	21.067	-0.465	-22.017	3.00	17.60
1449	CD	LYS	169	15.612	3.934	-16.303	3.00	17.60	22.355	-1.708	-22.431	3.00	17.60
1450	CE	LYS	169	16.128	4.237	-17.450	3.00	17.60	22.823	1.384	-19.562	3.00	17.60
1451	NE	LYS	169	15.663	5.500	-18.226	3.00	17.60	23.992	1.518	-15.192	3.00	17.60
1452	C	LYS	169	11.124	2.966	-15.969	3.00	17.60	21.863	2.023	-14.916	3.00	17.60
1453	O	LYS	169	10.911	1.801	-15.203	3.00	17.60	22.282	2.964	-13.888	3.00	17.60
1454	N	ARG	190	10.559	3.539	-16.439	3.00	17.60	21.137	3.913	-13.599	3.00	17.60
1455	CA	ARG	190	9.003	2.726	-17.356	3.00	17.60	20.785	4.568	-14.400	3.00	17.60
1456	CB	ARG	190	8.757	3.613	-18.208	3.00	17.60	19.404	4.308	-15.392	3.00	17.60
1457	CG	ARG	190	7.847	2.691	-19.029	3.00	17.60	19.303	6.976	-16.676	3.00	17.60
1458	CD	ARG	190	6.387	3.051	-19.316	3.00	17.60	19.173	2.015	-15.341	3.00	17.60
1459	CE	ARG	190	6.243	2.826	-20.731	3.00	17.60	18.398	5.000	-14.424	3.00	17.60
1460	CZ	ARG	190	5.277	2.166	-21.314	3.00	17.60	21.528	4.965	-12.542	3.00	17.60
1461	NH1	ARG	190	6.285	1.604	-20.639	3.00	17.60	22.722	2.265	-12.614	3.00	17.60
1462	NH2	ARG	190	5.353	2.118	-22.627	3.00	17.60	22.056	1.332	-12.218	3.00	17.60
1463	C	ARG	190	10.819	2.181	-18.542	3.00	17.60	23.620	2.581	-13.937	3.00	17.60
1464	O	ARG	190	11.256	2.944	-19.388	3.00	17.60	24.011	1.956	-10.630	3.00	17.60
1465	N	VAL	191	11.324	0.968	-18.484	3.00	17.60	25.457	1.928	-10.093	3.00	17.60
1466	CA	VAL	191	12.278	0.954	-19.480	3.00	17.60	26.704	-0.165	-9.175	3.00	17.60
1467	CB	VAL	191	13.690	0.066	-19.118	3.00	17.60	26.508	1.905	-7.827	3.00	17.60
1468	CGI	VAL	191	14.404	1.268	-18.194	3.00	17.60	23.235	2.868	-9.714	3.00	17.60
1469	CD2	VAL	191	13.045	-0.841	-17.946	3.00	17.60	22.164	2.365	-9.157	3.00	17.60
1470	C	VAL	191	11.722	-0.691	-20.031	3.00	17.60	21.362	3.034	-8.179	3.00	17.60
1471	O	VAL	191	11.328	-1.638	-19.334	3.00	17.60	20.494	4.098	-8.872	3.00	17.60
1472	N	LYS	192	11.757	-0.678	-21.355	3.00	17.60	19.489	3.416	-10.182	3.00	17.60
1473	CA	LYS	192	11.285	-1.863	-22.010	3.00	17.60	20.513	1.939	-7.527	3.00	17.60
1474	CB	LYS	192	10.357	-1.481	-23.344	3.00	17.60	20.264	0.867	-8.150	3.00	17.60
1475	CG	LYS	192	9.499	-2.664	-23.563	3.00	17.60	20.212	2.238	-6.235	3.00	17.60
1476	CD	LYS	192	8.442	-2.034	-24.415	3.00	17.60	19.421	1.400	-5.333	3.00	17.60
1477	CE	LYS	192	7.131	-1.894	-23.730	3.00	17.60	20.619	2.665	-3.762	3.00	17.60
1478	NE	LYS	192	7.259	-1.140	-22.516	3.00	17.60	19.715	0.871	-2.871	3.00	17.60
1479	C	LYS	192	12.492	-2.606	-22.489	3.00	17.60	20.327	1.170	-0.488	3.00	17.60
1480	O	LYS	192	12.465	-3.040	-23.629	3.00	17.60	19.230	1.276	-0.488	3.00	17.60
1481	N	GLY	193	13.521	-2.824	-21.676	3.00	17.60	20.619	2.665	-3.762	3.00	17.60
1482	CA	GLY	193	14.720	-3.574	-22.136	3.00	17.60	20.880	-1.091	-1.352	3.00	17.60
1483	C	GLY	193	15.583	-3.781	-20.927	3.00	17.60	22.546	0.159	-0.914	3.00	17.60
1484	O	GLY	193	15.118	-4.209	-19.852	3.00	17.60	23.092	1.489	-0.403	3.00	17.60
1485	N	ARG	194	16.823	-3.393	-21.137	3.00	17.60	22.616	-0.360	-0.870	3.00	17.60
1486	CA	ARG	194	17.878	-3.572	-20.168	3.00	17.60	26.570	1.134	-0.345	3.00	17.60
1487	CB	ARG	194	19.031	-4.371	-20.795	3.00	17.60					
1488	CG	ARG	194	18.834	-4.889	-22.302	3.00	17.60					
1489	CD	ARG	194	19.128	-5.977	-23.579	3.00	17.60					
1490	CE	ARG	194	18.904	-5.529	-23.379	3.00	17.60					
1491	CZ	ARG	194	19.206	-1.549	-24.256	3.00	17.60					
1492	NH1	ARG	194	19.757	-1.773	-25.484	3.00	17.60					

1558	C	PRO	201	21.157	-2.268	-0.480	1.00	17.60	ATOM	1623	CD1	ILE	210	29.667	-5.033	-6.015	1.00	17.60
1559	O	PRO	201	21.356	-3.157	-1.270	1.00	17.60	ATOM	1624	C	ILE	210	28.381	-4.708	-9.766	1.00	17.60
1560	N	GLU	201	21.551	-2.591	0.646	1.00	17.60	ATOM	1625	O	ILE	210	29.454	-4.186	-10.090	1.00	17.60
1561	CA	GLU	201	21.758	-3.989	0.997	1.00	17.60	ATOM	1626	CA	LEU	211	28.106	-5.964	-10.156	1.00	17.60
1562	CB	GLU	201	21.902	-4.183	2.507	1.00	17.60	ATOM	1627	CB	LEU	211	29.045	-6.491	-11.203	1.00	17.60
1563	CD	GLU	201	21.311	-3.024	3.263	1.00	17.60	ATOM	1628	CD	LEU	211	28.979	-7.970	-11.168	1.00	17.60
1564	CE	GLU	201	21.311	-3.024	3.263	1.00	17.60	ATOM	1629	CE	LEU	211	30.036	-8.790	-10.509	1.00	17.60
1565	CE1	GLU	201	21.769	-2.189	4.899	1.00	17.60	ATOM	1630	CE1	LEU	211	31.005	-7.998	-9.705	1.00	17.60
1566	E2	GLU	201	21.597	-1.002	3.099	1.00	17.60	ATOM	1631	CE2	LEU	211	29.239	-9.772	-9.709	1.00	17.60
1567	C	GLU	201	21.155	-4.641	0.199	1.00	17.60	ATOM	1632	C	LEU	211	28.310	-5.930	-12.426	1.00	17.60
1568	O	GLU	201	20.910	-5.851	0.231	1.00	17.60	ATOM	1633	O	LEU	211	29.566	-6.706	-13.033	1.00	17.60
1569	N	TYR	204	20.484	-3.743	-0.506	1.00	17.60	ATOM	1634	N	TYR	212	28.460	-4.643	-12.792	1.00	17.60
1570	CA	TYR	204	19.383	-4.131	-1.320	1.00	17.60	ATOM	1635	CA	TYR	212	27.570	-3.987	-13.749	1.00	17.60
1571	CB	TYR	204	18.334	-2.999	-1.302	1.00	17.60	ATOM	1636	CB	TYR	212	27.958	-2.495	-13.832	1.00	17.60
1572	CD	TYR	204	17.497	-3.272	-0.063	1.00	17.60	ATOM	1637	CD	TYR	212	29.350	-2.146	-13.703	1.00	17.60
1573	CD1	TYR	204	16.576	-4.312	-0.091	1.00	17.60	ATOM	1638	CD1	TYR	212	27.352	-4.308	-15.174	1.00	17.60
1574	CE1	TYR	204	15.913	-4.696	1.078	1.00	17.60	ATOM	1639	CE1	TYR	212	28.027	-4.155	-16.132	1.00	17.60
1575	CE2	TYR	204	17.744	-2.601	1.142	1.00	17.60	ATOM	1640	CE2	TYR	212	26.355	-5.367	-15.333	1.00	17.60
1576	CE3	TYR	204	17.094	-2.995	2.309	1.00	17.60	ATOM	1641	CE3	TYR	212	26.003	-6.107	-16.542	1.00	17.60
1577	C2	TYR	204	16.201	-4.043	2.285	1.00	17.60	ATOM	1642	C2	TYR	212	27.382	-8.055	-15.687	1.00	17.60
1578	OH	TYR	204	15.501	-4.452	3.408	1.00	17.60	ATOM	1643	OH	TYR	212	27.170	-9.314	-14.813	1.00	17.60
1579	O	TYR	204	19.877	-4.427	-2.708	1.00	17.60	ATOM	1644	O	TYR	212	26.164	-10.378	-15.409	1.00	17.60
1580	N	LEU	205	19.171	-5.107	-3.439	1.00	17.60	ATOM	1645	N	LEU	213	26.696	-11.668	-14.795	1.00	17.60
1581	CA	LEU	205	21.072	-3.985	1.124	1.00	17.60	ATOM	1646	CA	LEU	213	24.746	-6.439	-18.366	1.00	17.60
1582	CB	LEU	205	22.960	-3.470	-6.649	1.00	17.60	ATOM	1647	CB	LEU	213	24.655	-5.740	-17.235	1.00	17.60
1583	CC	LEU	205	21.328	-3.133	-5.266	1.00	17.60	ATOM	1648	CC	LEU	213	23.926	-4.657	-16.893	1.00	17.60
1584	CD	LEU	205	21.859	-3.172	-3.282	1.00	17.60	ATOM	1649	CD	LEU	213	22.655	-4.464	-17.551	1.00	17.60
1585	CE	LEU	205	24.309	-1.460	-4.622	1.00	17.60	ATOM	1650	CE	LEU	213	21.866	-5.376	-16.839	1.00	17.60
1586	CD2	LEU	205	21.559	-5.594	-4.916	1.00	17.60	ATOM	1651	CD2	LEU	213	21.033	-6.585	-16.603	1.00	17.60
1587	C	LEU	205	21.856	-6.889	-4.127	1.00	17.60	ATOM	1652	C	LEU	213	20.614	-4.680	-16.458	1.00	17.60
1588	N	ALA	206	21.087	-5.038	-6.143	1.00	17.60	ATOM	1653	N	ALA	215	19.846	-5.310	-15.619	1.00	17.60
1589	CA	ALA	206	21.160	-7.133	-6.746	1.00	17.60	ATOM	1654	CA	ALA	215	19.937	-4.883	-14.156	1.00	17.60
1590	CB	ALA	206	20.188	-7.216	-7.650	1.00	17.60	ATOM	1655	CB	ALA	215	20.034	-3.384	-13.771	1.00	17.60
1591	CC	ALA	206	22.556	-7.266	-7.296	1.00	17.60	ATOM	1656	CC	ALA	215	18.955	-2.853	-13.164	1.00	17.60
1592	C	ALA	206	23.043	-6.355	-7.645	1.00	17.60	ATOM	1657	C	ALA	215	18.911	-1.569	-12.743	1.00	17.60
1593	O	ALA	206	23.242	-8.404	-7.472	1.00	17.60	ATOM	1658	O	ALA	215	21.120	-2.943	-13.960	1.00	17.60
1594	CA	PRO	207	22.622	-9.695	-7.672	1.00	17.60	ATOM	1659	CA	PRO	215	21.081	-1.228	-13.542	1.00	17.60
1595	CB	PRO	207	24.675	-8.524	-7.627	1.00	17.60	ATOM	1660	CB	PRO	215	19.949	-0.758	-12.930	1.00	17.60
1596	CC	PRO	207	25.063	-9.914	-7.546	1.00	17.60	ATOM	1661	CC	PRO	215	19.756	0.537	-12.505	1.00	17.60
1597	C	PRO	207	23.756	-10.662	-7.352	1.00	17.60	ATOM	1662	C	PRO	215	18.259	-4.930	-16.107	1.00	17.60
1598	CA	PRO	207	26.123	-7.789	-9.592	1.00	17.60	ATOM	1663	CA	PRO	215	18.007	-4.081	-16.966	1.00	17.60
1599	CB	PRO	207	24.034	-8.322	-10.214	1.00	17.60	ATOM	1664	CB	PRO	215	17.315	-5.574	-15.675	1.00	17.60
1600	O	PRO	207	24.253	-7.872	-11.546	1.00	17.60	ATOM	1665	O	PRO	215	15.905	-5.309	-15.915	1.00	17.60
1601	CA	GLU	208	23.118	-8.277	-12.480	1.00	17.60	ATOM	1666	CA	GLU	216	15.806	-6.618	-15.901	1.00	17.60
1602	CB	GLU	208	21.773	-7.703	-12.166	1.00	17.60	ATOM	1667	CB	GLU	216	16.398	-7.224	-16.334	1.00	17.60
1603	CC	GLU	208	20.791	-8.657	-11.532	1.00	17.60	ATOM	1668	CC	GLU	216	16.693	-8.078	-15.132	1.00	17.60
1604	C	GLU	208	19.582	-8.409	-11.672	1.00	17.60	ATOM	1669	C	GLU	216	16.658	-8.886	-17.198	1.00	17.60
1605	CA	GLU	208	21.237	-9.593	-10.890	1.00	17.60	ATOM	1670	CA	GLU	216	15.153	-5.753	-14.669	1.00	17.60
1606	CB	GLU	208	24.369	-6.368	-11.527	1.00	17.60	ATOM	1671	CB	GLU	216	15.613	-5.836	-13.540	1.00	17.60
1607	C	GLU	208	24.954	-5.809	-12.441	1.00	17.60	ATOM	1672	C	GLU	216	12.852	-6.326	-13.994	1.00	17.60
1608	O	GLU	208	23.881	-5.605	-10.583	1.00	17.60	ATOM	1673	O	GLU	216	12.612	-7.037	-14.631	1.00	17.60
1609	N	ILE	209	24.037	-4.176	-10.681	1.00	17.60	ATOM	1674	N	ILE	217	11.066	-5.144	-16.061	1.00	17.60
1610	CA	ILE	209	23.019	-3.466	-9.826	1.00	17.60	ATOM	1675	CA	ILE	217	10.678	-6.632	-16.021	1.00	17.60
1611	CB	ILE	209	21.832	-1.955	-9.989	1.00	17.60	ATOM	1676	CB	ILE	217	11.315	-4.316	-17.221	1.00	17.60
1612	CC	ILE	209	21.656	-3.963	-10.252	1.00	17.60	ATOM	1677	CC	ILE	217	12.779	-7.246	-17.387	1.00	17.60
1613	CD	ILE	209	20.589	-3.274	-9.398	1.00	17.60	ATOM	1678	CD	ILE	217	13.337	-7.402	-17.387	1.00	17.60
1614	CE	ILE	209	25.419	-2.790	-10.264	1.00	17.60	ATOM	1679	CE	ILE	217	12.702	-7.150	-17.387	1.00	17.60
1615	C	ILE	209	25.930	-2.849	-10.853	1.00	17.60	ATOM	1680	C	ILE	217	14.381	-8.079	-13.909	1.00	17.60
1616	CA	ILE	209	26.085	-4.445	-9.290	1.00	17.60	ATOM	1681	CA	ILE	218	14.771	-9.089	-12.135	1.00	17.60
1617	CB	ILE	210	27.371	-3.979	-8.925	1.00	17.60	ATOM	1682	CB	ILE	218	15.675	-10.083	-12.804	1.00	17.60
1618	CC	ILE	210	27.695	-4.023	-7.360	1.00	17.60	ATOM	1683	CC	ILE	218	15.462	-8.567	-10.886	1.00	17.60
1619	C	ILE	210	26.413	-4.151	-6.511	1.00	17.60	ATOM	1684	C	ILE	218	15.348	-9.254	-9.850	1.00	17.60
1620	CA	ILE	210	28.514	-5.238	-7.062	1.00	17.60	ATOM	1685	CA	ILE	219	15.901	-7.300	-10.952	1.00	17.60
1621	CB	ILE	210						ATOM	1686	CB	ILE	219					
1622	CC	ILE	210						ATOM	1687	CC	ILE	219					

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1688	CA	VAL	219	16.693	-6.689	-9.787	1.00	17.60	15.122	-0.756	2.945	1.00	17.60
1689	CB	VAL	219	17.088	-5.299	-9.997	1.00	17.60	13.915	-7.855	1.335	1.00	17.60
1690	CC1	VAL	219	18.182	-5.527	-10.931	1.00	17.60	12.927	-7.404	1.335	1.00	17.60
1691	CC2	VAL	219	16.118	-4.231	-10.769	1.00	17.60	11.758	-6.862	1.546	1.00	17.60
1692	C	VAL	219	15.420	-6.511	-8.736	1.00	17.60	10.845	-6.168	2.472	1.00	17.60
1693	O	VAL	219	15.582	-6.634	-7.532	1.00	17.60	10.431	-4.853	2.831	1.00	17.60
1694	N	ASP	220	14.182	-6.314	-9.163	1.00	17.60	9.557	-5.891	1.814	1.00	17.60
1695	CA	ASP	220	13.121	-6.116	-8.224	1.00	17.60	12.429	-8.573	3.039	1.00	17.60
1696	CB	ASP	220	11.862	-5.622	-9.950	1.00	17.60	12.454	-8.492	4.255	1.00	17.60
1697	CC1	ASP	220	11.891	-4.095	-9.182	1.00	17.60	12.066	-9.708	2.458	1.00	17.60
1698	CC2	ASP	220	12.629	-3.655	-10.036	1.00	17.60	11.512	-10.836	3.225	1.00	17.60
1699	O	ASP	220	11.183	-3.318	-8.536	1.00	17.60	13.123	-12.015	2.287	1.00	17.60
1700	C	ASP	220	12.055	-7.400	-7.693	1.00	17.60	10.621	-13.152	3.135	1.00	17.60
1701	O	ASP	220	12.293	-7.339	-6.404	1.00	17.60	9.973	-11.655	1.361	1.00	17.60
1702	N	TRP	221	13.275	-8.568	-7.968	1.00	17.60	9.616	-12.704	0.299	1.00	17.60
1703	CA	TRP	221	12.092	-9.736	-7.235	1.00	17.60	12.505	-11.325	6.256	1.00	17.60
1704	CB	TRP	221	12.018	-10.867	-8.137	1.00	17.60	12.083	-11.642	5.341	1.00	17.60
1705	C	TRP	221	13.309	-10.554	-8.782	1.00	17.60	13.797	-11.388	4.925	1.00	17.60
1706	CC2	TRP	221	10.063	-10.346	-8.240	1.00	17.60	14.755	-11.843	4.937	1.00	17.60
1707	CC3	TRP	221	9.339	-10.089	-9.383	1.00	17.60	16.053	-12.035	6.312	1.00	17.60
1708	CC3	TRP	221	9.313	-10.317	-7.052	1.00	17.60	17.190	-12.622	5.093	1.00	17.60
1709	CC1	TRP	221	11.349	-10.436	-10.106	1.00	17.60	17.769	-12.048	6.189	1.00	17.60
1710	CC1	TRP	221	10.126	-10.155	-10.432	1.00	17.60	18.078	-12.635	6.713	1.00	17.60
1711	CC2	TRP	221	7.994	-9.610	-9.433	1.00	17.60	17.708	-13.784	4.578	1.00	17.60
1712	CC3	TRP	221	8.005	-10.037	-7.070	1.00	17.60	18.803	-14.386	5.125	1.00	17.60
1713	CC3	TRP	221	7.312	-9.786	-8.271	1.00	17.60	19.388	-13.795	6.194	1.00	17.60
1714	C	TRP	221	13.947	-10.105	-6.287	1.00	17.60	20.573	-14.356	6.616	1.00	17.60
1715	O	TRP	222	13.571	-10.661	-5.248	1.00	17.60	14.891	-10.782	6.046	1.00	17.60
1716	N	TRP	222	15.207	-9.748	-6.595	1.00	17.60	14.986	-11.107	7.238	1.00	17.60
1717	CA	TRP	222	16.292	-9.946	-5.828	1.00	17.60	14.916	-9.531	5.676	1.00	17.60
1718	CB	TRP	222	17.644	-9.488	-6.138	1.00	17.60	15.215	-8.438	6.591	1.00	17.60
1719	C	TRP	222	18.922	-9.618	-5.173	1.00	17.60	15.285	-7.203	5.656	1.00	17.60
1720	CC2	TRP	222	19.762	-10.596	-5.142	1.00	17.60	15.172	-5.709	6.007	1.00	17.60
1721	CC2	TRP	222	20.642	-10.127	-4.192	1.00	17.60	16.309	-6.864	6.559	1.00	17.60
1722	CC3	TRP	222	20.024	-11.777	-5.757	1.00	17.60	17.052	-4.267	5.813	1.00	17.60
1723	CC1	TRP	222	19.125	-8.637	-4.215	1.00	17.60	16.353	-4.754	7.773	1.00	17.60
1724	CC1	TRP	222	20.249	-8.957	-3.651	1.00	17.60	14.150	-6.448	7.659	1.00	17.60
1725	CC2	TRP	222	11.779	-10.825	-3.855	1.00	17.60	14.163	-6.521	8.865	1.00	17.60
1726	CC3	TRP	222	21.165	-12.481	-5.423	1.00	17.60	12.910	-6.506	7.203	1.00	17.60
1727	CC2	TRP	222	22.082	-12.018	-4.481	1.00	17.60	11.754	-8.524	8.073	1.00	17.60
1728	C	TRP	222	15.917	-9.088	-4.619	1.00	17.60	10.508	-8.593	7.237	1.00	17.60
1729	O	TRP	222	15.795	-9.674	-3.342	1.00	17.60	10.214	-7.355	6.413	1.00	17.60
1730	N	ALA	223	15.760	-7.761	-4.538	1.00	17.60	8.603	-7.416	5.592	1.00	17.60
1731	CA	ALA	223	15.375	-6.950	-3.399	1.00	17.60	9.024	-6.484	4.278	1.00	17.60
1732	CB	ALA	223	15.127	-5.547	-3.843	1.00	17.60	11.783	-9.700	9.029	1.00	17.60
1733	C	ALA	223	14.115	-7.495	-2.705	1.00	17.60	11.784	-9.434	10.207	1.00	17.60
1734	O	ALA	223	14.101	-7.474	-2.705	1.00	17.60	11.849	-10.972	8.821	1.00	17.60
1735	N	LEU	224	13.069	-8.096	-3.274	1.00	17.60	11.867	-12.131	9.508	1.00	17.60
1736	CA	LEU	224	11.974	-8.685	-2.486	1.00	17.60	11.626	-13.428	8.744	1.00	17.60
1737	CB	LEU	224	10.917	-9.485	-3.241	1.00	17.60	13.133	-12.352	10.303	1.00	17.60
1738	C	LEU	224	9.485	-9.501	-2.844	1.00	17.60	13.076	-12.872	11.426	1.00	17.60
1739	CC1	LEU	224	8.965	-10.816	-3.302	1.00	17.60	14.306	-12.048	9.752	1.00	17.60
1740	CC2	LEU	224	9.261	-9.281	-3.373	1.00	17.60	15.523	-12.212	10.519	1.00	17.60
1741	C	LEU	224	12.550	-9.740	-3.578	1.00	17.60	16.732	-12.390	9.667	1.00	17.60
1742	O	LEU	224	12.169	-9.823	-0.424	1.00	17.60	15.874	-11.065	11.426	1.00	17.60
1743	N	GLY	225	13.476	-10.529	-2.085	1.00	17.60	16.606	-11.314	12.371	1.00	17.60
1744	CA	GLY	225	14.103	-11.542	-3.294	1.00	17.60	15.425	-9.830	11.189	1.00	17.60
1745	C	GLY	225	14.813	-10.920	-0.130	1.00	17.60	15.777	-8.652	12.007	1.00	17.60
1746	O	GLY	225	14.956	-11.510	0.918	1.00	17.60	16.903	-7.776	11.438	1.00	17.60
1747	N	VAL	226	15.224	-9.667	-0.270	1.00	17.60	17.185	-6.645	11.856	1.00	17.60
1748	CA	VAL	226	15.967	-9.059	0.807	1.00	17.60	17.544	-8.336	10.415	1.00	17.60
1749	CB	VAL	226	16.911	-8.010	0.214	1.00	17.60	18.657	-7.705	9.737	1.00	17.60
1750	CC1	VAL	226	17.879	-7.675	1.367	1.00	17.60	19.921	-7.936	10.566	1.00	17.60
1751	CC2	VAL	226	17.847	-8.600	-0.901	1.00	17.60	20.159	-9.387	10.836	1.00	17.60
1752	C	VAL	226	14.987	-6.492	1.772	1.00	17.60	19.731	-9.817	12.013	1.00	17.60

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1818	CE1 TYR	235	19.844	-11.132	32.322	1.00 17.60	1.00 17.60	2.559	1.00 17.60
1819	CD2 TYR	235	20.710	-10.222	8.800	1.00 17.60	1.00 17.60	2.184	1.00 17.60
1820	CE2 TYR	235	20.824	-11.549	10.353	1.00 17.60	1.00 17.60	1.490	1.00 17.60
1821	CE TYR	235	20.378	-11.957	11.393	1.00 17.60	1.00 17.60	0.678	1.00 17.60
1822	OK TYR	235	20.512	-13.250	11.795	1.00 17.60	1.00 17.60	1.493	1.00 17.60
1823	C TYR	235	18.889	-8.172	8.273	1.00 17.60	1.00 17.60	0.859	1.00 17.60
1824	O TYR	235	18.316	-9.177	7.841	1.00 17.60	1.00 17.60	0.777	1.00 17.60
1825	N PRO	236	19.715	-7.471	7.462	1.00 17.60	1.00 17.60	0.416	1.00 17.60
1826	CD PRO	236	20.393	-6.191	7.706	1.00 17.60	1.00 17.60	-0.289	1.00 17.60
1827	CB PRO	236	19.963	-7.777	6.097	1.00 17.60	1.00 17.60	-0.286	1.00 17.60
1828	CB PRO	236	20.765	-6.562	5.646	1.00 17.60	1.00 17.60	0.946	1.00 17.60
1829	CB PRO	236	20.254	-5.412	6.426	1.00 17.60	1.00 17.60	-1.511	1.00 17.60
1830	C PRO	236	20.657	-9.132	6.038	1.00 17.60	1.00 17.60	-2.590	1.00 17.60
1831	O PRO	236	21.267	-9.553	7.025	1.00 17.60	1.00 17.60	0.301	1.00 17.60
1832	N PRO	237	19.919	-9.211	3.665	1.00 17.60	1.00 17.60	-0.390	1.00 17.60
1833	CD PRO	237	21.200	-11.022	4.536	1.00 17.60	1.00 17.60	-3.488	1.00 17.60
1834	CA PRO	237	20.651	-11.392	3.228	1.00 17.60	1.00 17.60	3.385	1.00 17.60
1835	CB PRO	237	19.512	-10.475	2.998	1.00 17.60	1.00 17.60	3.821	1.00 17.60
1836	CG PRO	237	22.707	-10.963	4.672	1.00 17.60	1.00 17.60	4.226	1.00 17.60
1837	C PRO	237	23.426	-11.959	4.649	1.00 17.60	1.00 17.60	5.540	1.00 17.60
1838	O PRO	237	23.063	-9.748	4.015	1.00 17.60	1.00 17.60	5.626	1.00 17.60
1839	N PHE	238	24.396	-9.244	3.800	1.00 17.60	1.00 17.60	2.111	1.00 17.60
1840	CA PHE	238	24.788	-8.943	2.931	1.00 17.60	1.00 17.60	1.815	1.00 17.60
1841	CB PHE	238	24.056	-8.806	1.343	1.00 17.60	1.00 17.60	2.086	1.00 17.60
1842	C PHE	238	22.885	-9.311	0.981	1.00 17.60	1.00 17.60	1.751	1.00 17.60
1843	CD1 PHE	238	24.527	-11.056	1.047	1.00 17.60	1.00 17.60	1.025	1.00 17.60
1844	CD2 PHE	238	22.172	-10.362	-0.032	1.00 17.60	1.00 17.60	1.808	1.00 17.60
1845	CD1 PHE	238	23.809	-11.866	0.206	1.00 17.60	1.00 17.60	3.069	1.00 17.60
1846	CE2 PHE	238	22.630	-11.429	-0.236	1.00 17.60	1.00 17.60	3.034	1.00 17.60
1847	CE PHE	238	24.163	-7.886	4.382	1.00 17.60	1.00 17.60	-2.427	1.00 17.60
1848	C PHE	238	23.085	-7.216	4.145	1.00 17.60	1.00 17.60	0.320	1.00 17.60
1849	N PHE	239	23.228	-7.500	5.099	1.00 17.60	1.00 17.60	0.014	1.00 17.60
1850	CA PHE	239	25.462	-6.215	5.740	1.00 17.60	1.00 17.60	-2.014	1.00 17.60
1851	CB PHE	239	24.525	-6.053	6.947	1.00 17.60	1.00 17.60	-5.582	1.00 17.60
1852	CB PHE	239	24.660	-7.010	8.125	1.00 17.60	1.00 17.60	-2.736	1.00 17.60
1853	CB PHE	239	24.064	-8.253	8.082	1.00 17.60	1.00 17.60	-2.729	1.00 17.60
1854	CD1 PHE	239	25.398	-6.622	9.225	1.00 17.60	1.00 17.60	-3.330	1.00 17.60
1855	CD2 PHE	239	24.228	-9.187	9.323	1.00 17.60	1.00 17.60	-2.230	1.00 17.60
1856	CE1 PHE	239	25.553	-7.489	10.269	1.00 17.60	1.00 17.60	-2.279	1.00 17.60
1857	CE2 PHE	239	24.972	-8.735	10.278	1.00 17.60	1.00 17.60	-0.728	1.00 17.60
1858	CE PHE	239	26.956	-6.091	6.191	1.00 17.60	1.00 17.60	-3.007	1.00 17.60
1859	C PHE	239	27.628	-7.128	6.209	1.00 17.60	1.00 17.60	-2.508	1.00 17.60
1860	O PHE	239	27.542	-4.889	6.409	1.00 17.60	1.00 17.60	-6.866	1.00 17.60
1861	N ALA	240	28.837	-4.616	7.050	1.00 17.60	1.00 17.60	-7.721	1.00 17.60
1862	CA ALA	240	29.937	-4.786	6.125	1.00 17.60	1.00 17.60	-1.520	1.00 17.60
1863	CB ALA	240	28.804	-3.127	7.354	1.00 17.60	1.00 17.60	-1.679	1.00 17.60
1864	C ALA	240	27.004	-2.518	6.978	1.00 17.60	1.00 17.60	-0.382	1.00 17.60
1865	O ALA	240	29.697	-2.372	7.992	1.00 17.60	1.00 17.60	-0.173	1.00 17.60
1866	N ASP	241	29.457	-0.922	8.027	1.00 17.60	1.00 17.60	-10.360	1.00 17.60
1868	CB ASP	241	29.690	-0.330	9.428	1.00 17.60	1.00 17.60	-11.422	1.00 17.60
1869	CD ASP	241	30.505	-1.230	10.345	1.00 17.60	1.00 17.60	-10.290	1.00 17.60
1870	OD1 ASP	241	31.687	-1.461	10.058	1.00 17.60	1.00 17.60	-9.443	1.00 17.60
1871	OD2 ASP	241	29.966	-1.732	11.340	1.00 17.60	1.00 17.60	-10.306	1.00 17.60
1872	C ASP	241	30.473	-0.415	7.021	1.00 17.60	1.00 17.60	-9.405	1.00 17.60
1873	O ASP	241	31.443	-0.535	7.347	1.00 17.60	1.00 17.60	-10.511	1.00 17.60
1874	N GLN	242	30.121	0.011	5.798	1.00 17.60	1.00 17.60	-10.265	1.00 17.60
1875	CA GLN	242	32.536	0.171	4.714	1.00 17.60	1.00 17.60	-9.080	1.00 17.60
1876	CB GLN	242	33.588	1.136	5.358	1.00 17.60	1.00 17.60	-8.994	1.00 17.60
1877	CG GLN	242	33.096	2.009	6.520	1.00 17.60	1.00 17.60	-10.140	1.00 17.60
1878	CD GLN	242	32.391	3.002	6.315	1.00 17.60	1.00 17.60	-10.775	1.00 17.60
1879	OE1 GLN	242	33.411	1.676	7.772	1.00 17.60	1.00 17.60	-11.889	1.00 17.60
1880	OE2 GLN	242	30.563	-0.337	3.542	1.00 17.60	1.00 17.60	-9.615	1.00 17.60
1881	C GLN	242	30.838	-1.547	3.494	1.00 17.60	1.00 17.60	-9.590	1.00 17.60
1882	GLN	242						-8.092	1.00 17.60
1883	GLN	242						-2.359	1.00 17.60
1884	CD PRO	243	29.877	0.216					
1885	CA PRO	243	29.893	1.599					
1886	CB PRO	243	29.018	-0.521					
1887	CG PRO	243	28.497	0.537					
1888	C PRO	243	28.534	1.735					
1889	O PRO	243	29.930	-1.541					
1890	N PRO	244	29.514	-2.678					
1891	CA ILE	244	31.305	-1.388					
1892	CB ILE	244	32.047	-2.051					
1893	CG2 ILE	244	33.420	-1.276					
1894	CG1 ILE	244	34.314	-1.396					
1895	CD1 ILE	244	34.088	-1.764					
1896	C ILE	244	33.694	-0.698					
1897	O ILE	244	32.150	-3.452					
1898	N GLN	245	32.083	-4.486					
1899	CA GLN	245	32.162	-3.488					
1900	CB GLN	245	32.245	-4.681					
1901	CG GLN	245	32.359	-4.238					
1902	CD GLN	245	33.738	-3.631					
1903	OE1 GLN	245	33.687	-2.625					
1904	OE2 GLN	245	34.599	-3.866					
1905	C GLN	245	31.044	-5.612					
1906	O GLN	245	31.844	-6.823					
1907	N ILE	246	29.904	-6.978					
1908	CA ILE	246	28.612	-5.532					
1909	CB ILE	246	27.524	-4.441					
1910	CG2 ILE	246	26.211	-5.178					
1911	CG1 ILE	246	27.607	-3.554					
1912	CD1 ILE	246	26.584	-2.427					
1913	C ILE	246	28.732	-6.123					
1914	O ILE	246	28.544	-7.318					
1915	CA TYR	247	29.198	-5.272					
1916	CB TYR	247	29.202	-5.582					
1917	CG TYR	247	29.889	-4.467					
1918	CD TYR	247	29.145	-3.139					
1919	CE1 TYR	247	29.809	-2.074					
1920	CE2 TYR	247	29.199	-0.617					
1921	CD2 TYR	247	27.882	-2.998					
1922	CE3 TYR	247	27.335	-1.785					
1923	CE TYR	247	27.916	-0.728					
1924	ON TYR	247	27.298	0.476					
1925	C TYR	247	29.866	-6.866					
1926	O TYR	247	29.321	-7.721					
1927	N GLU	248	30.959	-7.039					
1928	CA GLU	248	31.782	-8.221					
1929	CB GLU	248	32.844	-8.026					
1930	CG GLU	248	34.807	-9.014					
1931	CD GLU	248	33.591	-10.360					
1932	OE1 GLU	248	33.919	-11.422					
1933	OE2 GLU	248	32.922	-10.290					
1934	C GLU	248	30.928	-9.443					
1935	O GLU	248	30.890	-10.306					
1936	N LYS	249	30.157	-9.405					
1937	CA LYS	249	29.321	-10.511					
1938	CB LYS	249	28.797	-10.265					
1939	CG LYS	249	29.881	-9.080					
1940	CD LYS	249	28.889	-8.994					
1941	CE LYS	249	28.134	-10.140					
1942	OE LYS	249	27.635	-11.889					
1943	N ILE	250	27.640	-9.615					
1944	O ILE	250	26.441	-9.590					
1945	CA ILE	250	26.178	-8.092					
1946	CB ILE	250							

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1918	CG2	ILE	250	25.909	-7.738	-3.840	1.00	17.60	ATOK	2013	C	PRO	258	18.080	-19.017	11.184	1.00	17.60
1919	CG1	ILE	250	24.998	-7.718	-1.462	1.00	17.60	ATOK	2014	O	PRO	258	17.476	-20.464	10.316	1.00	17.60
1920	CD1	ILE	250	25.529	-7.871	-1.968	1.00	17.60	ATOK	2015	N	SER	259	18.544	-20.223	12.436	1.00	17.60
1921	C	ILE	250	26.816	-10.405	-3.321	1.00	17.60	ATOK	2016	CA	SER	259	17.652	-21.470	12.911	1.00	17.60
1922	O	ILE	250	26.135	-11.384	-3.663	1.00	17.60	ATOK	2017	CB	SER	259	18.197	-21.803	14.309	1.00	17.60
1923	N	VAL	251	27.994	-10.077	-3.835	1.00	17.60	ATOK	2018	O2	SER	259	16.408	-20.589	15.064	1.00	17.60
1924	CA	VAL	251	28.492	-10.717	-5.003	1.00	17.60	ATOK	2019	C	SER	259	16.129	-21.414	12.985	1.00	17.60
1925	CB	VAL	251	30.539	-9.017	-5.416	1.00	17.60	ATOK	2020	O	SER	259	15.476	-22.451	12.091	1.00	17.60
1926	CG1	VAL	251	29.038	-8.689	-6.423	1.00	17.60	ATOK	2021	N	HIS	260	15.524	-20.217	13.235	1.00	17.60
1927	CG2	VAL	251	28.033	-12.173	-4.762	1.00	17.60	ATOK	2022	CA	HIS	260	14.070	-20.105	13.298	1.00	17.60
1928	C	VAL	251	28.539	-12.956	-5.653	1.00	17.60	ATOK	2023	CB	HIS	260	13.732	-18.943	14.251	1.00	17.60
1929	N	SER	252	29.389	-12.661	-3.638	1.00	17.60	ATOK	2024	CC2	HIS	260	14.351	-17.562	13.011	1.00	17.60
1930	CA	SER	252	29.765	-14.078	-3.692	1.00	17.60	ATOK	2025	ND1	HIS	260	15.467	-17.094	13.752	1.00	17.60
1931	CB	SER	252	30.662	-14.234	-2.308	1.00	17.60	ATOK	2026	CE1	HIS	260	13.362	-16.566	13.455	1.00	17.60
1932	CG	SER	252	30.253	-13.510	-1.147	1.00	17.60	ATOK	2027	CE2	HIS	260	14.309	-15.511	13.189	1.00	17.60
1933	C	SER	252	28.571	-15.081	-3.317	1.00	17.60	ATOK	2028	N2	HIS	260	15.371	-15.014	13.373	1.00	17.60
1934	N	GLY	253	27.639	-14.412	-2.469	1.00	17.60	ATOK	2029	C	HIS	260	12.103	-19.314	12.003	1.00	17.60
1935	CA	GLY	253	26.565	-16.562	-1.094	1.00	17.60	ATOK	2030	O	HIS	260	13.271	-19.314	12.003	1.00	17.60
1936	CG	GLY	253	26.746	-17.698	-2.502	1.00	17.60	ATOK	2031	N	PHE	261	13.836	-20.427	10.901	1.00	17.60
1937	O	GLY	253	26.506	-16.591	-0.599	1.00	17.60	ATOK	2032	CA	PHE	261	13.273	-20.300	9.641	1.00	17.60
1938	N	LYE	254	26.495	-17.657	-0.295	1.00	17.60	ATOK	2033	CB	PHE	261	14.396	-19.027	0.707	1.00	17.60
1939	CA	LYE	254	27.782	-17.887	1.041	1.00	17.60	ATOK	2034	CG	PHE	261	14.538	-18.327	0.566	1.00	17.60
1940	CG	LYE	254	28.099	-17.988	0.081	1.00	17.60	ATOK	2035	CD1	PHE	261	13.971	-17.432	9.495	1.00	17.60
1941	CD	LYE	254	30.176	-18.339	0.800	1.00	17.60	ATOK	2036	CD2	PHE	261	15.254	-17.812	7.498	1.00	17.60
1942	CE	LYE	254	31.293	-18.136	-0.230	1.00	17.60	ATOK	2037	CB1	PHE	261	14.127	-16.062	9.347	1.00	17.60
1943	N2	LYE	254	31.331	-18.921	-1.459	1.00	17.60	ATOK	2038	CR2	PHE	261	15.403	-16.446	7.349	1.00	17.60
1944	C	LYE	254	23.433	-16.953	1.142	1.00	17.60	ATOK	2039	CA	PHE	261	14.812	-15.578	0.275	1.00	17.60
1945	N	VAL	255	25.746	-15.818	1.500	1.00	17.60	ATOK	2040	C	PHE	261	12.285	-21.005	9.003	1.00	17.60
1946	CA	VAL	255	24.201	-17.354	1.402	1.00	17.60	ATOK	2041	O	PHE	261	11.490	-20.459	0.362	1.00	17.60
1947	CB	VAL	255	23.342	-16.510	2.349	1.00	17.60	ATOK	2042	N	SER	262	12.465	-22.302	9.171	1.00	17.60
1948	CG1	VAL	255	21.813	-16.318	1.096	1.00	17.60	ATOK	2043	CA	SER	262	11.352	-23.169	8.427	1.00	17.60
1949	CG2	VAL	255	21.370	-15.030	2.420	1.00	17.60	ATOK	2044	CB	SER	262	9.090	-22.682	8.169	1.00	17.60
1950	C	VAL	255	23.603	-16.482	0.380	1.00	17.60	ATOK	2045	CG	SER	262	9.666	-23.938	6.986	1.00	17.60
1951	N	ARG	256	23.912	-17.568	6.063	1.00	17.60	ATOK	2046	C	SER	262	11.955	-23.265	7.043	1.00	17.60
1952	CA	ARG	256	24.105	-17.351	6.890	1.00	17.60	ATOK	2047	O	SER	262	12.594	-22.331	6.503	1.00	17.60
1953	CB	ARG	256	25.219	-18.082	6.447	1.00	17.60	ATOK	2048	N	SER	263	11.727	-23.066	6.163	1.00	17.60
1954	CD	ARG	256	24.903	-19.535	6.867	1.00	17.60	ATOK	2049	CA	SER	263	11.723	-24.469	6.494	1.00	17.60
1955	CE	ARG	256	25.672	-20.115	8.025	1.00	17.60	ATOK	2050	CG	SER	263	12.283	-24.832	5.191	1.00	17.60
1956	CH1	ARG	256	27.014	-18.783	8.015	1.00	17.60	ATOK	2051	O2	SER	263	11.097	-26.302	4.915	1.00	17.60
1957	CH2	ARG	256	27.060	-20.395	9.808	1.00	17.60	ATOK	2052	C	SER	263	12.062	-27.159	6.098	1.00	17.60
1958	C	ARG	256	21.660	-17.605	9.910	1.00	17.60	ATOK	2053	O	SER	263	11.727	-23.066	6.163	1.00	17.60
1959	N	PHE	257	21.452	-16.912	7.898	1.00	17.60	ATOK	2054	N	ASP	264	10.411	-23.700	3.535	1.00	17.60
1960	CA	PHE	257	20.816	-18.518	6.545	1.00	17.60	ATOK	2055	CA	ASP	264	8.211	-22.772	3.851	1.00	17.60
1961	CB	PHE	257	19.555	-18.599	7.179	1.00	17.60	ATOK	2056	CB	ASP	264	7.203	-23.880	3.522	1.00	17.60
1962	CD	PHE	257	18.723	-19.553	6.453	1.00	17.60	ATOK	2057	CC	ASP	264	9.639	-22.927	3.318	1.00	17.60
1963	CE	PHE	257	18.611	-19.193	4.082	1.00	17.60	ATOK	2058	CO1	ASP	264	7.482	-24.901	2.857	1.00	17.60
1964	CH1	PHE	257	18.001	-20.344	4.082	1.00	17.60	ATOK	2059	CO2	ASP	264	10.316	-21.573	3.004	1.00	17.60
1965	CH2	PHE	257	18.223	-17.339	4.646	1.00	17.60	ATOK	2060	C	ASP	264	10.463	-21.298	3.017	1.00	17.60
1966	C	PHE	257	18.754	-19.817	3.322	1.00	17.60	ATOK	2061	O	ASP	264	10.477	-20.800	4.065	1.00	17.60
1967	CA	PHE	257	18.097	-17.614	3.322	1.00	17.60	ATOK	2062	N	LEU	265	11.003	-19.477	3.860	1.00	17.60
1968	CB	PHE	257	18.362	-18.596	2.400	1.00	17.60	ATOK	2063	CA	LEU	265	11.071	-18.608	5.039	1.00	17.60
1969	C	PHE	257	19.566	-19.006	8.637	1.00	17.60	ATOK	2064	CB	LEU	265	11.562	-17.191	4.914	1.00	17.60
1970	O	PHE	257	19.995	-20.125	9.991	1.00	17.60	ATOK	2065	CG	LEU	265	10.632	-16.531	3.829	1.00	17.60
1971	N	PRO	258	19.001	-18.123	9.511	1.00	17.60	ATOK	2066	CD2	LEU	265	11.649	-16.403	6.091	1.00	17.60
1972	CA	PRO	258	18.688	-16.516	9.235	1.00	17.60	ATOK	2067	CD3	LEU	265	12.404	-18.432	3.374	1.00	17.60
1973	CB	PRO	258	18.667	-19.445	10.912	1.00	17.60	ATOK	2068	C	LEU	265	12.688	-19.009	2.357	1.00	17.60
1974	CD	PRO	258	17.717	-17.370	11.379	1.00	17.60	ATOK	2069	O	LEU	265	13.225	-20.508	3.944	1.00	17.60
1975	CG	PRO	258	17.509	-16.312	10.200	1.00	17.60	ATOK	2070	N	LYS	266	14.591	-20.485	3.489	1.00	17.60
1976	N	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2071	CA	LYS	266	15.242	-21.871	4.108	1.00	17.60
1977	CA	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2072	CB	LYS	266	15.626	-21.725	5.535	1.00	17.60
1978	CB	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2073	CG	LYS	266	16.311	-23.032	5.808	1.00	17.60
1979	CD	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2074	CD	LYS	266	16.304	-23.317	7.305	1.00	17.60
1980	CE	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2075	CE	LYS	266	16.969	-23.329	7.936	1.00	17.60
1981	CH1	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2076	N2	LYS	266	14.631	-20.936	2.002	1.00	17.60
1982	CH2	ARG	259	17.509	-16.312	10.200	1.00	17.60	ATOK	2077	C	LYS	266	14.631	-20.936	2.002	1.00	17.60

2078	D	LYS	266	15.461	-20.442	1.261	1.00	17.60	2078	D	LYS	266	20.062	-16.311	-9.294	1.00	17.60
2079	N	ASP	267	13.601	-21.651	1.618	1.00	17.60	2079	N	ASP	267	21.058	-19.500	-7.481	1.00	17.60
2080	CA	ASP	267	13.439	-22.100	0.294	1.00	17.60	2080	CA	ASP	267	20.747	-15.696	-7.421	1.00	17.60
2081	CB	ASP	267	12.412	-23.144	0.246	1.00	17.60	2081	CB	ASP	267	20.352	-14.512	-7.988	1.00	17.60
2082	CG	ASP	267	12.441	-23.703	-1.355	1.00	17.60	2082	CG	ASP	267	21.973	-15.060	-7.758	1.00	17.60
2083	DI	ASP	267	13.479	-24.764	-1.538	1.00	17.60	2083	DI	ASP	267	21.941	-15.123	-8.329	1.00	17.60
2084	OD2	ASP	267	11.453	-23.513	-1.869	1.00	17.60	2084	OD2	ASP	267	24.315	-15.698	-8.033	1.00	17.60
2085	C	ASP	267	13.018	-20.983	-0.607	1.00	17.60	2085	C	ASP	267	25.404	-14.781	-8.570	1.00	17.60
2086	N	LEU	268	13.476	-20.936	-1.756	1.00	17.60	2086	N	LEU	268	24.499	-15.803	-6.540	1.00	17.60
2087	N	LEU	268	12.135	-20.084	-0.212	1.00	17.60	2087	N	LEU	268	22.764	-14.832	-9.818	1.00	17.60
2088	CA	LEU	268	11.825	-19.036	-1.164	1.00	17.60	2088	CA	LEU	268	23.037	-13.758	-10.430	1.00	17.60
2089	CB	LEU	268	10.565	-18.374	-0.662	1.00	17.60	2089	CB	LEU	268	22.308	-15.958	-10.354	1.00	17.60
2090	CG	LEU	268	9.979	-17.186	-1.342	1.00	17.60	2090	CG	LEU	268	22.149	-16.146	-11.764	1.00	17.60
2091	DI	LEU	268	9.629	-17.167	-2.039	1.00	17.60	2091	DI	LEU	268	22.474	-17.635	-13.990	1.00	17.60
2092	OD2	LEU	268	8.717	-16.925	-0.465	1.00	17.60	2092	OD2	LEU	268	22.696	-18.178	-13.434	1.00	17.60
2093	C	LEU	268	13.032	-18.082	-1.273	1.00	17.60	2093	C	LEU	268	22.619	-19.160	-13.651	1.00	17.60
2094	D	LEU	268	13.415	-17.603	-2.339	1.00	17.60	2094	D	LEU	268	22.913	-17.260	-14.316	1.00	17.60
2095	N	LEU	269	13.740	-17.927	-0.174	1.00	17.60	2095	N	LEU	269	20.771	-15.751	-12.262	1.00	17.60
2096	CA	LEU	269	14.897	-17.067	-0.347	1.00	17.60	2096	CA	LEU	269	19.813	-16.508	-12.101	1.00	17.60
2097	CB	LEU	269	15.249	-16.967	-1.312	1.00	17.60	2097	CB	LEU	269	20.641	-14.624	-12.931	1.00	17.60
2098	CG	LEU	269	14.508	-15.887	2.089	1.00	17.60	2098	CG	LEU	269	19.377	-14.232	-13.555	1.00	17.60
2099	DI	LEU	269	13.094	-15.659	1.679	1.00	17.60	2099	DI	LEU	269	19.595	-13.082	-14.556	1.00	17.60
2100	OD2	LEU	269	14.562	-16.213	3.560	1.00	17.60	2100	OD2	LEU	269	19.963	-11.735	-13.917	1.00	17.60
2101	C	LEU	269	16.078	-17.569	-0.966	1.00	17.60	2101	C	LEU	269	19.660	-10.451	-14.766	1.00	17.60
2102	N	ARG	270	16.692	-16.845	-3.750	1.00	17.60	2102	N	ARG	270	18.057	-11.628	-12.712	1.00	17.60
2103	N	ARG	270	16.336	-18.847	-0.079	1.00	17.60	2103	N	ARG	270	17.359	-15.465	-14.130	1.00	17.60
2104	CA	ARG	270	17.451	-19.404	-1.586	1.00	17.60	2104	CA	ARG	270	19.376	-16.134	-14.987	1.00	17.60
2105	CB	ARG	270	17.755	-20.782	-3.029	1.00	17.60	2105	CB	ARG	270	18.372	-17.243	-15.617	1.00	17.60
2106	CG	ARG	270	16.508	-21.698	-3.262	1.00	17.60	2106	CG	ARG	270	19.643	-17.883	-16.527	1.00	17.60
2107	DI	ARG	270	16.759	-23.140	-0.094	1.00	17.60	2107	DI	ARG	270	20.167	-16.780	-17.326	1.00	17.60
2108	NE	ARG	270	16.966	-23.232	0.349	1.00	17.60	2108	NE	ARG	270	19.095	-19.080	-17.353	1.00	17.60
2109	C2	ARG	270	17.099	-24.451	1.196	1.00	17.60	2109	C2	ARG	270	17.859	-18.249	-14.695	1.00	17.60
2110	NH1	ARG	270	17.047	-25.616	0.538	1.00	17.60	2110	NH1	ARG	270	16.909	-18.985	-15.117	1.00	17.60
2111	NH2	ARG	270	17.287	-26.439	2.336	1.00	17.60	2111	NH2	ARG	270	18.762	-18.427	-13.447	1.00	17.60
2112	C	ARG	270	17.062	-19.465	-3.058	1.00	17.60	2112	C	ARG	270	17.528	-19.292	-12.534	1.00	17.60
2113	O	ARG	270	17.967	-19.643	-3.990	1.00	17.60	2113	O	ARG	270	18.534	-20.258	-12.048	1.00	17.60
2114	N	ASN	271	15.795	-19.377	-3.415	1.00	17.60	2114	N	ASN	271	19.377	-20.905	-13.147	1.00	17.60
2115	CA	ASN	271	15.368	-19.268	-4.813	1.00	17.60	2115	CA	ASN	271	20.538	-21.679	-12.474	1.00	17.60
2116	CB	ASN	271	13.997	-19.901	-5.459	1.00	17.60	2116	CB	ASN	271	20.130	-21.739	-13.426	1.00	17.60
2117	CG	ASN	271	13.780	-21.417	-5.278	1.00	17.60	2117	CG	ASN	271	21.325	-22.429	-10.933	1.00	17.60
2118	OD1	ASN	271	12.743	-21.913	-4.807	1.00	17.60	2118	OD1	ASN	271	16.790	-18.583	-11.351	1.00	17.60
2119	ND2	ASN	271	14.614	-22.258	-5.924	1.00	17.60	2119	ND2	ASN	271	16.134	-19.232	-10.330	1.00	17.60
2120	C	ASN	271	15.173	-17.803	-5.208	1.00	17.60	2120	C	ASN	271	16.230	-16.476	-10.266	1.00	17.60
2121	O	ASN	271	14.920	-17.565	-6.309	1.00	17.60	2121	O	ASN	271	16.431	-14.984	-10.371	1.00	17.60
2122	N	LEU	272	15.026	-15.417	-4.878	1.00	17.60	2122	N	LEU	272	16.030	-14.078	-9.408	1.00	17.60
2123	CA	LEU	272	15.160	-16.770	-4.362	1.00	17.60	2123	CA	LEU	272	16.787	-12.737	-9.345	1.00	17.60
2124	CB	LEU	272	14.030	-14.590	-4.078	1.00	17.60	2124	CB	LEU	272	18.231	-12.942	-9.196	1.00	17.60
2125	CG	LEU	272	12.524	-14.755	-4.354	1.00	17.60	2125	CG	LEU	272	19.132	-11.946	-9.404	1.00	17.60
2126	OD1	LEU	272	11.600	-16.164	-3.288	1.00	17.60	2126	OD1	LEU	272	18.697	-10.723	-9.742	1.00	17.60
2127	OD2	LEU	272	12.314	-16.077	-5.697	1.00	17.60	2127	OD2	LEU	272	20.431	-12.382	-9.353	1.00	17.60
2128	C	LEU	272	16.359	-14.767	-4.814	1.00	17.60	2128	C	LEU	272	14.719	-16.649	-10.208	1.00	17.60
2129	O	LEU	272	16.756	-14.083	-5.736	1.00	17.60	2129	O	LEU	272	14.054	-16.601	-11.265	1.00	17.60
2130	N	LEU	273	17.158	-14.997	-3.789	1.00	17.60	2130	N	LEU	273	14.094	-16.867	-9.010	1.00	17.60
2131	CA	LEU	273	18.478	-14.433	-3.662	1.00	17.60	2131	CA	LEU	273	12.646	-16.973	-9.000	1.00	17.60
2132	CB	LEU	273	18.712	-14.301	-2.191	1.00	17.60	2132	CB	LEU	273	12.104	-17.399	-7.450	1.00	17.60
2133	CG	LEU	273	18.081	-13.074	-4.609	1.00	17.60	2133	CG	LEU	273	13.552	-18.593	-7.423	1.00	17.60
2134	OD1	LEU	273	17.460	-13.322	-0.284	1.00	17.60	2134	OD1	LEU	273	12.056	-19.689	-8.102	1.00	17.60
2135	OD2	LEU	273	19.166	-12.083	-3.404	1.00	17.60	2135	OD2	LEU	273	10.514	-18.743	-6.518	1.00	17.60
2136	C	LEU	273	19.486	-15.337	-4.363	1.00	17.60	2136	C	LEU	273	11.024	-18.743	-6.518	1.00	17.60
2137	O	LEU	273	20.508	-15.007	-2.843	1.00	17.60	2137	O	LEU	273	11.507	-18.926	-7.866	1.00	17.60
2138	N	GLN	274	19.128	-15.468	-5.652	1.00	17.60	2138	N	GLN	274	9.965	-19.972	-6.383	1.00	17.60
2139	CA	GLN	274	19.727	-16.398	-6.619	1.00	17.60	2139	CA	GLN	274	10.461	-21.054	-6.951	1.00	17.60
2140	CB	GLN	274	18.642	-16.958	-7.577	1.00	17.60	2140	CB	GLN	274	12.114	-15.639	-9.416	1.00	17.60
2141	CG	GLN	274	18.856	-18.439	-8.016	1.00	17.60	2141	CG	GLN	274	12.463	-14.621	-9.038	1.00	17.60
2142	CD	GLN	274	20.301	-18.797	-8.317	1.00	17.60	2142	CD	GLN	274					

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2208	N	GLY	282	31.101	-15.177	-10.740	1.00	17.60	6.474	-16.312	-6.425	1.00	17.60
2209	CA	GLY	282	10.603	-14.791	-10.639	1.00	17.60	6.563	-14.970	-6.302	1.00	17.60
2210	C	GLY	282	11.221	-13.972	-11.972	1.00	17.60	6.664	-16.035	-4.829	1.00	17.60
2211	N	GLY	282	10.756	-13.048	-12.640	1.00	17.60	6.052	-14.467	-6.604	1.00	17.60
2212	N	ASN	283	12.748	-14.665	-12.445	1.00	17.60	8.564	-14.549	-8.022	1.00	17.60
2213	CA	ASN	283	12.743	-14.346	-13.767	1.00	17.60	5.173	-16.989	-5.783	1.00	17.60
2214	CR	ASN	283	14.253	-14.225	-13.769	1.00	17.60	5.173	-17.690	-4.781	1.00	17.60
2215	CG	ASN	283	14.726	-13.157	-16.756	1.00	17.60	4.067	-16.535	-5.911	1.00	17.60
2216	OD1	ASN	283	15.915	-12.830	-14.738	1.00	17.60	2.724	-16.893	-5.911	1.00	17.60
2217	HD2	ASN	283	13.911	-12.500	-15.629	1.00	17.60	1.594	-16.437	-5.896	1.00	17.60
2218	C	ASN	283	12.301	-15.324	-14.908	1.00	17.60	1.458	-14.895	-7.147	1.00	17.60
2219	O	ASN	283	12.462	-15.015	-16.088	1.00	17.60	0.312	-13.032	-8.107	1.00	17.60
2220	N	LEU	284	11.932	-16.517	-14.517	1.00	17.60	0.355	-14.500	-8.107	1.00	17.60
2221	CA	LEU	284	11.718	-17.686	-15.581	1.00	17.60	-0.907	-12.702	-8.233	1.00	17.60
2222	CB	LEU	284	11.626	-18.940	-14.491	1.00	17.60	2.428	-18.376	-5.617	1.00	17.60
2223	CG	LEU	284	12.476	-18.906	-13.225	1.00	17.60	1.648	-18.632	-4.701	1.00	17.60
2224	CD1	LEU	284	11.028	-19.039	-12.251	1.00	17.60	3.021	-19.198	-6.228	1.00	17.60
2225	CD2	LEU	284	13.929	-19.198	-13.510	1.00	17.60	2.531	-20.742	-5.988	1.00	17.60
2226	C	LEU	284	10.419	-17.549	-16.226	1.00	17.60	2.310	-21.531	-7.291	1.00	17.60
2227	O	LEU	284	9.615	-16.696	-15.871	1.00	17.60	1.479	-20.958	-8.248	1.00	17.60
2228	N	LVS	285	10.220	-18.339	-17.305	1.00	17.60	0.486	-20.383	-8.248	1.00	17.60
2229	CA	LVS	285	8.926	-18.322	-18.055	1.00	17.60	1.645	-20.914	-9.574	1.00	17.60
2230	CB	LVS	285	6.971	-19.381	-19.222	1.00	17.60	3.425	-21.911	-4.998	1.00	17.60
2231	CG	LVS	285	10.315	-19.010	-20.105	1.00	17.60	3.462	-22.618	-4.822	1.00	17.60
2232	CD	LVS	285	10.130	-19.092	-21.630	1.00	17.60	4.208	-20.597	-6.300	1.00	17.60
2233	CE	LVS	285	11.372	-18.332	-22.220	1.00	17.60	5.265	-21.176	-3.053	1.00	17.60
2234	ME	LVS	285	11.437	-18.246	-23.694	1.00	17.60	6.272	-20.755	-3.053	1.00	17.60
2235	C	LVS	285	7.899	-18.680	-16.999	1.00	17.60	7.522	-20.993	-2.614	1.00	17.60
2236	O	LVS	285	8.161	-19.657	-16.328	1.00	17.60	8.579	-21.241	-3.477	1.00	17.60
2237	K	ASP	286	5.878	-17.868	-15.610	1.00	17.60	7.909	-21.224	-1.380	1.00	17.60
2238	CA	ASP	286	6.043	-19.163	-14.595	1.00	17.60	9.162	-21.724	-1.491	1.00	17.60
2239	CB	ASP	286	5.773	-20.638	-15.087	1.00	17.60	9.564	-21.675	-2.745	1.00	17.60
2240	CC	ASP	286	4.767	-20.877	-15.812	1.00	17.60	4.559	-21.590	-2.337	1.00	17.60
2241	OD1	ASP	286	6.515	-21.549	-14.746	1.00	17.60	3.915	-20.676	-1.707	1.00	17.60
2242	OD2	ASP	286	6.234	-16.620	-14.950	1.00	17.60	5.100	-22.737	-2.049	1.00	17.60
2243	C	ASP	286	5.837	-15.541	-15.415	1.00	17.60	4.797	-22.486	-0.890	1.00	17.60
2244	N	GLY	287	7.004	-16.550	-13.886	1.00	17.60	5.771	-24.624	-0.775	1.00	17.60
2245	CA	GLY	287	7.401	-15.275	-13.364	1.00	17.60	6.025	-25.636	-1.991	1.00	17.60
2246	C	GLY	287	6.755	-15.095	-12.039	1.00	17.60	6.093	-25.162	-3.393	1.00	17.60
2247	O	GLY	287	7.018	-15.879	-11.132	1.00	17.60	8.166	-24.629	-3.400	1.00	17.60
2248	N	VAL	288	5.817	-14.136	-11.946	1.00	17.60	8.562	-24.062	-4.677	1.00	17.60
2249	CA	VAL	288	5.110	-13.879	-10.718	1.00	17.60	4.682	-22.556	0.338	1.00	17.60
2250	CB	VAL	288	3.978	-12.923	-10.820	1.00	17.60	3.745	-22.673	1.107	1.00	17.60
2251	CG	VAL	288	4.073	-12.103	-9.586	1.00	17.60	5.330	-21.520	0.456	1.00	17.60
2252	CD	VAL	288	3.966	-12.148	-12.093	1.00	17.60	5.439	-20.578	1.591	1.00	17.60
2253	C	VAL	288	4.917	-15.140	-10.221	1.00	17.60	6.964	-18.046	2.840	1.00	17.60
2254	N	ASN	289	4.450	-15.419	-9.074	1.00	17.60	6.771	-17.488	3.002	1.00	17.60
2255	CA	ASN	289	3.825	-15.994	-11.059	1.00	17.60	7.198	-17.290	4.317	1.00	17.60
2256	CB	ASN	289	3.190	-17.197	-10.541	1.00	17.60	6.314	-16.431	2.248	1.00	17.60
2257	CG	ASN	289	3.306	-17.880	-11.598	1.00	17.60	7.484	-19.413	3.984	1.00	17.60
2258	CD	ASN	289	1.085	-16.878	-11.756	1.00	17.60	7.614	-18.450	4.866	1.00	17.60
2259	OD1	ASN	289	1.126	-16.013	-12.524	1.00	17.60	7.187	-15.043	4.917	1.00	17.60
2260	OD2	ASN	289	0.000	-17.132	-10.992	1.00	17.60	6.307	-15.193	2.850	1.00	17.60
2261	N	ASP	290	4.174	-18.204	-10.012	1.00	17.60	6.737	-14.925	4.163	1.00	17.60
2262	C	ASP	290	3.734	-19.257	-9.588	1.00	17.60	4.165	-19.701	1.506	1.00	17.60
2263	N	ASP	290	5.486	-17.972	-9.994	1.00	17.60	3.579	-19.452	2.552	1.00	17.60
2264	CA	ASP	290	6.383	-18.905	-9.344	1.00	17.60	3.657	-19.184	0.394	1.00	17.60
2265	CB	ASP	290	7.806	-18.805	-9.886	1.00	17.60	2.687	-18.313	0.457	1.00	17.60
2266	CG	ASP	290	8.101	-19.139	-11.259	1.00	17.60	2.793	-17.272	-0.617	1.00	17.60
2267	CD	ASP	290	8.797	-18.348	-12.028	1.00	17.60	3.354	-16.198	-0.817	1.00	17.60
2268	OD1	ASP	290	7.687	-20.204	-11.835	1.00	17.60	3.212	-14.984	-0.219	1.00	17.60
2269	OD2	ASP	290	6.324	-18.357	-7.955	1.00	17.60	4.647	-16.437	-1.616	1.00	17.60
2270	C	P	290	6.118	-19.151	-7.038	1.00	17.60	4.153	-13.908	-0.108	1.00	17.60
2271	O	P	290	6.402	-17.022	-7.767	1.00	17.60	5.385	-15.447	-3.798	1.00	17.60
2272	N	P	291										

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ATOM	2403	N	TTR	306	-5.549	-8.194	-2.283	1.00	17.60
ATOM	2404	CA	TTR	306	-5.746	-7.125	-3.203	1.00	17.60
ATOM	2405	CB	TTR	306	-5.939	-7.133	-4.565	1.00	17.60
ATOM	2406	CG	TTR	306	-6.361	-6.684	-5.374	1.00	17.60
ATOM	2407	CD1	TTR	306	-7.659	-6.581	-6.040	1.00	17.60
ATOM	2408	CP1	TTR	306	-8.075	-5.493	-6.812	1.00	17.60
ATOM	2409	CD2	TTR	306	-5.437	-5.726	-5.897	1.00	17.60
ATOM	2410	CE2	TTR	306	-5.829	-4.660	-6.662	1.00	17.60
ATOM	2411	CE	TTR	306	-7.134	-4.532	-7.110	1.00	17.60
ATOM	2412	CH	TTR	306	-7.490	-3.393	-7.821	1.00	17.60
ATOM	2413	C	TTR	306	-6.889	-6.222	-2.785	1.00	17.60
ATOM	2414	O	TTR	306	-7.986	-6.799	-2.301	1.00	17.60
ATOM	2415	N	GLN	307	-9.195	-6.015	-1.971	1.00	17.60
ATOM	2416	CA	GLN	307	-10.433	-6.917	-1.855	1.00	17.60
ATOM	2417	CB	GLN	307	-11.113	-7.406	-3.146	1.00	17.60
ATOM	2418	CG	GLN	307	-11.418	-8.016	-3.125	1.00	17.60
ATOM	2419	CD1	GLN	307	-12.418	-9.302	-2.132	1.00	17.60
ATOM	2420	CE1	GLN	307	-11.727	-9.563	-4.248	1.00	17.60
ATOM	2421	CE2	GLN	307	-9.016	-5.261	-0.656	1.00	17.60
ATOM	2422	C	GLN	307	-10.044	-4.709	-0.122	1.00	17.60
ATOM	2423	O	GLN	307	-7.016	-5.379	-0.146	1.00	17.60
ATOM	2424	N	ARG	308	-7.325	-4.763	1.124	1.00	17.60
ATOM	2425	CA	ARG	308	-7.594	-2.275	1.194	1.00	17.60
ATOM	2426	CB	ARG	308	-6.828	-2.628	0.084	1.00	17.60
ATOM	2427	CG	ARG	308	-7.136	-1.203	0.288	1.00	17.60
ATOM	2428	CD	ARG	308	-6.016	-0.516	0.878	1.00	17.60
ATOM	2429	CE	ARG	308	-5.972	-0.300	2.183	1.00	17.60
ATOM	2430	C1	ARG	308	-6.352	-0.752	3.002	1.00	17.60
ATOM	2431	NH1	ARG	308	-5.004	0.524	2.613	1.00	17.60
ATOM	2432	NH2	ARG	308	-8.278	-5.502	2.045	1.00	17.60
ATOM	2433	O	ARG	308	-8.570	-6.670	1.800	1.00	17.60
ATOM	2434	N	LYS	309	-8.657	-4.856	3.121	1.00	17.60
ATOM	2435	CA	LYS	309	-9.609	-5.356	4.021	1.00	17.60
ATOM	2436	CB	LYS	309	-11.135	-5.108	3.338	1.00	17.60
ATOM	2437	CG	LYS	309	-11.535	-3.759	2.569	1.00	17.60
ATOM	2438	CD	LYS	309	-12.024	-3.488	3.363	1.00	17.60
ATOM	2439	CE	LYS	309	-11.843	-1.024	2.763	1.00	17.60
ATOM	2440	CE	LYS	309	-12.705	-0.667	1.625	1.00	17.60
ATOM	2441	N1	LYS	309	-9.401	-6.846	6.410	1.00	17.60
ATOM	2442	C	LYS	309	-8.782	-7.197	5.303	1.00	17.60
ATOM	2443	O	LYS	309	-8.956	-7.216	3.522	1.00	17.60
ATOM	2444	N	VAL	310	-10.010	-9.134	3.772	1.00	17.60
ATOM	2445	CA	VAL	310	-10.474	-9.846	2.386	1.00	17.60
ATOM	2446	CB	VAL	310	-11.106	-11.205	2.768	1.00	17.60
ATOM	2447	CG1	VAL	310	-11.537	-9.078	1.524	1.00	17.60
ATOM	2448	CG2	VAL	310	-8.859	-9.765	4.443	1.00	17.60
ATOM	2449	C	VAL	310	-9.125	-10.420	5.459	1.00	17.60
ATOM	2450	O	VAL	310	-7.567	-9.845	6.075	1.00	17.60
ATOM	2451	N	GLU	311	-6.596	-10.263	4.971	1.00	17.60
ATOM	2452	CA	GLU	311	-5.242	-10.629	4.315	1.00	17.60
ATOM	2453	CB	GLU	311	-5.257	-12.036	3.644	1.00	17.60
ATOM	2454	CG	GLU	311	-4.553	-13.265	4.251	1.00	17.60
ATOM	2455	CD	GLU	311	-4.936	-11.396	3.822	1.00	17.60
ATOM	2456	CE1	GLU	311	-3.611	-13.210	5.086	1.00	17.60
ATOM	2457	CE2	GLU	311	-6.424	-9.160	5.979	1.00	17.60
ATOM	2458	C	GLU	311	-5.955	-8.033	5.763	1.00	17.60
ATOM	2459	O	GLU	311	-7.056	-9.479	7.031	1.00	17.60
ATOM	2460	N	ALA	312	-6.953	-8.486	6.165	1.00	17.60
ATOM	2461	CA	ALA	312	-7.929	-8.876	9.278	1.00	17.60
ATOM	2462	CB	ALA	312	-5.493	-8.486	6.655	1.00	17.60
ATOM	2463	C	ALA	312	-4.015	-9.515	8.702	1.00	17.60
ATOM	2464	O	ALA	312	-5.008	-7.248	8.779	1.00	17.60
ATOM	2465	N	PRO	313	-5.764	-6.030	8.556	1.00	17.60
ATOM	2466	CA	PRO	313	-3.676	-6.869	9.148	1.00	17.60

2338	CE	PHE	297	5.744	-14.227	-1.138	1.00	17.60
2339	C	PHE	297	1.538	-19.393	0.061	1.00	17.60
2340	C	PHE	297	1.593	-19.926	-1.092	1.00	17.60
2341	N	ALA	298	0.713	-19.049	0.881	1.00	17.60
2342	CA	ALA	298	-0.775	-20.841	0.574	1.00	17.60
2343	CB	ALA	298	0.378	-22.228	0.736	1.00	17.60
2344	C	ALA	298	-1.627	-20.745	1.320	1.00	17.60
2345	ALA	298	-2.685	-21.258	0.927	1.00	17.60	
2346	N	THR	299	-1.529	-20.026	2.424	1.00	17.60
2347	CA	THR	299	-2.622	-19.500	3.219	1.00	17.60
2348	CB	THR	299	-2.153	-19.417	4.567	1.00	17.60
2349	CG1	THR	299	-0.716	-19.680	4.730	1.00	17.60
2350	CG2	THR	299	-3.061	-20.284	5.495	1.00	17.60
2351	C	THR	299	-3.238	-18.062	2.701	1.00	17.60
2352	O	THR	299	-3.738	-17.286	3.304	1.00	17.60
2353	N	THR	300	-2.200	-17.633	1.433	1.00	17.60
2354	CA	THR	300	-2.331	-16.313	1.070	1.00	17.60
2355	CB	THR	300	-0.990	-15.959	0.461	1.00	17.60
2356	CG1	THR	300	-0.035	-16.067	1.459	1.00	17.60
2357	CG2	THR	300	-1.017	-14.548	-0.057	1.00	17.60
2358	C	THR	300	-3.459	-16.304	0.038	1.00	17.60
2359	O	THR	300	-3.424	-17.025	-0.923	1.00	17.60
2360	N	ASP	301	-4.539	-15.576	0.305	1.00	17.60
2361	CA	ASP	301	-5.601	-15.432	-0.643	1.00	17.60
2362	CB	ASP	301	-6.947	-15.959	0.055	1.00	17.60
2363	CG1	ASP	301	-8.111	-15.645	-0.895	1.00	17.60
2364	OD1	ASP	301	-8.274	-15.948	-0.385	1.00	17.60
2365	OD2	ASP	301	-8.048	-15.398	-2.116	1.00	17.60
2366	C	ASP	301	-5.370	-14.030	-1.192	1.00	17.60
2367	O	ASP	301	-5.717	-12.941	-0.684	1.00	17.60
2368	N	TRP	302	-4.712	-14.165	-2.331	1.00	17.60
2369	CA	TRP	302	-4.257	-13.050	-3.081	1.00	17.60
2370	CB	TRP	302	-3.619	-13.588	-4.389	1.00	17.60
2371	CG	TRP	302	-2.355	-14.329	-4.404	1.00	17.60
2372	CD1	TRP	302	-1.097	-13.821	-3.714	1.00	17.60
2373	CE2	TRP	302	-0.317	-14.976	-3.721	1.00	17.60
2374	CE3	TRP	302	-0.458	-12.619	-3.429	1.00	17.60
2375	CD1	TRP	302	-2.356	-15.478	-4.216	1.00	17.60
2376	N1	TRP	302	-1.099	-16.041	-4.013	1.00	17.60
2377	CE2	TRP	302	1.050	-14.961	-3.458	1.00	17.60
2378	CE3	TRP	302	0.900	-12.410	-3.170	1.00	17.60
2379	CH2	TRP	302	1.660	-13.749	-3.182	1.00	17.60
2380	C	TRP	302	-5.367	-12.097	-3.382	1.00	17.60
2381	O	TRP	302	-5.162	-10.909	-3.183	1.00	17.60
2382	N	ILE	303	-6.599	-12.471	-3.721	1.00	17.60
2383	CA	ILE	303	-7.576	-11.394	-3.977	1.00	17.60
2384	CB	ILE	303	-8.194	-11.859	-4.852	1.00	17.60
2385	CG1	ILE	303	-8.371	-12.642	-6.004	1.00	17.60
2386	CG2	ILE	303	-9.881	-12.679	-6.128	1.00	17.60
2387	C	ILE	303	-11.163	-13.150	-6.960	1.00	17.60
2388	O	ILE	303	-8.100	-10.771	-2.704	1.00	17.60
2389	N	ALA	304	-8.303	-9.585	-2.693	1.00	17.60
2390	CA	ALA	304	-8.316	-11.538	-1.620	1.00	17.60
2391	CB	ALA	304	-8.570	-11.002	-0.338	1.00	17.60
2392	CG	ALA	304	-8.314	-12.051	0.716	1.00	17.60
2393	C	ALA	304	-7.600	-9.809	-0.066	1.00	17.60
2394	O	ALA	304	-8.028	-8.655	0.063	1.00	17.60
2395	N	ILE	305	-6.298	-10.145	-0.231	1.00	17.60
2396	CA	ILE	305	-5.261	-9.130	-0.049	1.00	17.60
2397	CB	ILE	305	-3.915	-9.813	-0.249	1.00	17.60
2398	CG1	ILE	305	-2.818	-8.960	-0.712	1.00	17.60
2399	CG2	ILE	305	-3.624	-10.221	1.119	1.00	17.60
2400	CD1	ILE	305	-2.614	-11.325	1.068	1.00	17.60
2401	C	ILE	305	-5.462	-7.998	-0.968	1.00	17.60
2402	O	ILE	305	-5.606	-5.907	-0.489	1.00	17.60

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2468	CB	PRO	313	-3.645	-5.424	9.398	1.00	17.60	ATOM	2533	CA	PRO	321	-5.899	7.269	15.661	1.00	17.60
2469	CG	PRO	313	-5.116	-5.211	9.422	1.00	17.60	ATOM	2534	CB	PRO	321	-6.906	6.261	16.340	1.00	17.60
2470	CA	PRO	313	-3.234	-7.555	10.355	1.00	17.60	ATOM	2535	CC	PRO	321	-6.309	4.842	16.193	1.00	17.60
2471	C	PRO	313	-3.840	-7.521	10.424	1.00	17.60	ATOM	2536	C	PRO	321	-5.218	8.309	16.985	1.00	17.60
2472	N	PHE	314	-2.070	-8.111	10.111	1.00	17.60	ATOM	2537	N	GLY	322	-4.106	8.163	17.140	1.00	17.60
2473	CA	PHE	314	-1.459	-8.058	11.113	1.00	17.60	ATOM	2538	O	GLY	322	-5.837	9.490	16.675	1.00	17.60
2474	CB	PHE	314	-0.687	-9.945	10.418	1.00	17.60	ATOM	2539	CA	GLY	322	-5.707	10.583	17.415	1.00	17.60
2475	CG	PHE	314	-1.746	-10.940	9.923	1.00	17.60	ATOM	2540	C	GLY	322	-5.061	11.407	16.191	1.00	17.60
2476	CD1	PHE	314	-1.567	-11.481	8.349	1.00	17.60	ATOM	2541	O	GLY	322	-6.055	11.970	15.689	1.00	17.60
2477	CD2	PHE	314	-2.963	-11.205	10.525	1.00	17.60	ATOM	2542	N	ASP	323	-3.482	11.210	15.652	1.00	17.60
2478	CD3	PHE	314	-2.585	-12.243	7.995	1.00	17.60	ATOM	2543	CA	ASP	323	-3.418	11.016	14.316	1.00	17.60
2479	CD4	PHE	314	-3.992	-11.977	9.962	1.00	17.60	ATOM	2544	CB	ASP	323	-4.033	13.293	14.140	1.00	17.60
2480	CE	PHE	314	-3.790	-12.490	8.681	1.00	17.60	ATOM	2545	CC	ASP	323	-4.014	13.808	12.913	1.00	17.60
2481	C	PHE	314	-0.726	-7.741	11.792	1.00	17.60	ATOM	2546	CD1	ASP	323	-4.446	13.515	11.759	1.00	17.60
2482	O	PHE	314	0.290	-7.267	11.315	1.00	17.60	ATOM	2547	CD2	ASP	323	-5.753	14.592	13.159	1.00	17.60
2483	N	ILE	315	-1.476	-7.265	12.775	1.00	17.60	ATOM	2548	C	ASP	323	-3.936	11.099	14.412	1.00	17.60
2484	CA	ILE	315	-1.200	-6.160	13.664	1.00	17.60	ATOM	2549	O	ASP	323	-3.161	11.501	15.407	1.00	17.60
2485	CB	ILE	315	-0.477	-6.673	14.910	1.00	17.60	ATOM	2550	N	THR	324	-3.436	12.349	13.203	1.00	17.60
2486	CD2	ILE	315	-0.766	-5.688	16.108	1.00	17.60	ATOM	2551	CA	THR	324	-0.230	13.066	12.903	1.00	17.60
2487	CD3	ILE	315	-1.027	-6.000	15.341	1.00	17.60	ATOM	2552	CB	THR	324	-0.750	14.616	12.979	1.00	17.60
2488	CD1	ILE	315	-0.018	-9.021	14.952	1.00	17.60	ATOM	2553	CD1	THR	324	-1.975	14.756	12.200	1.00	17.60
2489	C	ILE	315	-0.439	-5.104	12.934	1.00	17.60	ATOM	2554	CD2	THR	324	0.221	15.626	12.350	1.00	17.60
2490	O	ILE	315	-1.100	-4.628	12.030	1.00	17.60	ATOM	2555	C	THR	324	1.080	12.836	13.655	1.00	17.60
2491	N	PRO	316	0.759	-4.541	13.195	1.00	17.60	ATOM	2556	O	THR	324	1.114	12.806	14.892	1.00	17.60
2492	CD	PRO	316	1.780	-4.283	12.185	1.00	17.60	ATOM	2557	N	SER	325	2.202	12.046	12.892	1.00	17.60
2493	CA	PRO	316	0.953	-3.524	14.218	1.00	17.60	ATOM	2558	CA	SER	325	3.552	12.666	13.424	1.00	17.60
2494	CB	PRO	316	2.463	-3.332	14.272	1.00	17.60	ATOM	2559	CB	SER	325	4.023	12.666	13.424	1.00	17.60
2495	CG	PRO	316	2.016	-3.332	12.832	1.00	17.60	ATOM	2560	CC	SER	325	4.010	15.039	13.469	1.00	17.60
2496	C	PRO	316	0.161	-2.328	13.692	1.00	17.60	ATOM	2561	C	SER	325	3.532	13.445	14.328	1.00	17.60
2497	O	PRO	316	0.786	-1.306	13.318	1.00	17.60	ATOM	2562	O	SER	325	2.794	10.492	13.982	1.00	17.60
2498	N	LYS	317	-1.195	-2.497	13.714	1.00	17.60	ATOM	2563	N	ASP	326	4.402	11.359	15.357	1.00	17.60
2499	CA	LYS	317	-2.159	-2.611	13.141	1.00	17.60	ATOM	2564	CA	ASP	326	3.378	10.477	17.371	1.00	17.60
2500	CB	LYS	317	-3.589	-2.209	13.411	1.00	17.60	ATOM	2565	CB	ASP	326	1.979	9.906	17.233	1.00	17.60
2501	CG	LYS	317	-4.309	-2.788	12.194	1.00	17.60	ATOM	2566	CC	ASP	326	3.119	10.266	16.450	1.00	17.60
2502	CD	LYS	317	-5.497	-2.739	12.510	1.00	17.60	ATOM	2567	CD1	ASP	326	1.661	9.003	16.131	1.00	17.60
2503	CD2	LYS	317	-5.004	-5.115	13.110	1.00	17.60	ATOM	2568	CD2	ASP	326	4.549	8.826	15.874	1.00	17.60
2504	CD3	LYS	317	-5.943	-6.253	13.330	1.00	17.60	ATOM	2569	C	ASP	326	6.556	7.889	16.631	1.00	17.60
2505	C	LYS	317	-2.063	-0.255	13.757	1.00	17.60	ATOM	2570	O	ASP	326	6.910	7.300	13.910	1.00	17.60
2506	O	LYS	317	-3.024	0.213	14.305	1.00	17.60	ATOM	2571	N	PHE	327	4.561	8.573	14.514	1.00	17.60
2507	N	PHE	318	-0.938	0.394	13.563	1.00	17.60	ATOM	2572	CA	PHE	327	3.775	6.794	13.133	1.00	17.60
2508	CA	PHE	318	-0.480	1.576	14.241	1.00	17.60	ATOM	2573	CB	PHE	327	2.819	6.144	14.059	1.00	17.60
2509	CB	PHE	318	0.222	2.334	13.098	1.00	17.60	ATOM	2574	CC	PHE	327	3.185	5.032	14.738	1.00	17.60
2510	CG	PHE	318	-0.833	3.104	12.472	1.00	17.60	ATOM	2575	CD1	PHE	327	1.565	6.437	14.184	1.00	17.60
2511	CD1	PHE	318	-1.983	2.639	11.940	1.00	17.60	ATOM	2576	CD2	PHE	327	2.273	4.396	15.546	1.00	17.60
2512	CD2	PHE	318	-0.790	4.514	12.652	1.00	17.60	ATOM	2577	CD3	PHE	327	0.677	5.989	15.002	1.00	17.60
2513	CD3	PHE	318	-3.060	3.470	11.650	1.00	17.60	ATOM	2578	CD4	PHE	327	1.014	4.873	15.678	1.00	17.60
2514	CE2	PHE	318	-1.874	5.372	12.349	1.00	17.60	ATOM	2579	C	PHE	327	6.088	7.523	12.954	1.00	17.60
2515	CE	PHE	318	-3.032	4.838	11.849	1.00	17.60	ATOM	2580	O	PHE	327	6.726	6.606	12.415	1.00	17.60
2516	C	PHE	318	-3.529	2.398	15.079	1.00	17.60	ATOM	2581	N	ASP	328	6.236	6.003	12.697	1.00	17.60
2517	O	PHE	318	-2.671	2.735	14.691	1.00	17.60	ATOM	2582	CA	ASP	328	7.306	9.287	11.796	1.00	17.60
2518	N	LYS	319	-1.110	2.836	16.269	1.00	17.60	ATOM	2583	CB	ASP	328	6.631	10.208	10.723	1.00	17.60
2519	CA	LYS	319	-1.990	3.543	17.380	1.00	17.60	ATOM	2584	CC	ASP	328	5.795	9.527	9.586	1.00	17.60
2520	CB	LYS	319	-1.265	3.655	18.504	1.00	17.60	ATOM	2585	CD	ASP	328	6.358	9.002	8.611	1.00	17.60
2521	CC	LYS	319	-0.071	4.698	18.531	1.00	17.60	ATOM	2586	CD1	ASP	328	6.550	9.366	9.629	1.00	17.60
2522	CD	LYS	319	1.096	4.268	19.454	1.00	17.60	ATOM	2587	CD2	ASP	328	8.183	9.955	12.915	1.00	17.60
2523	CE	LYS	319	1.457	2.791	19.216	1.00	17.60	ATOM	2588	O	ASP	328	7.767	10.083	14.110	1.00	17.60
2524	NE	LYS	319	1.850	2.572	17.830	1.00	17.60	ATOM	2589	N	ASP	328	9.418	10.356	12.526	1.00	17.60
2525	C	LYS	319	-2.514	4.892	16.750	1.00	17.60	ATOM	2590	CA	ASP	329	10.444	10.915	13.401	1.00	17.60
2526	O	LYS	319	-3.365	5.439	17.605	1.00	17.60	ATOM	2591	CB	ASP	329	11.750	10.306	12.999	1.00	17.60
2527	N	GLY	320	-2.104	5.417	15.629	1.00	17.60	ATOM	2592	CC	ASP	329	12.133	8.994	13.649	1.00	17.60
2528	CA	GLY	320	-2.475	6.713	15.254	1.00	17.60	ATOM	2593	CD	ASP	329	11.573	8.707	14.724	1.00	17.60
2529	C	GLY	320	-3.718	6.923	14.673	1.00	17.60	ATOM	2594	CD1	ASP	329	13.006	8.100	15.025	1.00	17.60
2530	CD	GLY	320	-3.818	7.575	15.412	1.00	17.60	ATOM	2595	CD2	ASP	329	10.663	12.427	13.528	1.00	17.60
2531	N	PRO	321	-4.083	6.396	15.031	1.00	17.60	ATOM	2596	C	ASP	329	9.893	13.259	13.039	1.00	17.60
2532	CD	PRO	321	-5.317	4.919	16.979	1.00	17.60	ATOM	2597	O	ASP	329					

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[illegible]

2728	CR	LVS	345	16.550	24.757	-20.001	1.00	17.60	ATOM	2722	CB	THR	352	23.791	-12.917	13.103	3.00	17.60
2729	CD	LVS	345	16.439	26.149	-20.707	1.00	17.60	ATOM	2723	CO1	THR	352	24.747	-14.257	13.192	1.00	17.60
2730	CD	LVS	345	15.141	26.300	-21.603	1.00	17.60	ATOM	2724	CO2	THR	352	24.243	-12.365	11.787	1.00	17.60
2731	CE	LVS	345	14.781	27.680	-22.294	1.00	17.60	ATOM	2725	C	THR	352	23.664	-10.863	14.572	1.00	17.60
2732	N2	LVS	345	15.570	28.070	-23.478	1.00	17.60	ATOM	2726	O	THR	352	24.125	-9.768	14.206	1.00	17.60
2733	C	LVS	345	15.444	22.975	-19.680	1.00	17.60	ATOM	2727	N	THR	353	22.575	-10.972	15.330	1.00	17.60
2734	O	LVS	345	14.965	22.114	-19.396	1.00	17.60	ATOM	2728	CB	THR	353	21.774	-9.430	15.710	1.00	17.60
2735	H	GLU	346	15.951	22.728	-17.494	1.00	17.60	ATOM	2729	CB	THR	353	20.659	-10.292	16.619	1.00	17.60
2736	CA	GLU	346	16.142	21.386	-16.968	1.00	17.60	ATOM	2730	CO1	THR	353	19.488	-8.180	17.295	1.00	17.60
2737	CB	GLU	346	17.187	21.506	-15.808	1.00	17.60	ATOM	2731	CO2	THR	353	18.415	-7.793	17.099	1.00	17.60
2738	CG	GLU	346	18.431	22.078	-16.553	1.00	17.60	ATOM	2732	CE1	THR	353	18.410	-9.630	15.713	1.00	17.60
2739	CD	GLU	346	20.007	23.495	-16.229	1.00	17.60	ATOM	2733	CE2	THR	353	17.381	-8.779	15.525	1.00	17.60
2740	OE1	GLU	346	18.223	24.482	-16.529	1.00	17.60	ATOM	2734	CE3	THR	353	17.400	-7.509	16.213	1.00	17.60
2741	OE2	GLU	346	14.866	20.498	-16.446	1.00	17.60	ATOM	2735	CH	THR	353	16.387	-6.614	15.997	1.00	17.60
2742	O	GLU	346	14.562	19.536	-16.000	1.00	17.60	ATOM	2736	C	THR	353	22.722	-8.902	16.436	1.00	17.60
2743	H	PHE	347	14.120	21.484	-15.632	1.00	17.60	ATOM	2737	D	THR	353	22.879	-7.788	15.968	1.00	17.60
2744	CA	PHE	347	12.931	21.043	-16.949	1.00	17.60	ATOM	2738	N	ALA	354	24.301	-8.692	18.240	1.00	17.60
2745	CB	PHE	347	13.069	21.421	-13.529	1.00	17.60	ATOM	2739	CB	ALA	354	25.168	-9.399	19.238	1.00	17.60
2746	CD	PHE	347	16.260	20.778	-12.950	1.00	17.60	ATOM	2740	C	ALA	354	25.436	-8.010	17.325	1.00	17.60
2747	CE1	PHE	347	15.403	21.513	-12.844	1.00	17.60	ATOM	2741	O	ALA	354	25.914	-6.910	17.654	1.00	17.60
2748	CE2	PHE	347	16.544	20.913	-12.371	1.00	17.60	ATOM	2742	N	ASP	355	25.786	-8.462	16.195	1.00	17.60
2749	CE3	PHE	347	15.355	18.893	-12.318	1.00	17.60	ATOM	2743	CA	ASP	355	26.756	-8.021	15.301	1.00	17.60
2750	C	PHE	347	16.321	19.606	-12.012	1.00	17.60	ATOM	2744	CB	ASP	355	27.351	-9.029	14.329	1.00	17.60
2751	H	THR	348	11.677	21.654	-13.524	1.00	17.60	ATOM	2745	CO1	ASP	355	28.176	-10.069	15.078	1.00	17.60
2752	C	THR	348	10.775	22.080	-14.811	1.00	17.60	ATOM	2746	CO2	ASP	355	29.318	-9.813	15.673	1.00	17.60
2753	N	THR	348	11.633	21.689	-16.849	1.00	17.60	ATOM	2747	C	ASP	355	26.156	-6.866	14.515	1.00	17.60
2754	CA	THR	348	10.520	22.291	-17.545	1.00	17.60	ATOM	2748	H	PHE	356	26.765	-5.790	14.356	1.00	17.60
2755	CB	THR	348	10.922	22.743	-18.277	1.00	17.60	ATOM	2749	CA	PHE	356	24.928	-7.038	14.072	1.00	17.60
2756	CG	THR	348	11.925	21.889	-19.494	1.00	17.60	ATOM	2750	CG	PHE	356	27.947	-6.491	12.844	1.00	17.60
2757	C	THR	348	11.285	21.458	-17.680	1.00	17.60	ATOM	2751	CG	PHE	356	27.159	-5.551	11.964	1.00	17.60
2758	H	GLU	349	8.185	23.040	-17.741	1.00	17.60	ATOM	2752	CD1	PHE	356	27.118	-5.033	10.815	1.00	17.60
2759	CA	GLU	349	9.349	20.342	-17.702	1.00	17.60	ATOM	2753	CD2	PHE	356	20.879	-1.216	12.306	1.00	17.60
2760	CB	GLU	349	8.747	18.203	-18.829	1.00	17.60	ATOM	2754	CE1	PHE	356	22.014	-4.200	9.981	1.00	17.60
2761	CG	GLU	349	7.851	17.036	-19.108	1.00	17.60	ATOM	2755	CE2	PHE	356	20.182	-4.415	11.472	1.00	17.60
2762	C	GLU	349	8.581	15.690	-19.016	1.00	17.60	ATOM	2756	CE3	PHE	356	20.740	-2.890	10.318	1.00	17.60
2763	CA	GLU	349	9.316	15.355	-19.964	1.00	17.60	ATOM	2757	C	PHE	356	24.078	-4.758	14.274	1.00	17.60
2764	CB	GLU	349	8.412	14.981	-17.997	1.00	17.60	ATOM	2758	O	PHE	356	24.431	-3.616	13.822	1.00	17.60
2765	CG	GLU	349	7.629	18.855	-16.604	1.00	17.60	ATOM	2759	N	ILE	357	23.853	-4.863	15.533	1.00	17.60
2766	C	GLU	349	7.462	17.668	-16.330	1.00	17.60	ATOM	2760	CA	ILE	357	23.277	-3.693	16.366	1.00	17.60
2767	CA	PHE	350	6.982	19.393	-14.376	1.00	17.60	ATOM	2761	CB	ILE	357	22.452	-4.012	17.593	1.00	17.60
2768	CB	PHE	350	8.291	19.497	-13.503	1.00	17.60	ATOM	2762	CG	ILE	357	21.154	-4.485	17.047	1.00	17.60
2769	CG	PHE	350	8.275	18.888	-12.078	1.00	17.60	ATOM	2763	CO1	ILE	357	22.954	-3.265	18.382	1.00	17.60
2770	C	PHE	350	7.306	19.222	-11.144	1.00	17.60	ATOM	2764	CD1	ILE	357	24.120	-5.106	19.429	1.00	17.60
2771	CA	PHE	350	9.206	17.330	-11.758	1.00	17.60	ATOM	2765	CD2	ILE	357	24.362	-2.800	16.852	1.00	17.60
2772	CB	PHE	350	7.253	18.606	-9.914	1.00	17.60	ATOM	2766	O	ILE	357	24.145	-1.714	17.374	1.00	17.60
2773	CG	PHE	350	9.149	17.315	-10.527	1.00	17.60	ATOM	2767	N	ALA	358	25.527	-3.387	16.658	1.00	17.60
2774	C	PHE	350	8.379	17.644	-9.609	1.00	17.60	ATOM	2768	CA	ALA	358	26.812	-2.773	16.874	1.00	17.60
2775	CA	PHE	350	5.877	20.337	-13.924	1.00	17.60	ATOM	2769	CB	ALA	358	27.706	-3.880	17.370	1.00	17.60
2776	CB	PHE	350	5.936	21.507	-14.301	1.00	17.60	ATOM	2770	C	ALA	358	27.370	-2.116	15.569	1.00	17.60
2777	CG	PHE	350	5.019	19.339	-13.118	1.00	17.60	ATOM	2771	O	ALA	358	28.382	-1.398	13.623	1.00	17.60
2778	C	THR	351	26.441	-12.459	17.857	1.00	17.60	ATOM	2772	N	SPR	359	26.787	-2.218	14.366	1.00	17.60
2779	CA	THR	351	25.021	-13.778	17.829	1.00	17.60	ATOM	2773	CA	SPR	359	27.387	-1.679	13.177	1.00	17.60
2780	CB	THR	351	24.486	-13.256	19.195	1.00	17.60	ATOM	2774	CB	SPR	359	27.198	-2.614	12.067	1.00	17.60
2781	CG	THR	351	23.449	-12.380	18.834	1.00	17.60	ATOM	2775	OC	SPR	359	25.855	-2.919	11.925	1.00	17.60
2782	C	THR	351	25.504	-12.532	20.177	1.00	17.60	ATOM	2776	C	SPR	359	26.536	-0.461	12.895	1.00	17.60
2783	CA	THR	351	24.785	-12.806	16.555	1.00	17.60	ATOM	2777	O	SPR	359	25.464	-0.262	13.462	1.00	17.60
2784	CB	THR	351	25.320	-11.841	16.867	1.00	17.60	ATOM	2778	N	GLY	360	27.052	0.286	11.918	1.00	17.60
2785	CG	THR	351	24.295	-13.134	15.672	1.00	17.60	ATOM	2779	CA	GLY	360	26.463	1.519	11.458	1.00	17.60
2786	C	THR	351	24.364	-12.206	14.339	1.00	17.60	ATOM	2780	C	GLY	360	25.270	1.300	10.553	1.00	17.60
2787	CA	THR	351						ATOM	2781	O	GLY	360	24.670	2.221	10.439	1.00	17.60
2788	CB	THR	351						ATOM	2782	N	ARG	361	24.983	0.151	9.936	1.00	17.60

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2857 CA ARG 361 23.826 0.011 9.036 1.00 17.60
 2858 CB ARG 361 23.953 0.139 9.139 1.00 17.60
 2859 CD ARG 361 25.149 -1.350 9.129 1.00 17.60
 2860 CE ARG 361 25.502 -0.180 9.169 1.00 17.60
 2861 CF ARG 361 24.559 0.737 9.704 1.00 17.60
 2862 CG ARG 361 24.592 1.425 9.612 1.00 17.60
 2863 CH ARG 361 25.602 2.359 9.899 1.00 17.60
 2864 CI ARG 361 23.589 1.139 9.794 1.00 17.60
 2865 CJ ARG 361 22.467 -0.095 9.718 1.00 17.60
 2866 CK ARG 361 21.518 -0.584 9.092 1.00 17.60
 2867 CL ARG 361 22.365 0.282 11.002 1.00 17.60
 2868 CM ARG 361 21.116 0.199 11.746 1.00 17.60
 2869 CN ARG 361 23.614 0.053 13.275 1.00 17.60
 2870 CO ARG 361 22.348 1.035 13.731 1.00 17.60
 2871 CP ARG 361 21.947 -1.349 13.524 1.00 17.60
 2872 CQ ARG 361 20.275 1.427 13.466 1.00 17.60
 2873 CR ARG 361 19.060 1.468 13.557 1.00 17.60
 2874 CS ARG 361 21.009 2.451 13.006 1.00 17.60
 2875 CT ARG 361 20.449 3.747 10.637 1.00 17.60
 2876 CU ARG 361 19.450 3.604 9.510 1.00 17.60
 2877 CV ARG 361 19.183 2.508 9.028 1.00 17.60
 2878 CW ARG 361 18.004 1.685 9.073 1.00 17.60
 2879 CX ARG 361 17.987 1.653 7.992 1.00 17.60
 2880 CY ARG 361 17.061 5.018 7.857 1.00 17.60
 2881 CZ ARG 361 15.958 5.373 8.638 1.00 17.60
 2882 DA ARG 361 14.635 6.271 8.675 1.00 17.60
 2883 DB ARG 361 14.157 6.592 7.317 1.00 17.60
 2884 DC ARG 361 16.099 7.070 6.919 1.00 17.60
 2885 DD ARG 361 14.464 8.040 7.011 1.00 17.60
 2886 DE ARG 361 13.792 8.187 5.694 1.00 17.60
 2887 DF ARG 361 19.075 4.752 6.855 1.00 17.60
 2888 DG ARG 361 19.910 5.658 6.856 1.00 17.60
 2889 DH ARG 361 19.152 3.717 6.032 1.00 17.60
 2890 DI ARG 361 20.194 3.586 5.035 1.00 17.60
 2891 DJ ARG 361 20.235 3.128 6.717 1.00 17.60
 2892 DK ARG 361 20.533 3.412 5.999 1.00 17.60
 2893 DL ARG 361 20.355 -0.009 5.766 1.00 17.60
 2894 DM ARG 361 19.066 -0.376 5.269 1.00 17.60
 2895 DN ARG 361 18.127 -0.945 5.911 1.00 17.60
 2896 DO ARG 361 16.974 -1.283 5.400 1.00 17.60
 2897 DP ARG 361 16.275 -2.111 7.210 1.00 17.60
 2898 DQ ARG 361 19.889 4.509 3.807 1.00 17.60
 2899 DR ARG 361 18.788 4.551 3.299 1.00 17.60
 2900 DS ARG 361 20.847 5.361 3.519 1.00 17.60
 2901 DT ARG 361 20.821 6.231 3.453 1.00 17.60
 2902 DU ARG 361 21.567 7.384 2.628 1.00 17.60
 2903 DV ARG 361 20.706 6.492 2.470 1.00 17.60
 2904 DW ARG 361 20.544 9.037 1.343 1.00 17.60
 2905 DX ARG 361 20.762 5.546 1.139 1.00 17.60
 2906 DY ARG 361 21.353 4.461 1.009 1.00 17.60
 2907 DZ ARG 361 20.271 6.203 0.040 1.00 17.60
 2908 EA ARG 361 20.282 5.676 -1.296 1.00 17.60
 2909 EB ARG 361 19.127 6.173 -2.110 1.00 17.60
 2910 EC ARG 361 21.517 6.095 -1.036 1.00 17.60
 2911 ED ARG 361 21.859 7.269 -1.958 1.00 17.60
 2912 EE ARG 361 22.153 5.154 -2.719 1.00 17.60
 2913 EF ARG 361 23.319 5.440 -3.450 1.00 17.60
 2914 EG ARG 361 24.241 4.218 -3.703 1.00 17.60
 2915 EH ARG 361 23.032 4.315 -2.455 1.00 17.60
 2916 EI ARG 361 23.629 2.938 -4.076 1.00 17.60
 2917 EJ ARG 361 21.627 2.182 -4.878 1.00 17.60
 2918 EK ARG 361 23.201 6.060 -1.806 1.00 17.60
 2919 EL ARG 361 22.153 6.507 -5.218 1.00 17.60
 2920 EN ARG 361 24.374 6.102 -5.593 1.00 17.60

2921 CA ARG 361 24.369 6.697 -6.795 1.00 17.60
 2922 CB ARG 361 23.363 6.032 -7.248 1.00 17.60
 2923 CD ARG 361 23.112 6.072 -8.024 1.00 17.60
 2924 CE ARG 361 21.953 7.571 -9.335 1.00 17.60
 2925 CF ARG 361 26.063 7.167 -9.937 1.00 17.60
 2926 CG ARG 361 23.459 7.987 -10.783 1.00 17.60
 2927 CH ARG 361 22.193 8.212 -10.834 1.00 17.60
 2928 CI ARG 361 26.027 8.162 -6.029 1.00 17.60
 2929 CJ ARG 361 22.910 8.697 -6.651 1.00 17.60
 2930 CK ARG 361 25.077 8.671 -7.473 1.00 17.60
 2931 CL ARG 361 25.319 9.962 -8.033 1.00 17.60
 2932 CM ARG 361 25.602 11.006 -8.846 1.00 17.60
 2933 CN ARG 361 25.040 10.621 -5.423 1.00 17.60
 2934 CO ARG 361 23.831 10.619 -5.222 1.00 17.60
 2935 CP ARG 361 25.719 10.239 -4.512 1.00 17.60
 2936 CQ ARG 361 26.598 9.431 -8.865 1.00 17.60
 2937 CR ARG 361 27.783 9.719 -8.502 1.00 17.60
 2938 CS ARG 361 26.366 8.538 -9.720 1.00 17.60

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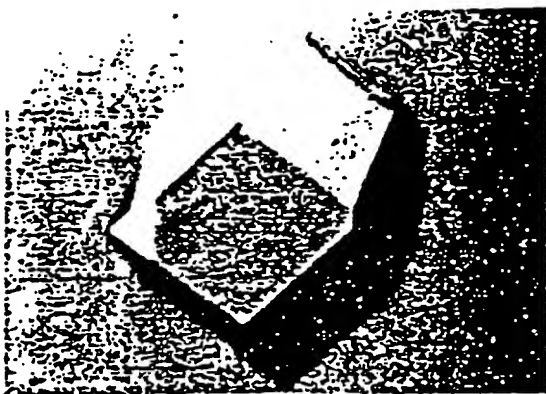


FIGURE 18A



FIGURE 18B

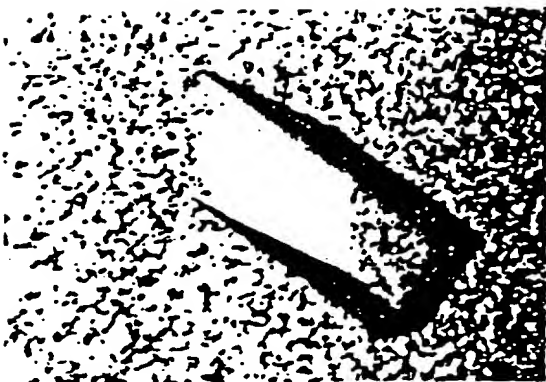


FIGURE 18C

TABLE 1 Structure Solution Statistics

a). Diffraction Data:							
Data Sets	No. of Crystals	d _{min} (Å)	No. of Measurements	No. of Reflections	<I/σ(I)>	Completeness (%)	R _{sym}
Native-1	2	2.7	58889	12713	12.9	98.1	0.061
Native-2	1	2.7	27067	11291	13.2	87.3	0.040
PIIMB-1	1	3.0	30973	7233	13.3	76.1	0.063
PIIMB-2	1	3.0	23476	8809	6.9	92.1	0.075
MgATP	1	2.7	26464	11840	11.1	91.1	0.048

(b) **SIRAS Phasing Statistics:**

	Overall	11.72	7.70	Average	Shell	Resolution (Å)	
Mean figure of merit	0.57	0.74	0.75	0.68	0.62	0.53	0.43
PIIMB-1							
acentric r.m.s. f_h/E_{iso}	2.73	3.27	3.80	3.70	3.00	2.54	2.26
r.m.s. $\Delta F_{anom}/E_{anom}$	0.96	1.53	1.68	1.41	1.19	1.05	0.78
R_c	0.50	0.34	0.38	0.45	0.51	0.78	0.70
PIIMB-2							
acentric r.m.s. f_h/E_{iso}	2.26	3.89	3.72	3.35	2.81	2.34	1.83
R_c	0.60	0.37	0.53	0.58	0.64	0.75	0.65

(c) Refinement:

Model	No. of Residues/Chains	Initial R-factor	Final R-factor	B	Data Selection
A. First unrefined partial	275/4	0.473	0.304	overall	10-2.7 Å, $F/\sigma > 2$
B. First unrefined full	356/2	0.434	0.228	overall	10-2.7 Å, $F/\sigma > 2$
C. Latest X-PLOR	356/2	—	0.195	individual	10-2.7 Å, $F/\sigma > 2$
D. TNT	356/2	0.221	0.212	individual	40-2.7 Å

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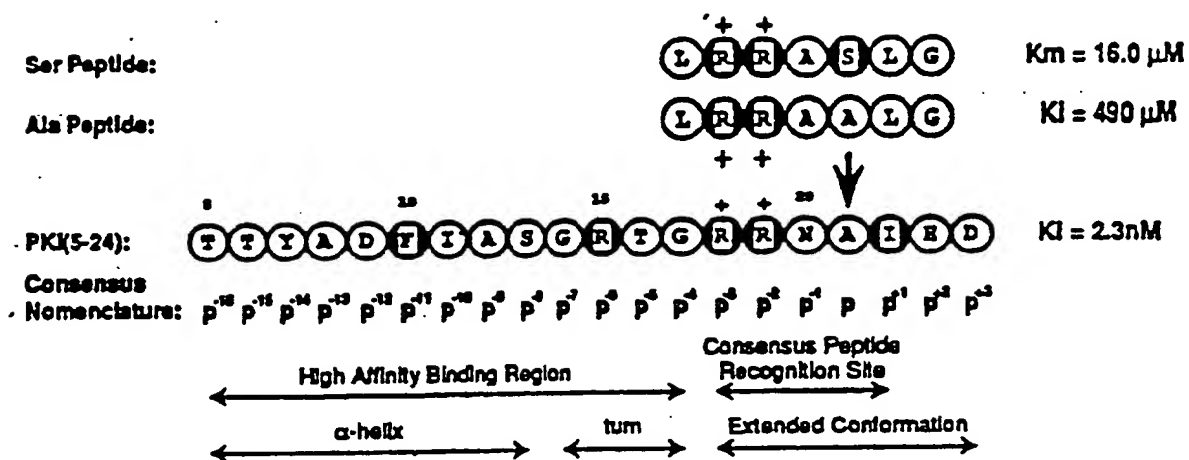


TABLE 2

TABLE 3

<u>POINTS OF CONTACT</u>	<u>POSITION</u>	<u>cAPK</u>	<u>CKII</u>
P+1	197	Thr	Val
	198	Leu	Arg
	199	Cys	Val
	200	Gly	Ala
	201	Thr	Ser
	202	Pro	Arg
	203	Glu	Tyr
	204	Tyr	Phe
	205	Leu	Lys
P-2	170	Glu	His
	230	Glu	Glu
P-3	127	Glu	Asp
	331	Glu	
P-6	203	Leu	
P-11	235	Tyr	
	236	Pro	
	237	Pro	
	238	Phe	
	239	Phe	

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TABLE 4

Angstroms apart	Atom 1	Atom 2
5.29	ASP 184 CA	GLY 186 CA
5.73	GLU 91 CA	GLY 186 CA
6.46	ASN 171 CA	ASP 184 CA
7.41	ASN 171 CA	ASP 166 CA
7.61	ASP 166 CA	GLY 186 CA
7.87	ASP 184 CA	GLU 91 CA
8.20	ASP 166 CA	ASP 184 CA
9.20	ASP 184 CA	LYS 72 CA
9.90	GLY 52 CA	LYS 72 CA
10.15	ASN 171 CA	GLY 186 CA
10.29	ASP 184 CA	GLY 52 CA
10.53	GLY 52 CA	GLY 186 CA
10.78	ASN 171 CA	GLY 52 CA
10.91	GLY 186 CA	LYS 72 CA
11.29	GLU 91 CA	LYS 72 CA
11.80	ARG 280 CA	GLU 208 CA
12.27	ASP 166 CA	GLU 91 CA
12.65	ASP 166 CA	GLY 52 CA
13.52	ASN 171 CA	LYS 72 CA
14.07	ASN 171 CA	GLU 91 CA
15.02	GLU 91 CA	GLY 52 CA
15.07	ASP 166 CA	GLU 208 CA
16.54	ASP 166 CA	LYS 72 CA
18.58	ARG 280 CA	ASP 166 CA
19.99	GLU 208 CA	GLY 186 CA
22.00	ASN 171 CA	GLU 208 CA
22.82	ASP 184 CA	GLU 208 CA
23.37	GLU 91 CA	GLU 208 CA
23.49	ARG 280 CA	ASN 171 CA
24.87	ARG 280 CA	GLY 186 CA
25.18	GLU 208 CA	GLY 52 CA
25.61	ARG 280 CA	ASP 184 CA
27.34	ARG 280 CA	GLU 91 CA
30.53	GLU 208 CA	LYS 72 CA
30.83	ARG 280 CA	GLY 52 CA
34.67	ARG 280 CA	LYS 72 CA

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 92/06137

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ⁶		
According to International Patent Classification (IPC) or to both National Classification and IPC		
Int.Cl. 5 C12Q1/00; C07K13/00;	C12Q1/48; A61K37/64;	C12N9/99; C07K15/00 G01N33/68
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁷		
Classification System	Classification Symbols	
Int.Cl. 5	C12Q ; C12N ; G01N ; A61K C07K	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁸		
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹		
Category ^o	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
A	WO,A,8 907 654 (PROGENICS PHARMACEUTICALS) 24 August 1989 see claims; example 1 ---	1
A	EP,A,0 359 981 (BOEHRINGER) 28 March 1990 see claims ---	1,29
	-/--	
^o Special categories of cited documents : ¹⁰ "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "A" document member of the same patent family		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search 04 NOVEMBER 1992		Date of Mailing of this International Search Report 16. 11. 92
International Searching Authority EUROPEAN PATENT OFFICE		Signature of Authorized Officer DELANGHE L.L.M.

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	<p>CHEMICAL ABSTRACTS, vol. 105, no. 21, 24 November 1986, Columbus, Ohio, US; abstract no. 186487d, CLORE, G.MARIUS ET AL. 'Stereochemistry of binding of the tetrapeptide acetyl-Pro-Ala-Pro-Tyr-NH₂ to porcine pancreatic elastase. Combined use of two-dimensional transferred nuclear Overhauser enhancement measurements, restrained molecular dynamics, X-ray crystallography and molecular modelling.' page 314 ; see abstract & J.MOL.BIOL. vol. 190, no. 2, 1986, ENG pages 259 - 267</p>	1
P,X	<p>--- CHEMICAL ABSTRACTS, vol. 115, no. 13, 30 September 1991, Columbus, Ohio, US; abstract no. 130637s, KNIGHTON, DANIEL R. ET AL. 'Crystallization of cAMP-dependent protein kinase. Cocystals of the catalytic subunit with a 20 amino acid residue peptide inhibitor and magnesium-ATP diffract to 3.0 Å resolution.' page 453 ; see abstract & J.MOL.BIOL. vol. 220, no. 2, 1991, ENG pages 217 - 220</p>	1
P,X	<p>--- SCIENCE vol. 253, no. 5018, 26 July 1991, LANCASTER, PA US pages 414 - 420 D.R.KNIGHTON ET AL. 'Structure of a peptide inhibitor bound to the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase.' see page 420 see the whole document</p> <p>-----</p>	1-66

**ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO.**

US 9206137
SA 62983

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 04/11/92

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		JP-T-	3503598	15-08-91

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